

OLSZÓWKA, Magdalena, TOCZEK, Wiktoria Oliwia, BOGDAN, Klaudia, JANKOWSKI, Mikołaj, JANICKA, Urszula, CIEPLUCH, Natalia and SŁOMIŃSKI, Szymon Stanisław. The Gut Microbiome in Athletes: Mechanisms, Training Adaptations, and Implications for Performance — A Comprehensive Review. *Quality in Sport*. 2025;48:67288. eISSN 2450-3118.

<https://doi.org/10.12775/QS.2025.48.67288>

<https://apcz.umk.pl/QS/article/view/67288>

The journal has been awarded 20 points in the parametric evaluation by the Ministry of Higher Education and Science of Poland. This is according to the Annex to the announcement of the Minister of Higher Education and Science dated 05.01.2024, No. 32553. The journal has a Unique Identifier: 201398. Scientific disciplines assigned: Economics and Finance (Field of Social Sciences); Management and Quality Sciences (Field of Social Sciences).

Punkty Ministerialne z 2019 - aktualny rok 20 punktów. Załącznik do komunikatu Ministra Szkolnictwa Wyższego i Nauki z dnia 05.01.2024 Lp. 32553. Posiada Unikatowy Identyfikator Czasopisma: 201398. Przypisane dyscypliny naukowe: Ekonomia i finanse (Dziedzina nauk społecznych); Nauki o zarządzaniu i jakości (Dziedzina nauk społecznych). © The Authors 2025.

This article is published with open access under the License Open Journal Systems of Nicolaus Copernicus University in Torun, Poland. Open Access: This article is distributed under the terms of the Creative Commons Attribution Noncommercial License, which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited. This is an open access article licensed under the terms of the Creative Commons Attribution Non-commercial Share Alike License (<http://creativecommons.org/licenses/by-nc-sa/4.0/>), which permits unrestricted, non-commercial use, distribution, and reproduction in any medium, provided the work is properly cited.

The authors declare that there is no conflict of interest regarding the publication of this paper.

Received: 09.12.2025. Revised: 25.12.2025. Accepted: 25.12.2025. Published: 29.12.2025.

## THE GUT MICROBIOME IN ATHLETES: MECHANISMS, TRAINING ADAPTATIONS, AND IMPLICATIONS FOR PERFORMANCE — A COMPREHENSIVE REVIEW

Magdalena Olszówka<sup>1</sup>, ORCID <https://orcid.org/0009-0007-5196-3906>

E-mail: [magdalenaolszowka2@gmail.com](mailto:magdalenaolszowka2@gmail.com)

<sup>1</sup>Stefan Cardinal Wyszyński Provincial Specialist Hospital SPZOZ in Lublin, Lublin, Poland

Wiktoria Oliwia Toczek<sup>2</sup>, ORCID <https://orcid.org/0009-0009-3530-6660>

E-mail: [toczek.wiktoria2@gmail.com](mailto:toczek.wiktoria2@gmail.com)

<sup>2</sup>Ludwik Rydygier Specialist Hospital in Cracow, os. Złotej Jesieni 1, 31-826 Kraków, Poland

Klaudia Bogdan<sup>2</sup>, ORCID <https://orcid.org/0009-0003-7260-2799>

E-mail: [klaudiabogdan27@gmail.com](mailto:klaudiabogdan27@gmail.com)

<sup>2</sup>Ludwik Rydygier Specialist Hospital in Cracow, os. Złotej Jesieni 1, 31-826 Kraków, Poland

Mikołaj Jankowski<sup>2</sup>, ORCID <https://orcid.org/0009-0009-6542-9143>

E-mail: [mr.mikolajjankowski@gmail.com](mailto:mr.mikolajjankowski@gmail.com)

<sup>2</sup>Ludwik Rydygier Specialist Hospital in Cracow, os. Złotej Jesieni 1, 31-826 Kraków, Poland

Urszula Janicka<sup>3</sup>, ORCID <https://orcid.org/0009-0001-7324-2137>

E-mail: [ujanicka.uj@gmail.com](mailto:ujanicka.uj@gmail.com)

<sup>3</sup>Lower Silesian Center of Oncology, Pulmonology and Hematology, Plac Ludwika Hirszfelda 12, 53-413 Wrocław, Poland

Natalia Ciepluch<sup>4</sup>, ORCID <https://orcid.org/0009-0005-1703-4674>

E-mail: [nw.ciepluch@gmail.com](mailto:nw.ciepluch@gmail.com)

<sup>4</sup>Municipal Hospital No. 4 in Gliwice, Zygmunta Starego 20, 44-100 Gliwice, Poland

Szymon Stanisław Słomiński<sup>5</sup>, ORCID <https://orcid.org/0009-0006-0208-0608>

E-mail: [szymonslominski085@gmail.com](mailto:szymonslominski085@gmail.com)

<sup>5</sup>University Clinical Hospital in Poznań, Przybyszewskiego 49, 60-355 Poznań, Poland

### Corresponding Author

Magdalena Olszówka, E-mail: [magdalenaolszowka2@gmail.com](mailto:magdalenaolszowka2@gmail.com)

**Abstract****Background**

The gut microbiota is increasingly seen as a key regulator of functions relevant to athletic performance. Exercise alters microbial composition, while microbial metabolites influence metabolism, immunity, gut barrier integrity, and recovery. Despite rapid growth of research, findings are inconsistent, underscoring the need for clearer mechanisms. This review summarizes current evidence on the bidirectional relationship between training and the gut microbiota and its implications for performance.

**Aim**

To consolidate open-access research on links between gut microbiota, exercise physiology, and performance, focusing on metabolic pathways, gut–muscle communication, immune modulation, intestinal permeability, and training-specific effects.

**Material and Methods**

This narrative review draws on open-access literature up to 2025, identified through PubMed, PMC, and Google Scholar. Included studies comprise mechanistic animal work, observational data, randomized trials, and systematic reviews examining physical activity, microbiota composition, microbial metabolites, and performance in athletes.

**Results**

Endurance training induces microbial adaptations, including increases in taxa metabolizing lactate and generating propionate. Strength and HIIT training show more variable changes shaped by diet, training load, and metabolic health. Microbial metabolites—especially SCFAs—may enhance energy regulation, immune balance, and gut barrier function, though evidence remains fragmented and causality unclear.

**Conclusion**

The gut microbiota is a dynamic component of exercise adaptation. While diverse and SCFA-producing microbes appear beneficial for metabolism, inflammation control, and recovery, overall findings are heterogeneous. More interdisciplinary research is needed to clarify causal pathways and support personalized, microbiome-based strategies in sports nutrition and training.

**Keywords:**

gut microbiome; athletic performance; exercise physiology; short-chain fatty acids (SCFAs); gut–muscle axis; intestinal barrier; endurance training; HIIT; strength training; microbial diversity; sports nutrition

## 1. Introduction

The gut microbiome is a dynamic, complex ecosystem consisting of bacteria, fungi, viruses, and archaea, that reside primarily in the human digestive tract. The main phyla of bacteria include *Firmicutes*, *Bacteroidetes*, *Proteobacteria*, *Actinobacteria* and *Verrucomicrobia* [1]. They interact with each other and with the host, playing a pivotal role in digestion, vitamin synthesis, short-chain fatty acids (SCFAs) production, and affecting brain health, immune system and inflammatory response [2, 3].

The gut microbiome is primarily shaped by the mode of delivery at birth, and is typically affected by factors such as age, diet, exercise, sleep, stress, illness, and drugs use [4, 5, 6]. Dysbiosis, an imbalance of the gut microbiome composition, may impact the integrity of the intestinal barrier, and is a cause to diseases such as diabetes mellitus, obesity, arthritis, and cardiovascular and neurodegenerative diseases [2].

In recent years, research has increasingly highlighted the importance of the gut microbiome in the context of human physical activity. According to Mach and Fuster-Botella (2017), regular exercise may affect the composition of the gut microbiota, increasing its diversity, especially in the *Firmicutes* phylum [4]. This diversity is associated with nutrient absorption, and enhanced production of short-chain fatty acids, that can result in the improvement of the endurance performance of the host. Moderate exercise can reduce inflammation, by causing a rise in interleukin-6, whereas prolonged, high-intensity training is connected to temporary immunosuppression, oxidative stress, and increased risk of illnesses [7]. Furthermore, certain bacterial genus may metabolize exercise-related compounds, such as lactate, into substrates for subsequent energy production. A study by Scheiman et al. showed that *Veillonella* species use lactate, producing propionate, that can increase VO<sub>2</sub> max and overall performance [8].

Moreover, the microbiome is modified by factors fundamental for athletes: type of training and diet. For example, a high-protein diet typically associated with strength training can impact the diversity of the gut microbiota [9]. Endurance athletes often consume a high-carbohydrate diet, which can positively influence fermentation efficiency of the gut microbiome through the impact on an abundance of *Prevotella* [10]. Exercise itself modulates microbial composition by affecting gut transit time, immune signaling, and mucus secretion [11]. Accumulating studies also indicate that the increase of bacterial fermentation products can reduce recovery time of the athletes [10, 12].

Despite growing interest, no universal model fully explains how exercise, diet, and microbiota converge to shape performance outcomes. Available evidence is fragmented and varies across disciplines. Thus, the purpose of this review is to synthesize current open-access research and evaluate the connection between the gut microbiome and metabolism, type of training and performance. Specifically, the review focuses on:

- (1) biological foundations of microbiome function;
- (2) mechanistic links between microbiota and exercise physiology;
- (3) effects of different training modalities;
- (4) distinctive features of elite athletes;
- (5) dietary strategies modulating microbiome composition; and
- (6) emerging biomarkers and future directions.

By consolidating these findings, this work aims to support the development of personalized nutrition and training approaches informed by the gut microbiome.

## **2. Biological Foundations of the Gut Microbiome**

The gut microbiome is an integral component of human physiology, influencing metabolic, endocrine, and immune systems. One of its most important functions involves the fermentation of non-digestible carbohydrates from dietary fiber. The metabolites of that fermentation are short-chain fatty acids, such as acetate, propionate and butyrate. They are not only substrates for colonocytes, but also take part in maintaining the integrity of the gut barrier and regulate gut endocrine functions [13]. Studies show that SCFAs can also have an impact on satiety, by influencing the peptide YY and glucagon-like peptide-1 production [14]. Regular physical activity enhances microbial diversity, leading to the increase of the SCFAs production [15].

The gut microbiota also metabolises bile acids (BAs) produced in the liver. In the intestine, BAs regulate digestion and metabolism of lipids and glucose by activating nuclear receptors. They also impact the composition of the microbiota, through their antimicrobial features [16]. Moreover, secondary BAs present anti-inflammatory effect by mediating the reduction of inflammatory cytokine production [17].

### **2.1 Gut barrier and immune regulation**

The intestinal barrier is crucial for preventing translocation of harmful molecules, especially during strenuous physical activity, and SCFAs play a key role in maintaining a healthy gut barrier. For example, butyrate is the main energy source for the intestinal epithelial cells (IEC) and it reduces the permeability of the IEC. Furthermore, butyrate affects the immune system, by decreasing the levels of pro-inflammatory cytokines and increasing the synthesis of antimicrobial peptides [14]. SCFAs also modulate mucosal secretory functions, thereby protecting the mucosa [13]. BAs maintain the integrity of the gut barrier by influencing the expression of tight junction proteins [18].

A compromised barrier and increased intestinal permeability—often referred to as “leaky gut”—can increase local and systemic inflammation and impair recovery [19]. Ultimately, reduced SCFAs production may elevate the risk of the injuries among athletes [20].

### **2.2 Circadian rhythms and exercise**

Studies demonstrate that there is a bidirectional influence between the circadian clock and the gut microbiota. It is determined by the feeding time, dietary composition and activity cycles [21]. Disruptions, such as those caused by irregular sleeping patterns or travel (common in competitive athletes), may impact microbial diversity and lead to metabolic dysfunction [22, 23, 24]. Synchronization between training, nutrition, and feeding/fasting phases with circadian rhythms may preserve microbial homeostasis and therefore optimize physiological responses of athletes [25].

### **2.3 Gut-muscle axis**

Animal models research shows that the gut microbiota affects skeletal muscle metabolism and function [26]. Possible mechanisms include participating in inducing insulin growth factor 1, that promotes anabolism. SCFAs may stimulate mitochondrial energy production, contributing

to the muscle gain [27]. Moreover, it has been shown that the dysbiosis leads to increased expression of the markers associated with muscle atrophy [28]. BAs may also influence muscle metabolism through inducing fibroblast growth factor 19 in the intestine [28].

### **3. Mechanisms Linking the Gut Microbiome With Exercise Physiology**

#### **3.1 Energy metabolism**

Regular physical activity is linked to the abundance of the bacteria of the phylum *Firmicutes*, such as *Ruminococcus*, *Lachnospiraceae* and *Holdemanella*. *Ruminococcus* produces the SCFA succinate, which is involved in intestinal gluconeogenesis and thereby impacts glucose homeostasis [29]. It is also connected to increased insulin sensitivity. *Lachnospiraceae* produces acetate, amongst others, which is shown to improve glucose metabolism. *Holdemanella* may reduce hyperglycemia by regulating the production of the GLP-1 hormone [30]. SCFAs also elevate the production of peptide YY [31], that modulates intestinal transit and impacts satiety, ultimately decreasing food intake. Intense aerobic exercises are also associated with an increase in the abundance of *Bacteroidetes*. *Bacteroidetes* participate in the metabolism of carbohydrates, utilizing them through Carbohydrate-Active Enzymes (CAZymes) [32].

SCFAs, mainly propionate and acetate, activate free fatty acid receptors, FFAR2 and FFAR3, that regulate lipid storage and maintain metabolic homeostasis. The activation of receptors promotes the usage of excess energy, instead of storing it. It has also been shown to reduce the insulin sensitivity of adipocytes, ultimately leading to elevated energy expenditure [33, 34, 35]. Studies also show the increase of *Veillonella* in the marathon runners' gut [8]. This species can utilize lactate into acetate and propionate, which are the energy sources, thereby improving the endurance of the host [36].

#### **3.2 Immune modulation**

High training loads, particularly in endurance athletes, are associated with transient immunosuppression [37]. Intense exercise may increase the risk of infection through many pathways. For example, it influences production of the cytokines, that regulate the immune system [38]. Excessive training load has been associated with a higher risk of upper respiratory tract infections [39]. However, higher microbiome diversity contributes to the modulation of the immune system of the host and decrease of the upper respiratory tract infection risk [40]. Furthermore, probiotic supplementation has demonstrated benefits in modulating immune responses and reducing infection incidence [41, 42]. The gut microbiota also influences the immune system by stimulating the IgA secretion and shaping the gut associated lymphoid tissues (GALT) [43].

#### **3.3 Regulation of inflammation and oxidative stress**

Strenuous physical activity induces the production of reactive oxygen species (ROS), that leads to oxidative damage and inflammation, and can subsequently reduce athletic performance. It also impacts the release of glucocorticoids, that decrease the production of anti-inflammatory cytokines [44, 45]. It has been shown that probiotic interventions in athletes can reduce oxidative stress levels and improve inflammatory responses [46, 47]. Certain *Lactobacillus*

species may decrease cytokines production and mitigate systemic inflammation [47]. SCFAs, mainly butyrate and propionate, inhibit the ROS production and expression of the pro-inflammatory cytokines [48, 49].

### **3.4 Gut barrier integrity during exercise**

The integrity of gut barrier is crucial for maintaining an efficient immune system, preventing systemic inflammation and absorbing energy and nutrients. While moderate exercise can have a positive effect on the gut barrier permeability, high-intensity endurance training can reduce splanchnic perfusion, causing ischemia and leading to increased permeability [50]. It can result in endotoxemia, as lipopolysaccharides (LPS) and bacteria start circulating in the bloodstream [51]. Subsequently, increased LPS blood concentration triggers systemic inflammatory response and impacts nutrients absorption and overall athletic performance [52].

A diverse microbiome, especially rich in butyrate-producing taxa, helps maintain barrier function. Butyrate is the main energy source for colonocytes, it influences their proliferation and apoptosis, thereby impacting the integrity of the gut barrier [49]. SCFAs also upregulate mucus production and reinforce intercellular tight junctions, which restrict transit of bacteria [53]. Bacteria such as *Holdemanella biformis* can regulate the expression of gut barrier integrity markers and thereby impact its permeability [30].

Maintaining the integrity of gut barrier is essential not only for health but also for preventing performance-limiting gastrointestinal symptoms commonly reported by athletes [44].

## **4. Effects of Different Training Modalities on the Gut Microbiome**

### **4.1 Endurance training**

Endurance training induces significant shifts in microbial composition. Zhao et al. (2018) observed that long-distance endurance running results in rapid changes in the gut microbiota, for example an increase of the *Actinobacteria* phylum, that participates in the metabolism of steroids [54]. Ampe et al. (2025) showed the enhancement of *Bacteroidota* and diminution of *Firmicutes* in elite female cyclists microbiome in comparison to the non-athlete control group [55]. Moreover, Petersen et al. (2017) reported the increased *Methanobrevibacter smithii* abundance in professional cyclist, compared to amateurs. *M. smithii* contributes to metabolic efficiency of the microbiota through carbohydrate metabolism [10]. Furthermore, Shalmon et al. (2024) observed that there are differences in the composition of gut microbiome between runners and cyclists, suggesting the occurring changes are sport-specific [56]. Uchida et al. (2023) found that aerobic exercise training impacts the diversity of murine gut microbiota, promoting the increase of *Erysipelotrichaceae* and *Alcaligenaceae* families, that may have a positive effect on endurance capacity [57].

Although it is difficult to determine universal impact the endurance training has on the gut microbiota, it has been shown that some of bacteria genera, such as *Lactobacillus* and *Bifidobacterium* are typically increased in respond to exercise [50]. Moreover, Grosicki et al. (2019) observed rapid changes in ultramarathoner's microbiome after the race, suggesting that the microbiome adapts acutely to physiological demands [58].

### **4.2 Strength training**

Resistance training was historically assumed to have limited impact on the microbiome. Indeed, numerous studies have reported no significant alterations in the gut microbiome composition after strength training [59, 60]. However, Jang et al. (2019) demonstrated that strength-trained individuals exhibit distinct microbial patterns, such as increased abundance of *Sutterella*, *Clostridium*, and *Faecalibacterium* [61]. This suggests that muscular load—independently of cardiorespiratory stress—may shape microbial function. Prokopidis et al. (2023) observed that supplementation of probiotics improved muscle strength and enhanced muscle mass, suggesting bidirectional link between the gut microbiota and exercise [62].

#### **4.3 High-intensity interval training (HIIT)**

HIIT represents a unique physiological stressor combining anaerobic bursts with brief recovery cycles. Wang et al. (2025) observed that HIIT promotes enhancement of *Lactobacillus*, *Eisenbergiella*, and *Limosilactobacillus*, that may improve muscle strength, endurance capacity, and have a positive impact on cardiovascular health [63]. Nechalová et al. (2024) reported that a 12-week program led to a reduction of the *Firmicutes/Bacteroidetes* ratio and an increase in the abundance of *Akkermansia muciniphila*, and decrease of such SCFAs producers as *Erysipelatoclostridium ramosum*, *Coprococcus comes*, and *Butyrivibrio fibrisolvens* [64]. However, many studies show that HIIT does not alter the composition of the gut microbiota, although it does improve glucose metabolism, insulin sensitivity and SCFAs production [65, 66, 67]. This suggests the changes in the microbiome diversity may be the result of the overall metabolic health improvement. For example, Couvert et al. (2024) documented the correlation between the reduction of abdominal fat and occurring changes in the gut microbiota diversity in obese men after 12-week cycling or running HIIT programs [68].

#### **4.4 Team sports**

Team-sport athletes typically undergo varied training combining strength, endurance, sprinting, and tactical drills—likely explaining the complexity of their microbial profiles. Clarke et al. (2014) demonstrated that professional rugby players possess highly diverse gut microbiota compared with non-athletes [69]. These differences were attributed partly to high caloric and protein intake but also to the metabolic demands of the sport. Another research, carried out by Petri et al. (2024) highlighted the differences in the gut microbiota composition between elite soccer players and sedentary men [70]. Both groups were similar in terms of age and BMI and were following the Mediterranean diet, which involved higher amounts of fiber and lower protein intake than in the previous studies. The study revealed that the composition of gut microbiome in athletes compared to sedentary individuals included an increase in the abundance of *Roseburia hominis*, *Ruminococcaceae*, and *Prevotella albensis*, and increase in butyrate production.

### **5. The microbiome of Elite Athletes**

Elite athletes are at increased risk of systemic inflammation and developing respiratory infections due to cumulative stressors such as strenuous training, frequent travel, psychological stress, and strict dietary regimens. These factors may both challenge and enhance microbial diversity depending on context. Increasing evidence suggests that the gut microbiome

represents an additional layer of biological adaptation contributing to elite performance. Scheiman et al. (2019) provided groundbreaking evidence that *Veillonella atypica*, increased in marathon runners, metabolizes exercise-induced lactate into propionate [8]. Transplantation of this bacterium into mice improved endurance capacity, suggesting a potential performance-enhancing mechanism. A study conducted by Hintikka et al. (2022) documented that elite athletes are characterised by lower phylogenetic diversity and reduced abundance of *Akkermansia* and *Ruminococcus torques* in comparison to control group [71]. Their study also showed a healthier lipid profile in elite athletes, as well as the positive correlation between *Butyricicoccus* abundance and HDL cholesterol levels. Furthermore, a study by Li et al. (2023) revealed how the gut microbiota differs depending on the sport type, including wrestling, aerobics, and rowing, suggesting that microbiome adapts to specific training pattern [72]. In conclusion, current evidence indicates that the gut microbiome is a dynamic, adaptive system that both reflects and supports the physiological demands of elite athletic performance.

## **6. Microbiome as a Biomarker of Training Adaptation**

Growing evidence suggests that the microbiome can serve as a biomarker of training adaptation and performance capacity, and correlates with training and recovery periods.

For example, Akazawa et al. (2023) found that training periodization alters the gut microbiota composition in Japanese elite athletes [73]. The training phase impacted the abundance of *Bacteroides*, *Blautia*, *Bifidobacterium*, and *Prevotella*. Moreover, according to the study, the alteration of *Bacteroides* abundance is linked to a change in VO<sub>2</sub>max. Similarly, a research conducted by Carlone et al. (2025) showed how elite athletes gut microbiota changes depending on training, competition and recovery phases [74]. The main adaptations included fluctuations in the *Firmicutes/Bacteroidetes* ratio and increased abundance of *Rikenellaceae* in the recovery phase. Authors suggested that an analysis of the gut microbiota may be a useful tool in predicting recovery effectiveness and athletic performance.

As was mentioned earlier, a study by Uchida et al. (2023) showed that the abundance of certain bacterial families are associated with endurance performance [57]. This finding was also supported by the observation that transplantation of the gut microbiota from exercise-trained mice enhanced endurance performance of recipient.

Taken together, these studies indicate that the gut microbiome dynamically reflects training load and recovery status, making it a promising biomarker for assessing and predicting athletic adaptation and performance.

## **7. Dietary Interventions Modulating the Microbiome**

### **7.1 Probiotics**

Probiotics are live microorganisms—most commonly beneficial bacteria—that, when consumed in adequate amounts, provide health benefits to the host [75]. They support gut microbiota by introducing beneficial strains that enhance microbial balance, improve gut barrier function, reduce inflammation, and modulate immune responses [76]. Probiotics can also increase the abundance of health-promoting species and counteract disruptions caused by



various stressors, including intense training, thereby contributing to overall gut health and metabolic stability. Their effects have been widely investigated in the context of sports performance. For example, a research by Lin et al. (2020) showed that *Bifidobacterium longum* subsp. *longum* Olympic No. 1 not only increased the abundance of the microbiota, but also enhanced aerobic endurance of the participants [77].

Similarly, Huang et al. (2020) reported that supplementation of *Lactobacillus plantarum* PS128 is linked to an increased endurance capacity [78]. The research also showed an increase in abundance of *Akkermansia*, *Lactobacillus*, *Bifidobacterium*, and *Butyricimonas*, as well as higher levels of SCFAs. However, PS128 supplementation did not improve VO<sub>2</sub>max. A study conducted by Jäger et al. (2016) showed that *Bacillus oagulans* GBI-30 supplementation significantly improves recovery and reduces muscle damage following intense exercise [79]. It is also involved in maintaining performance during strenuous training. Additionally, there are numerous animal studies aiming to establish whether probiotic supplementation impacts an endurance capacity and overall physical performance. For instance, Soares et al. (2019) found that *Saccharomyces boulardii* increases VO<sub>2</sub>max and improves aerobic performance in rats [80].

## 7.2 Synbiotics

Synbiotics, a combination of probiotics and prebiotics, are designed to confer synergistic health benefits by simultaneously introducing beneficial microorganisms and providing substrates that support their growth and activity [81, 82, 83]. Evidence from human studies indicates that synbiotic supplementation can modulate selected markers of gut and immune function, such as reducing the incidence of upper-respiratory tract infections in physically active individuals and influencing inflammatory and oxidative stress parameters in clinical populations. Some data from athletic settings also suggest that certain synbiotic formulations may contribute to improved training adaptations—for example, enhancing muscle strength in competitive athletes—although mechanistic pathways such as changes in short-chain fatty acids, intestinal permeability, or nutrient absorption have not been consistently demonstrated [84, 85]. Overall, while synbiotics show potential to benefit the gut–muscle axis and support aspects of performance and recovery [86], current findings remain heterogeneous, and further well-controlled studies are needed to confirm their specific physiological and ergogenic effects.

## 7.3 Dietary patterns

Given that diet can shape the gut microbiota and the microbiota may, in turn, impact athletic performance, supporting one's gut microbes should be regarded as an effective performance-optimizing approach for athletes [87]. Athletes frequently adopt nutritional strategies focused on performance: high protein intake, carbohydrate loading, supplementation, and often low dietary fiber — a combination that may lead to unintended consequences for microbial diversity [88]. A comprehensive review dedicated to athletes concluded that while regular exercise has the potential to benefit gut microbial activity and metabolic output, common sport-centric dietary practices (high protein, low fiber, simple carbohydrate load, food avoidance) may counteract these benefits [89]. Specifically, such diets may reduce microbial diversity, suppress SCFA-producing bacteria, and shift microbial metabolism toward proteolytic fermentation, with production of potentially harmful metabolites [9].

Dietary fiber provides fundamental substrates for microbial fermentation in the colon [90]. Studies show that high-fiber diet increases the diversity of gut microbiota and therefore impacts SCFAs production. SCFAs are associated with pathways involved in muscle function and energy metabolism, and emerging evidence suggests they may influence glycogen metabolism, which could potentially affect athletic performance [91].

Protein is indispensable for muscle repair, adaptation to training, and overall recovery. For athletes, recommendations often range between 1.2 and 2.0 g/kg/day depending on training type [90]. However, when high protein intake is not balanced with sufficient dietary fiber (or fermentable carbohydrate), it may lead to a shift in microbial metabolism: from carbohydrate fermentation to proteolytic fermentation in the colon [92]. Proteolytic fermentation can generate metabolites such as ammonia, hydrogen sulfide, phenols, indoles, and other nitrogenous compounds — some of which may damage the gut mucosa, impair barrier integrity, increase colonic pH, and favor growth of less beneficial (or even harmful) bacteria. In a study of bodybuilders consuming a very high-protein, low-fiber diet, the relative abundance of SCFA- and lactic acid-producing bacteria decreased, despite regular probiotic supplementation [93]. However, dietary strategies for athletes should not only consider protein quantity, but also the quality and balance of macronutrients — ensuring that protein is complemented by sufficient fermentable carbohydrates.

A well-structured dietary pattern that balances fiber-rich carbohydrates with adequate, but not excessive, protein intake therefore appears essential for maintaining a resilient gut microbiota that can effectively support athletes' health, recovery, and overall performance.

## **8. Future Directions**

Although research on the gut microbiome in athletes has progressed considerably, many aspects of this relationship remain insufficiently understood. A key challenge for future work is the development of unified research protocols. Studies should employ comparable approaches to stool sampling, sequencing methodologies, dietary assessment, and monitoring of training loads so that results from different laboratories can be meaningfully contrasted and combined. Equally important is the need for experimental and mechanistic studies that can determine how particular microorganisms and their metabolites—such as short-chain fatty acids or bile acids—shape physiological processes relevant to sport, including endurance capacity, muscular adaptations, post-exercise recovery, and overall metabolic regulation. Achieving this will require the use of comprehensive multi-omics strategies that integrate microbial composition with metabolic outputs, gene expression, and protein function.

Another promising avenue involves examining how microbiome responses differ according to sport discipline and sex, as emerging evidence suggests that training-specific demands and biological differences create unique microbial patterns. Longitudinal studies that track athletes over extended periods—through preparation phases, competition seasons, travel, and recovery—would offer valuable insight into how the microbiota fluctuates in response to both acute stress and chronic workload.

Given the strong influence of diet on microbial ecology, nutritional factors should remain a

central theme in upcoming investigations. Researchers should explore how various macronutrient profiles, fiber intake, and microbiome-directed supplementation strategies, including probiotics, prebiotics, and synbiotics, interact with athletic training to influence performance and health. Attention should also be paid to meal timing and circadian alignment, as these factors may help stabilize microbial rhythms and support optimal physiological functioning.

Finally, the potential clinical implications of microbiome disturbances in athletes warrant deeper exploration. Increasing evidence links alterations in microbial balance with heightened intestinal permeability, systemic inflammation, weakened immunity, and possibly even greater injury risk. Understanding these associations more clearly could enable the creation of personalized interventions aimed at preserving athlete health, improving adaptation to training, and reducing susceptibility to illness or injury.

## 9. Conclusions

Current evidence demonstrates that the gut microbiome plays a crucial role in modulating physiological pathways relevant to athletic performance, including energy metabolism, immune function, inflammation, oxidative stress, and gut barrier integrity. Exercise-induced changes in microbial composition—particularly increases in beneficial SCFA-producing taxa—may enhance metabolic efficiency and recovery while reducing susceptibility to illness. However, the magnitude and direction of microbiota alterations vary substantially across individuals, sport disciplines, and environmental factors such as diet, stress, and circadian disruption. Endurance athletes exhibit the most consistent microbial adaptations, including expansion of lactate-metabolizing species linked to improved performance. Conversely, strength training and HIIT produce inconsistent microbial shifts, often mediated by dietary intake or metabolic status rather than exercise alone. While elite athletes show distinct microbial profiles, these may not universally translate into enhanced diversity or uniform health benefits. Overall, the relationship between training modalities, microbial ecology, and performance outcomes is complex and multifactorial. A definitive mechanistic model linking gut microbiota and athletic performance has not yet been established.

## Disclosure

### Author contributions

Conceptualization: **Magdalena Olszówka**, and Wiktoria Oliwia Toczek

Methodology: Urszula Janicka

Check: Klaudia Bogdan, and Mikołaj Jankowski

Investigation: Szymon Stanisław Słomiński

Data curation: Szymon Stanisław Słomiński

Writing - rough preparation: Natalia Ciepluch, and Mikołaj Jankowski

Writing - review and editing: **Magdalena Olszówka**

Visualization: Natalia Ciepluch, and Wiktoria Oliwia Toczek

Supervision: Urszula Janicka

Project administration: Klaudia Bogdan

All authors have read and agreed with the published version of the manuscript.

### **Funding Statement**

The article did not receive any funding.

### **Institutional Review Board Statement**

Not Applicable.

### **Informed Consent Statement**

Not Applicable

### **Conflict Of Interest**

Authors declare no conflict of interest.

In preparing this work, authors used ChatGPT for the purpose of improving language and readability, and text formatting. After using this tool/service, the authors have reviewed and edited the content as needed and accept full responsibility for the substantive content of the publication.

### **References**

1. Jardon KM, Canfora EE, Goossens GH, Blaak EE. Dietary macronutrients and the gut microbiome: a precision nutrition approach to improve cardiometabolic health. *Gut*. 2022 Jun;71(6):1214-1226. doi: 10.1136/gutjnl-2020-323715
2. Chen Y, Zhou J, Wang L. Role and Mechanism of Gut Microbiota in Human Disease. *Front Cell Infect Microbiol*. 2021 Mar 17;11:625913. doi: 10.3389/fcimb.2021.625913
3. Humińska-Lisowska K, Łabaj PP, Zielińska K. Unique Athletic Gut Microbiomes and Their Role in Sports Performance: A Narrative Review. *J Hum Kinet*. 2025 Oct 1;99:79-97. doi: 10.5114/jhk/202642
4. Núria Mach, Dolors Fuster-Botella. Endurance exercise and gut microbiota: A review, *Journal of Sport and Health Science*, Volume 6, Issue 2, 2017, Pages 179-197, ISSN 2095-2546, <https://doi.org/10.1016/j.jshs.2016.05.001>
5. Lin Z, Jiang T, Chen M, Ji X, Wang Y. Gut microbiota and sleep: Interaction mechanisms and therapeutic prospects. *Open Life Sci*. 2024 Jul 18;19(1):20220910. doi: 10.1515/biol-2022-0910
6. Weersma RK, Zhernakova A, Fu J. Interaction between drugs and the gut microbiome. *Gut*. 2020 Aug;69(8):1510-1519. doi: 10.1136/gutjnl-2019-320204
7. Nieman DC, Wentz LM. The compelling link between physical activity and the body's defense system. *J Sport Health Sci*. 2019 May;8(3):201-217. doi: 10.1016/j.jshs.2018.09.009

8. Scheiman J, Lubner JM, Chavkin TA, MacDonald T, Tung A, Pham LD, Wibowo MC, Wurth RC, Punthambaker S, Tierney BT, Yang Z, Hattab MW, Avila-Pacheco J, Clish CB, Lessard S, Church GM, Kostic AD. Meta-omics analysis of elite athletes identifies a performance-enhancing microbe that functions via lactate metabolism. *Nat Med*. 2019 Jul;25(7):1104-1109. doi: 10.1038/s41591-019-0485-4
9. Jang LG, Choi G, Kim SW, Kim BY, Lee S, Park H. The combination of sport and sport-specific diet is associated with characteristics of gut microbiota: an observational study. *J Int Soc Sports Nutr*. 2019 May 3;16(1):21. doi: 10.1186/s12970-019-0290-y
10. Petersen LM, Bautista EJ, Nguyen H, Hanson BM, Chen L, Lek SH, Sodergren E, Weinstock GM. Community characteristics of the gut microbiomes of competitive cyclists. *Microbiome*. 2017 Aug 10;5(1):98. doi: 10.1186/s40168-017-0320-4
11. Procházková N, Falony G, Dragsted LO, Licht TR, Raes J, Roager HM. Advancing human gut microbiota research by considering gut transit time. *Gut*. 2023 Jan;72(1):180-191. doi: 10.1136/gutjnl-2022-328166
12. Marttinen M, Ala-Jaakkola R, Laitila A, Lehtinen MJ. Gut Microbiota, Probiotics and Physical Performance in Athletes and Physically Active Individuals. *Nutrients*. 2020 Sep 25;12(10):2936. doi: 10.3390/nu12102936
13. Martin-Gallausiaux C, Marinelli L, Blottière HM, Larraufie P, Lapaque N. SCFA: mechanisms and functional importance in the gut. *Proceedings of the Nutrition Society*. 2021;80(1):37-49. doi:10.1017/S0029665120006916
14. Hays KE, Pfaffinger JM, Ryznar R. The interplay between gut microbiota, short-chain fatty acids, and implications for host health and disease. *Gut Microbes*. 2024 Jan-Dec;16(1):2393270. Doi: 10.1080/19490976.2024.2393270
15. Varghese S, Rao S, Khattak A, Zamir F, Chaari A. Physical Exercise and the Gut Microbiome: A Bidirectional Relationship Influencing Health and Performance. *Nutrients*. 2024 Oct 28;16(21):3663. doi: 10.3390/nu16213663
16. Van Hul M, Cani PD, Petitfils C, De Vos WM, Tilg H, El-Omar EM. What defines a healthy gut microbiome? *Gut*. 2024 Oct 7;73(11):1893-1908. doi: 10.1136/gutjnl-2024-333378
17. Collins SL, Stine JG, Bisanz JE, Okafor CD, Patterson AD. Bile acids and the gut microbiota: metabolic interactions and impacts on disease. *Nat Rev Microbiol*. 2023 Apr;21(4):236-247. doi: 10.1038/s41579-022-00805-x
18. Shi L, Jin L, Huang W. Bile Acids, Intestinal Barrier Dysfunction, and Related Diseases. *Cells*. 2023 Jul 19;12(14):1888. doi: 10.3390/cells12141888
19. Usuda H, Okamoto T, Wada K. Leaky Gut: Effect of Dietary Fiber and Fats on Microbiome and Intestinal Barrier. *Int J Mol Sci*. 2021 Jul 16;22(14):7613. doi: 10.3390/ijms22147613
20. Álvarez-Herms J, González A, Corbi F, Odriozola I, Odriozola A. Possible relationship between the gut leaky syndrome and musculoskeletal injuries: the important role of gut microbiota as indirect modulator. *AIMS Public Health*. 2023 Aug 22;10(3):710-738. doi: 10.3934/publichealth.2023049
21. Butler TD, Gibbs JE. Circadian Host-Microbiome Interactions in Immunity. *Front Immunol*. 2020 Aug 14;11:1783. doi: 10.3389/fimmu.2020.01783
22. Voigt RM, Forsyth CB, Green SJ, Mutlu E, Engen P, Vitaterna MH, Turek FW, Keshavarzian A. Circadian disorganization alters intestinal microbiota. *PLoS One*. 2014 May 21;9(5):e97500. doi: 10.1371/journal.pone.0097500
23. Zhao E, Tait C, Minacapelli CD, Catalano C, Rustgi VK. Circadian Rhythms, the Gut Microbiome, and Metabolic Disorders. *Gastro Hep Adv*. 2022 Feb 3;1(1):93-105. doi: 10.1016/j.gastha.2021.10.008

24. Zeng Q, Feng X, Hu Y, Su S. The human gut microbiota is associated with host lifestyle: a comprehensive narrative review. *Front Microbiol.* 2025 Jun 23;16:1549160. doi: 10.3389/fmicb.2025.1549160
25. Franzago M, Alessandrelli E, Notarangelo S, Stuppia L, Vitacolonna E. Chrono-Nutrition: Circadian Rhythm and Personalized Nutrition. *Int J Mol Sci.* 2023 Jan 29;24(3):2571. doi: 10.3390/ijms24032571
26. Wen C, Wang Q, Gu S, Jin J, Yang N. Emerging perspectives in the gut-muscle axis: The gut microbiota and its metabolites as important modulators of meat quality. *Microb Biotechnol.* 2024 Jan;17(1):e14361. Doi: 10.1111/1751-7915.14361
27. Li T, Yin D, Shi R. Gut-muscle axis mechanism of exercise prevention of sarcopenia. *Front Nutr.* 2024 Aug 16;11:1418778. doi: 10.3389/fnut.2024.1418778
28. Mancin L, Wu GD, Paoli A. Gut microbiota-bile acid-skeletal muscle axis. *Trends Microbiol.* 2023 Mar;31(3):254-269. doi: 10.1016/j.tim.2022.10.003. Epub 2022 Oct 29. Erratum in: *Trends Microbiol.* 2023 Mar;31(3):322. doi: 10.1016/j.tim.2023.01.003
29. De Vadder F, Kovatcheva-Datchary P, Zitoun C, Duchampt A, Bäckhed F, Mithieux G. Microbiota-Produced Succinate Improves Glucose Homeostasis via Intestinal Gluconeogenesis. *Cell Metab.* 2016 Jul 12;24(1):151-7. doi: 10.1016/j.cmet.2016.06.013
30. Romaní-Pérez M, López-Almela I, Bullich-Vilarrubias C, Rueda-Ruzafa L, Gómez Del Pulgar EM, Benítez-Páez A, Liebisch G, Lamas JA, Sanz Y. Holdemanella biformis improves glucose tolerance and regulates GLP-1 signaling in obese mice. *FASEB J.* 2021 Jul;35(7):e21734. doi: 10.1096/fj.202100126R
31. Larraufie, P., Martin-Gallausiaux, C., Lapaque, N. et al. SCFAs strongly stimulate PYY production in human enteroendocrine cells. *Sci Rep* 8, 74 (2018). <https://doi.org/10.1038/s41598-017-18259-0>
32. Trovão F, Correia VG, Lourenço FM, Ribeiro DO, Carvalho AL, Palma AS, Pinheiro BA. The structure of a *Bacteroides thetaiotaomicron* carbohydrate-binding module provides new insight into the recognition of complex pectic polysaccharides by the human microbiome. *J Struct Biol X.* 2023 Jan 2;7:100084. doi: 10.1016/j.yjsbx.2022.100084
33. Chambers ES, Viardot A, Psichas A, Morrison DJ, Murphy KG, Zac-Varghese SE, MacDougall K, Preston T, Tedford C, Finlayson GS, Blundell JE, Bell JD, Thomas EL, Mt-Isa S, Ashby D, Gibson GR, Kolida S, Dhillon WS, Bloom SR, Morley W, Clegg S, Frost G. Effects of targeted delivery of propionate to the human colon on appetite regulation, body weight maintenance and adiposity in overweight adults. *Gut.* 2015 Nov;64(11):1744-54. doi: 10.1136/gutjnl-2014-307913
34. Morrison DJ, Preston T. Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes.* 2016 May 3;7(3):189-200. doi: 10.1080/19490976.2015.1134082
35. Kimura I, Ozawa K, Inoue D, Imamura T, Kimura K, Maeda T, Terasawa K, Kashiwara D, Hirano K, Tani T, Takahashi T, Miyauchi S, Shioi G, Inoue H, Tsujimoto G. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. *Nat Commun.* 2013;4:1829. doi: 10.1038/ncomms2852
36. Gross K, Santiago M, Krieger JM, Hagele AM, Zielinska K, Scheiman J, Jäger R, Kostic A, Kerkick CM. Impact of probiotic *Veillonella atypica* FB0054 supplementation on anaerobic capacity and lactate. *iScience.* 2023 Dec 8;27(1):108643. doi: 10.1016/j.isci.2023.108643
37. Simpson RJ, Campbell JP, Gleeson M, Krüger K, Nieman DC, Pyne DB, Turner JE, Walsh NP. Can exercise affect immune function to increase susceptibility to infection? *Exerc Immunol Rev.* 2020;26:8-22.

38. Peake JM, Della Gatta P, Suzuki K, Nieman DC. Cytokine expression and secretion by skeletal muscle cells: regulatory mechanisms and exercise effects. *Exerc Immunol Rev.* 2015;21:8-25.
39. Jäger R, Mohr AE, Carpenter KC, Kerksick CM, Purpura M, Moussa A, Townsend JR, Lamprecht M, West NP, Black K, Gleeson M, Pyne DB, Wells SD, Arent SM, Smith-Ryan AE, Kreider RB, Campbell BI, Bannock L, Scheiman J, Wissent CJ, Pane M, Kalman DS, Pugh JN, Ter Haar JA, Antonio J. International Society of Sports Nutrition Position Stand: Probiotics. *J Int Soc Sports Nutr.* 2019 Dec 21;16(1):62. doi: 10.1186/s12970-019-0329-0
40. Rawson ES, Miles MP, Larson-Meyer DE. Dietary Supplements for Health, Adaptation, and Recovery in Athletes. *Int J Sport Nutr Exerc Metab.* 2018 Mar 1;28(2):188-199. doi: 10.1123/ijsnem.2017-0340
41. Komano Y, Shimada K, Naito H, Fukao K, Ishihara Y, Fujii T, Kokubo T, Daida H. Efficacy of heat-killed *Lactococcus lactis* JCM 5805 on immunity and fatigue during consecutive high intensity exercise in male athletes: a randomized, placebo-controlled, double-blinded trial. *J Int Soc Sports Nutr.* 2018 Aug 2;15(1):39. doi: 10.1186/s12970-018-0244-9
42. Strasser B, Geiger D, Schauer M, Gostner JM, Gatterer H, Burtscher M, Fuchs D. Probiotic Supplements Beneficially Affect Tryptophan-Kynurenine Metabolism and Reduce the Incidence of Upper Respiratory Tract Infections in Trained Athletes: A Randomized, Double-Blinded, Placebo-Controlled Trial. *Nutrients.* 2016 Nov 23;8(11):752. doi: 10.3390/nu8110752
43. Jandhyala SM, Talukdar R, Subramanyam C, Vuyyuru H, Sasikala M, Nageshwar Reddy D. Role of the normal gut microbiota. *World J Gastroenterol.* 2015 Aug 7;21(29):8787-803. doi: 10.3748/wjg.v21.i29.8787
44. Clark A, Mach N. Exercise-induced stress behavior, gut-microbiota-brain axis and diet: a systematic review for athletes. *J Int Soc Sports Nutr.* 2016 Nov 24;13:43. doi: 10.1186/s12970-016-0155-6
45. Gleeson M. Immune function in sport and exercise. *J Appl Physiol* (1985). 2007 Aug;103(2):693-9. doi: 10.1152/jappphysiol.00008.2007
46. Lamprecht M, Bogner S, Schippinger G, Steinbauer K, Fankhauser F, Hallstroem S, Schuetz B, Greilberger JF. Probiotic supplementation affects markers of intestinal barrier, oxidation, and inflammation in trained men; a randomized, double-blinded, placebo-controlled trial. *J Int Soc Sports Nutr.* 2012 Sep 20;9(1):45. doi: 10.1186/1550-2783-9-45
47. Huang WC, Wei CC, Huang CC, Chen WL, Huang HY. The Beneficial Effects of *Lactobacillus plantarum* PS128 on High-Intensity, Exercise-Induced Oxidative Stress, Inflammation, and Performance in Triathletes. *Nutrients.* 2019 Feb 7;11(2):353. doi: 10.3390/nu11020353
48. Segain JP, Raingeard de la Blétière D, Bourreille A, Leray V, Gervois N, Rosales C, Ferrier L, Bonnet C, Blottière HM, Galmiche JP. Butyrate inhibits inflammatory responses through NFkappaB inhibition: implications for Crohn's disease. *Gut.* 2000 Sep;47(3):397-403. doi: 10.1136/gut.47.3.397
49. Corrêa-Oliveira R, Fachi JL, Vieira A, Sato FT, Vinolo MA. Regulation of immune cell function by short-chain fatty acids. *Clin Transl Immunology.* 2016 Apr 22;5(4):e73. doi: 10.1038/cti.2016.17
50. Clauss M, Gérard P, Mosca A, Leclerc M. Interplay Between Exercise and Gut Microbiome in the Context of Human Health and Performance. *Front Nutr.* 2021 Jun 10;8:637010. doi: 10.3389/fnut.2021.637010
51. van Wijck K, Lenaerts K, Grootjans J, Wijnands KA, Poeze M, van Loon LJ, Dejong CH, Buurman WA. Physiology and pathophysiology of splanchnic hypoperfusion and intestinal injury during exercise: strategies for evaluation and prevention. *Am J Physiol Gastrointest Liver Physiol.* 2012 Jul 15;303(2):G155-68. doi: 10.1152/ajpgi.00066.2012

52. Di Vincenzo F, Del Gaudio A, Petito V, Lopetuso LR, Scaldaferri F. Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Intern Emerg Med*. 2024 Mar;19(2):275-293. doi: 10.1007/s11739-023-03374-w
53. Yang J, Huang Q, Long J, Li J. Microbiota and inflammatory bowel disease: the dual effect mechanism of polysaccharide therapy. *Front Immunol*. 2025 Oct 31;16:1666866. doi: 10.3389/fimmu.2025.1666866
54. Zhao X, Zhang Z, Hu B, Huang W, Yuan C, Zou L. Response of Gut Microbiota to Metabolite Changes Induced by Endurance Exercise. *Front Microbiol*. 2018 Apr 20;9:765. doi: 10.3389/fmicb.2018.00765
55. Ampe T, Decroix L, De Pauw K, Meeusen R, Demuyser T, Roelands B. Nutritional and Physiological Demands Shape the Gut Microbiome of Female World Tour Cyclists. *Microorganisms*. 2025 Oct 13;13(10):2345. doi: 10.3390/microorganisms13102345
56. Shalmon G, Ibrahim R, Israel-Elgali I, Grad M, Shlayem R, Shapira G, Shomron N, Youngster I, Scheinowitz M. Differential Gut Microbiome Profiles in Long-Distance Endurance Cyclists and Runners. *Life (Basel)*. 2024 Dec 23;14(12):1703. doi: 10.3390/life14121703
57. Uchida M, Fujie S, Yano H, Iemitsu M. Aerobic exercise training-induced alteration of gut microbiota composition affects endurance capacity. *J Physiol*. 2023 Jun;601(12):2329-2344. doi: 10.1113/JP283995
58. Grosicki GJ, Durk RP, Bagley JR. Rapid gut microbiome changes in a world-class ultramarathon runner. *Physiol Rep*. 2019 Dec;7(24):e14313. doi: 10.14814/phy2.14313
59. Moore JH, Smith KS, Chen D, Lamb DA, Smith MA, Osburn SC, Rupé BA, Morrow CD, Huggins KW, McDonald JR, Brown MD, Young KC, Roberts MD, Frugé AD. Exploring the Effects of Six Weeks of Resistance Training on the Fecal Microbiome of Older Adult Males: Secondary Analysis of a Peanut Protein Supplemented Randomized Controlled Trial. *Sports (Basel)*. 2022 Apr 22;10(5):65. doi: 10.3390/sports10050065
60. Bycura D, Santos AC, Shiffer A, Kyman S, Winfree K, Sutcliffe J, Pearson T, Sonderegger D, Cope E, Caporaso JG. Impact of Different Exercise Modalities on the Human Gut Microbiome. *Sports (Basel)*. 2021 Jan 21;9(2):14. doi: 10.3390/sports9020014
61. Wagner A, Kapounková K, Struhár I. The relationship between the gut microbiome and resistance training: a rapid review. *BMC Sports Sci Med Rehabil*. 2024 Jan 2;16(1):4. doi: 10.1186/s13102-023-00791-4
62. Prokopidis K, Giannos P, Kirwan R, Ispoglou T, Galli F, Witard OC, Triantafyllidis KK, Kechagias KS, Morwani-Mangnani J, Ticinesi A, Isanejad M. Impact of probiotics on muscle mass, muscle strength and lean mass: a systematic review and meta-analysis of randomized controlled trials. *J Cachexia Sarcopenia Muscle*. 2023 Feb;14(1):30-44. doi: 10.1002/jcsm.13132
63. Wang Y, Bai S, Yang T, Guo J, Zhu X, Dong Y. Impact of exercise-induced alterations on gut microbiota diversity and composition: comparing effects of different training modalities. *Cell Regen*. 2025 Jul 2;14(1):28. doi: 10.1186/s13619-025-00244-y
64. Nechalová L, Bielik V, Hric I, Babicová M, Baranovičová E, Grendár M, Koška J, Penesová A. Gut microbiota and metabolic responses to a 12-week caloric restriction combined with strength and HIIT training in patients with obesity: a randomized trial. *BMC Sports Sci Med Rehabil*. 2024 Dec 5;16(1):239. doi: 10.1186/s13102-024-01029-7
65. Batitucci G, Almeida OG, De Martinis ECP, Solar I, Cintra DE, de Freitas EC. Intermittent fasting and high-intensity interval training do not alter gut microbiota composition in adult women with obesity. *Am J Physiol Endocrinol Metab*. 2024 Sep 1;327(3):E241-E257. doi: 10.1152/ajpendo.00310.2023
66. Maillard F, Vazeille E, Sauvanet P, Sirvent P, Combaret L, Sourdrille A, Chavanelle V, Bonnet R, Otero YF, Delcros G, Barnich N, Boisseau N. High intensity interval training



- promotes total and visceral fat mass loss in obese Zucker rats without modulating gut microbiota. *PLoS One*. 2019 Apr 9;14(4):e0214660. doi: 10.1371/journal.pone.0214660
67. Rettedal EA, Cree JME, Adams SE, MacRae C, Skidmore PML, Cameron-Smith D, Gant N, Blenkiron C, Merry TL. Short-term high-intensity interval training exercise does not affect gut bacterial community diversity or composition of lean and overweight men. *Exp Physiol*. 2020 Aug;105(8):1268-1279. doi: 10.1113/EP088744
  68. Couvert A, Goumy L, Maillard F, Esbrat A, Lanchais K, Saugrain C, Verdier C, Doré E, Chevarin C, Adjoutah D, Morel C, Pereira B, Martin V, Lancha AH Jr, Barnich N, Chassaing B, Rance M, Boisseau N. Effects of a Cycling versus Running HIIT Program on Fat Mass Loss and Gut Microbiota Composition in Men with Overweight/Obesity. *Med Sci Sports Exerc*. 2024 May 1;56(5):839-850. doi: 10.1249/MSS.0000000000003376
  69. Clarke SF, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, Hayes P, O'Reilly M, Jeffery IB, Wood-Martin R, Kerins DM, Quigley E, Ross RP, O'Toole PW, Molloy MG, Falvey E, Shanahan F, Cotter PD. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut*. 2014 Dec;63(12):1913-20. doi: 10.1136/gutjnl-2013-306541
  70. Petri C, Mascherini G, Izzicupo P, Rosati D, Cerboneschi M, Smeazzetto S, Arrones LS. Gut microbiota and physical activity level: characterization from sedentary to soccer players. *Biol Sport*. 2024 Jul;41(3):169-176. doi: 10.5114/biol sport.2024.134759
  71. Hintikka JE, Munukka E, Valtonen M, Luoto R, Ihalainen JK, Kallonen T, Waris M, Heinonen OJ, Ruuskanen O, Pekkala S. Gut Microbiota and Serum Metabolome in Elite Cross-Country Skiers: A Controlled Study. *Metabolites*. 2022 Apr 7;12(4):335. doi: 10.3390/metabo12040335
  72. Li Y, Cheng M, Zha Y, Yang K, Tong Y, Wang S, Lu Q, Ning K. Gut microbiota and inflammation patterns for specialized athletes: a multi-cohort study across different types of sports. *mSystems*. 2023 Aug 31;8(4):e0025923. doi: 10.1128/msystems.00259-23
  73. Akazawa N, Nakamura M, Eda N, Murakami H, Nakagata T, Nanri H, Park J, Hosomi K, Mizuguchi K, Kunisawa J, Miyachi M, Hoshikawa M. Gut microbiota alternation with training periodization and physical fitness in Japanese elite athletes. *Front Sports Act Living*. 2023 Jul 14;5:1219345. doi: 10.3389/fspor.2023.1219345
  74. Carlone J, Giampaoli S, Alladio E, Rosellini G, Barni F, Salata E, Parisi A, Fasano A, Tessitore A. Dynamic stability of gut microbiota in elite volleyball athletes: microbial adaptations during training, competition and recovery. *Front Sports Act Living*. 2025 Sep 3;7:1662964. doi: 10.3389/fspor.2025.1662964
  75. Oniszczuk A, Oniszczuk T, Gancarz M, Szymańska J. Role of Gut Microbiota, Probiotics and Prebiotics in the Cardiovascular Diseases. *Molecules*. 2021 Feb 22;26(4):1172. doi: 10.3390/molecules26041172
  76. Wieërs G, Belkhir L, Enaud R, Leclercq S, Philippart de Foy JM, Dequenne I, de Timary P, Cani PD. How Probiotics Affect the Microbiota. *Front Cell Infect Microbiol*. 2020 Jan 15;9:454. doi: 10.3389/fcimb.2019.00454
  77. Lin CL, Hsu YJ, Ho HH, Chang YC, Kuo YW, Yeh YT, Tsai SY, Chen CW, Chen JF, Huang CC, Lee MC. *Bifidobacterium longum* subsp. *longum* OLP-01 Supplementation during Endurance Running Training Improves Exercise Performance in Middle- and Long-Distance Runners: A Double-Blind Controlled Trial. *Nutrients*. 2020 Jul 2;12(7):1972. doi: 10.3390/nu12071972
  78. Huang WC, Pan CH, Wei CC, Huang HY. *Lactobacillus plantarum* PS128 Improves Physiological Adaptation and Performance in Triathletes through Gut Microbiota Modulation. *Nutrients*. 2020 Aug 1;12(8):2315. doi: 10.3390/nu12082315
  79. Jäger R, Shields KA, Lowery RP, De Souza EO, Partl JM, Hollmer C, Purpura M, Wilson JM. Probiotic *Bacillus coagulans* GBI-30, 6086 reduces exercise-induced muscle damage and increases recovery. *PeerJ*. 2016 Jul 21;4:e2276. doi: 10.7717/peerj.2276

80. Soares ADN, Wanner SP, Morais ESS, Hudson ASR, Martins FS, Cardoso VN. Supplementation with *Saccharomyces boulardii* Increases the Maximal Oxygen Consumption and Maximal Aerobic Speed Attained by Rats Subjected to an Incremental-Speed Exercise. *Nutrients*. 2019 Oct 2;11(10):2352. doi: 10.3390/nu11102352
81. Zhang L, Xiao H, Zhao L, Liu Z, Chen L, Liu C. Comparison of the Effects of Prebiotics and Synbiotics Supplementation on the Immune Function of Male University Football Players. *Nutrients*. 2023 Feb 25;15(5):1158. doi: 10.3390/nu15051158.
82. Li HY, Zhou DD, Gan RY, Huang SY, Zhao CN, Shang A, Xu XY, Li HB. Effects and Mechanisms of Probiotics, Prebiotics, Synbiotics, and Postbiotics on Metabolic Diseases Targeting Gut Microbiota: A Narrative Review. *Nutrients*. 2021 Sep 15;13(9):3211. doi: 10.3390/nu13093211
83. Markowiak P, Śliżewska K. Effects of Probiotics, Prebiotics, and Synbiotics on Human Health. *Nutrients*. 2017 Sep 15;9(9):1021. doi: 10.3390/nu9091021
84. Al-Habsi N, Al-Khalili M, Haque SA, Elias M, Olqi NA, Al Uraimi T. Health Benefits of Prebiotics, Probiotics, Synbiotics, and Postbiotics. *Nutrients*. 2024 Nov 19;16(22):3955. doi: 10.3390/nu16223955
85. Naseri K, Saadati S, Ghaemi F, Ashtary-Larky D, Asbaghi O, Sadeghi A, Afrisham R, de Courten B. The effects of probiotic and synbiotic supplementation on inflammation, oxidative stress, and circulating adiponectin and leptin concentration in subjects with prediabetes and type 2 diabetes mellitus: a GRADE-assessed systematic review, meta-analysis, and meta-regression of randomized clinical trials. *Eur J Nutr*. 2023 Mar;62(2):543-561. doi: 10.1007/s00394-022-03012-9
86. Imanian B, Hemmatinafar M, Maymandinejad I, Binazade MR, Jäger R, Jahan Z, Naseri K, Rezaei R, Suzuki K. Synbiotic Supplementation with Probiotics and Omega-3 Fatty Acids Enhances Upper-Body Muscle Strength in Elite Swimmers: Evidence for Gut-Muscle Axis Modulation During Race-Pace Training. *Nutrients*. 2025 Sep 15;17(18):2959. doi: 10.3390/nu17182959
87. Nay K, Jollet M, Goustard B, Baati N, Vernus B, Pontones M, Lefeuvre-Orfila L, Bendavid C, Rué O, Mariadassou M, Bonniieu A, Ollendorff V, Lepage P, Derbré F, Koechlin-Ramonatxo C. Gut bacteria are critical for optimal muscle function: a potential link with glucose homeostasis. *Am J Physiol Endocrinol Metab*. 2019 Jul 1;317(1):E158-E171. doi: 10.1152/ajpendo.00521.2018
88. Chen Y, Yang K, Xu M, Zhang Y, Weng X, Luo J, Li Y, Mao YH. Dietary Patterns, Gut Microbiota and Sports Performance in Athletes: A Narrative Review. *Nutrients*. 2024 May 26;16(11):1634. doi: 10.3390/nu16111634
89. Riley L Hughes, Hannah D Holscher. Fueling Gut Microbes: A Review of the Interaction between Diet, Exercise, and the Gut Microbiota in Athletes, *Advances in Nutrition*, Volume 12, Issue 6, 2021, Pages 2190-2215, ISSN 2161-8313, <https://doi.org/10.1093/advances/nmab077>.
90. Zhang L, Li H, Song Z, Liu Y, Zhang X. Dietary Strategies to Improve Exercise Performance by Modulating the Gut Microbiota. *Foods*. 2024; 13(11):1680. <https://doi.org/10.3390/foods13111680>
91. Okamoto T, Morino K, Ugi S, Nakagawa F, Lemecha M, Ida S, Ohashi N, Sato D, Fujita Y, Maegawa H. Microbiome potentiates endurance exercise through intestinal acetate production. *Am J Physiol Endocrinol Metab*. 2019 May 1;316(5):E956-E966. doi: 10.1152/ajpendo.00510.2018
92. Alex E Mohr, Núria Mach, Jamie Pugh, Gregory J Grosicki, Jacob M Allen, J Philip Karl, Corrie M Whisner, Mechanisms underlying alterations of the gut microbiota by exercise and their role in shaping ecological resilience, *FEMS Microbiology Reviews*, Volume 49, 2025, fuaf037, <https://doi.org/10.1093/femsre/fuaf037>

93. Son J, Jang LG, Kim BY, Lee S, Park H. The Effect of Athletes' Probiotic Intake May Depend on Protein and Dietary Fiber Intake. *Nutrients*. 2020 Sep 25;12(10):2947. doi: 10.3390/nu12102947