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Understanding the Pathophysiology of Atopic Dermatitis – insights into Immune Dysregulation and Skin Barrier Dysfunction

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Abstract:

Atopic dermatitis (AD) is a chronic inflammatory skin disease characterized by a disrupted skin barrier and immune dysregulation. The exact pathophysiology of atopic dermatitis despite extensive research remains complex. It includes genetic disorders, a defect in the epidermal barrier, an altered immune response, and disruption of the skin's microbial balance. Recent advances in research have provided deeper insights into the molecular mechanisms including the role of filaggrin mutations, Th2 cytokine-mediated inflammation, and the skin microbiome. Understanding the intricate interplay between these components is crucial for developing targeted therapeutic strategies.

Aim of the study: This review provides a comprehensive overview of the current knowledge on the pathophysiological mechanisms underlying AD, highlighting recent advances and areas for future research.

Material and methods: Comprehensive literature searches were performed across the main electronic databases of PubMed and GoogleScholar using the keywords: "atopic dermatitis", "eczema pathophysiology", "skin barrier".

Conclusions: Atopic dermatitis (AD) is a complex, chronic inflammatory skin disease with a multifaceted pathophysiology involving genetic, immunological, and environmental factors. Recent advances in understanding the molecular mechanisms underlying AD have highlighted the importance of skin barrier dysfunction, immune system dysregulation, and microbial interactions in the disease's progression.

Keywords: atopic dermatitis, skin barrier, eczema, pathophysiology

Introduction

Atopic dermatitis (AD), or atopic eczema, is a common chronic inflammatory skin disease affecting 15-20% of children and up to 1-3% of adults [1]. It is characterized by patches of dry, pruritic skin lesions usually accompanied by itch, erythema, papules, vesicles squames,

or lichenification. AD can manifest at any age in life. However, the onset of disease commonly presents before the fifth year of life (80% - 90%) [2] and is often the first step in the development of other atopic diseases ('atopic march'), characterized by a typical sequence of atopic diseases preceding the development of other allergic disorders later in life [3].

Although the pathophysiology of AD is not completely understood, numerous studies demonstrated that there is a strong genetic component, which leads to a compromised barrier function, resulting in increased transepidermal water loss (TEWL), and heightened permeability to allergens, irritants, and pathogens. This leads to immune activation that not only promotes inflammation but also further impairs barrier function, creating a vicious cycle of the disease. This article provides a comprehensive review of the pathophysiological mechanisms underlying AD, with a focus on recent advances. By elucidating such complex interactions, we aim to inform the development of targeted therapeutic strategies that address the causes of AD, ultimately improving patient outcomes and quality of life.

1. Genetic disorder

Recent research shows over 70 genes could be associated with AD in different populations [4]. Mutations in the human filaggrin gene (FLG) are among the most significant genetic mutations related to AD. Other variations associated with the disease include epidermal barriers, immune response mechanisms, and interleukin genes [5].

1.1 Filaggrin mutations

Filaggrin is a key protein involved in skin barrier function. During keratinocyte differentiation, profilaggrin is dephosphorylated and degraded into filaggrin monomers. Then it is aggregated in the keratin cytoskeleton to form a dense protein-lipid matrix, resulting in a fully differentiated epidermal barrier. During FLG deamination and breakdown, filaggrin monomers are degraded into amino acids, which contribute to the natural moisturizing factors (NMF), maintaining skin hydration, a low pH, and other aspects of the barrier function of the stratum corneum [6]. The less acidic pH caused by lack of NMF allows skin surface serine proteases to become activated leading to uncontrolled epidermis desquamation [7]. Moreover, there are research that shows that loss of NMFs affects the skin biome, because they favor adhesion of non-pathogenic bacteria, preventing the aggregation of *S. aureus* [8].

Because of its role in maintaining skin barrier FLG mutation currently stands out as a primary driver of atopic change. About half of the patients with moderate to severe AD have null mutations in the FLG gene [9]. Patients with AD who carry FLG mutations have more early-

onset, severe, and persistent disease [10,11]. It is important to remember that FLG mutations as such also represent a risk factor for other atopic manifestations, e.g., asthma, suggesting that FLG deficiency may have a broader systemic significance [12].

1.2. SPINK5 mutations

In 1958 Earl W. Netheron described a rare monogenic disease with AD-like lesions in the skin [13] nowadays known as Netheron Syndrome. Research by S. Chavanas et al. [14] proved that it is caused by a loss-of-funcion mutation in the SPINK5 gene. This gene encodes the serine peptidase inhibitor lymphoepithelial Kazal-type trypsin inhibitor (LEKTI), which is crucial for epidermal homeostasis. LEKTI inhibits the activity of kallikreins (KLKs), the serine protease in the epidermis [15]. Netherton Syndrome is characterized by an excessive KLKs activity, which provokes permeability barrier defects. SPINK5 mutations are associated with AD in certain populations, especially eastern Asian populations [16], which suggest the role of serine protease imbalance in the pathogenesis of AD.

1.3 Other mutations

The above-described genetic mutations are not the only ones that are associated with an increased risk of atopic dermatitis. Besides FLG, other genes involved in maintaining skin barrier were identified to be associated with AD. Research has included so far genes for LAMA3, TMEM79, filaggrin-2(FLG2) and Late Cornified Envelope-like Proline-rich 1 (LELP1) [17-19]

Connection between atopic dermatitis and genes encoding the Th2 pathway has also been replicated by multiple independent studies. Genes whose polymorphisms contribute to the development of AD include IL-4, IL13 IL-31, IL-33, signal transducer and activator of transcription 6 (STAT6), thymic stromal lymphopoietin (TSLP) and its receptors: IL-7R and TSLPR [20-22]

Additionally, recent studies demonstrated that vitamin D receptor polymorphisms and cytochrome P450 variants are also associated with AD. In this regard, vitamin D receptor (VDR) polymorphisms and CYD27A1 were found to be associated with AD severity [23].

2. Lipids

The main lipid classes in human stratum corneum (SC) are ceramides (CERs), cholesterol (CHOL), and free fatty acids (FFAs). These lipid classes are present in an approximately equal molar ratio and constitute the lipid matrix that is organized in lamellar bodies and located between corneocytes [24] The introduction of liquid chromatography combined with

mass spectrometry provided a rapid boost in research leading to discovering and describing 12 different subclasses by 2010 [25]. Each CER consists of at least one acyl chain chemically linked to a sphingoid base of either sphingosine (S), phytosphingosine (P), 6-hydroxy-sphingosine, (H) or dihydrosphingosine (DS) base. The acyl chain is either a nonhydroxyfatty acid (N), an α -hydroxy fatty acid (A), or an esterified ω -hydroxy fatty acid (EO). The CER-EO subclass is often referred as acyl-CERs and contains an additional fatty acid attached to the acyl chain.

2.1. Lipid Composition

When focusing on the ratios between the various classes of lipids research [26] has reported a reduced CER/CHOL ratio in lesional AD skin. Imokawa et al. proved that the level of CER-EOS was decreased in lesional as well as in nonlesional skin [27]. Investigations of the CER composition led to the conclusion that all changes in nonlesional skin were also observed in lesional skin, but the changes were more aggravated [28]. A recent study showed that the average chain length in nonlesional skin in patients with AD was reduced compared with controls [29].

2.2 Lipid Organization

Not only lipid composition but also organization is crucial for the skin barrier function. Human SC lipids assemble in a dense, orthorhombic, lateral packing [30]. In the research on nonlesional AD skin significant increase in the level of lipids adopting the hexagonal lateral packing was observed [29]. As the hexagonal packing is less dense than the orthorhombic lateral packing, this change in lateral packing may result in a higher permeation through the lipid domains in the SC [31]. It was the first study to demonstrate that these modulations occur already in nonlesional AD skin.

This is an important observation as this may implicate that a normalization of the lipid composition in nonlesional skin may contribute to the restoration of the barrier function and thus reduce the reoccurence of lesional skin in AD patients.

3. Microbiome

3.1 Staphylococcus aureus

It has been proven that the composition and diversity of microorganisms on the skin differ between people with eczema and those healthy ones. In atopic skin, there has been a reduction in commensal bacteria of the genera *Streptococcus, Corynebacterium, Cutibacterium*, and the type *Proteobacteria* with the increase towards the genus *Staphylococcus* (*S. aureus* in

particular) [32]. Also, the total bacterial load is significantly greater on AD skin compared with healthy control skin.

Staphylococcus aureus colonized the skin in 60–100% of AD patients compared with 5–30% of healthy controls [33]. There is evidence of a loss of community diversity preceding flares of AD. Moreover, a higher colonization index and increased pathogen density show a positive correlation between the skin lesions' severity and the severity of the disease [34].

Research has proven that *S. aureus* damages and penetrates the epidermal barrier [35]. Staphylococcal enterotoxin B increases the expression of IL-31, which lowers the level of fillagrin and can induce T-cell–independent B-cell expansion, upregulate proinflammatory cytokines, and stimulate mast cell degranulation [36]. Moreover, by realising proteases it can affect and deformed corneocytes leading to stratum corneum dissolution. All of the above contribute to Th2 overexpress, skin inflammation, and itching leading to further *S. aureus* colonization.

3.2 Commensal bacteria

Studies suggested that normal microbiota commensals may modulate skin resistance and protect against AD development. For example, *Staphylococcus epidermidis* causes keratinocytes to produce antimicrobial peptides, and these suppress cytokine release after minor epidermal injury. Thus, *S. epidermidis* contributes as a barrier against colonization of pathogenic microbes [37].

Therapeutic interventions such as topical treatments with corticosteroids, calcineurin inhibitors, or even moisturizers and emollients in patients with atopic eczema may restore barrier function and normalize the skin microbiome.

4. Immunological dysregulation

The impaired skin barrier allows for the penetration of allergens and microbes, which activate the innate immune system and subsequently the adaptive immune system. This leads to the production of cytokines and chemokines that perpetuate the inflammatory response, recruit more immune cells, and further damage the skin barrier.

4.1. Innate Immune Response

The innate immune system is the first line of defense against pathogens and is crucial for maintaining skin homeostasis. It consists of the epidermal barrier, cells of the immune system, cytokines, pattern recognition receptors (PRR), antimicrobial peptides and skin microbes [38]. A recent study showed that the skin of AD patients has an over-representation of innate immunity and angiogenesis markers [39]. There is a three to four times increased percentage

of proliferating Langerhans cells in atopic skin than in healthy control. On the contrary, patients with AD have reduced levels of Antimicrobial Peptides (AMPs), contributing to an increased risk of skin infections, which induce pro-inflammatory cytokines by activating TLR2 [40]. They can also enhance the upregulation of FceRI (high-affinity IgE receptor), which binds immunoglobins on skin dendritic cells and enhances the immune response and the skin's inflammatory reaction.

The skin barrier disruption activates inflammatory epidermal dendritic cells and stimulates keratinocytes to produce proinflammatory cytokines such as TSLP (Thymic stromal lymphopoietin), IL-25, L-33, and type 2 mediated responses [41]. TSLP activates immature dendritic cells and enhances the maturation of antigen-presenting cells (APCs). Research shows that TSLP is highly expressed in the skin of patients with AD, and its production is triggered by exposure to environmental factors such as allergens, microorganisms, diesel exhaust, cigarette smoke, and chemical irritants [42]. Korean birth cohort study showed elevated expression of TSLP in the skin of 2-month-old infants before the development of clinical AD at 24 months of age [43]. IL-25 induces the expression of various chemokines: eotaxin, CCL17, (thymus and activation-regulated chemokine), and MDC (macrophage-derived chemokine), which are necessary for the recruitment of eosinophils and Th2 cells. IL-33 activates NF-kB (nuclear factor kappa-light-chain-enhancer of activated B cells) and MAP (mitogen-activated protein kinases), which stimulates the production of cytokines related to Th2 response (such as IL-4, IL-5, and IL-13) [38].

4.2. Adaptive Immune Response

The adaptive immune response in the early phases of the disease process in AD is predominantly characterized by a Th2 signal. Switching from Th2 to Th1 seems to promote chronicity of the disease [44].

External irritants and damage to the epidermal barrier lead to the stimulation of the Th2 response. Activated Th2 cells secrete cytokines such as IL-4, IL-5, and IL-13. IL-4 and IL-13 promote IgE class switching in B cells via the signal transducer and activator of the transcription (STAT) pathway [45], leading to elevated serum IgE levels. IL-5 is involved in the recruitment and activation of eosinophils in peripheral blood and tissues [46]. In addition mentioned earlier cytokines also regulate the proteins needed in the epidermal barrier. IL-4 and IL-13 stimulation were both shown to inhibit FLG function and production, with an effect

strong enough to simulate a loss of function mutation in FLG already discussed above [47]. Another study demonstrates how IL-4 upregulates, via the JAK/STAT pathway, the expression and function of the histamine receptor 4, expressed on peripheral eosinophils [48].

5. Skin barrier dysfunction

Numerous studies have demonstrated that skin barrier dysfunction is a critical component of atopic dermatitis. An impaired barrier allows external antigens and toxins to penetrate the skin and induce inflammation. This defect causes degradation of intercellular connections, higher protease activity, increased epidermal permeability, infiltration of antigens, and stimulation of proinflammatory cytokines [49].

Keratinocytes of patients with AD show an increase in the apoptosis process. Genes related to apoptosis such as NOD2, DUSP1, and ADM were all induced by the IFN-γ. The loss of skin cells due to apoptosis may cause gaps in the skin barrier allowing increased water loss and antigen penetration [50].

5.1. Tight junctions

Tight junctions are groups of adhesive proteins placed on opposing membranes of keratinocytes in SC. They are crucial in maintaining skin barrier integrity as they control the passage of fluids and solutes paracellularly acting as a selective barrier. The components of tight junctions include claudins, occludins, tricellulin, zonula occludens, and junction adhesion molecules [51] Study conducted by De Benedetto et al. [52] shows a striking reduction in expression of TJ skin proteins in AD patients. Moreover, the expression of claudin-1 correlated with Th 2 cytokines, serum IgE, and serum eosinophils [53]. In another study on animal models, it was revealed that upregulation of claudin-1 attenuated the severity and natural course of AD. Furthermore, reduced expression of tight junction proteins such as claudin-1 and 4 may be caused by cytokines such as IL-17 and bacterial infection [54].

5.2. Increase TEWL

Cutaneous barrier lesion leads to a significant increase in transepidermal water loss (TEWL) [55]. Stratum corneum hydration declines in both lesional and nonlesional AD skin, moreover it is suggested that a high TEWL may correlate with disease severity. Dry skin promotes the survival and replication of invasive staphylococci and inhibits the growth of commensal organisms, which leads to disruption of the skin microbiome [56]. It also stimulates epidermal hyperplasia and early evidence of inflammation (eg, mast cell degranulation), even in normal skin.

Regular application of emollients has been reported to reduce TEWL, enhance skin hydration, and improve general skin barrier function.

6. Conclusion

Atopic dermatitis is the most common inflammatory skin condition that affects both children and adults. Multiple factors, including epidermal gene mutations, skin barrier dysfunction, immune dysregulation, altered lipid composition, and microbial imbalance, can contribute to the development of the disease. Recently, there has been substantial progress in understanding the pathogenesis of AD. These new insights related to the genetic, immunologic, and environmental impacts have paved the way for future novel treatments. Early diagnosis and treatment may help decrease the morbidity of the disease and prevent progression to other associated atopic diseases. Further advances in this matter will allow us to achieve a precision medicine approach to the prevention and treatment of AD.

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References:

- 1. Laughter MR, Maymone MBC, Mashayekhi S, Arents BWM, Karimkhani C, Langan SM, Dellavalle RP, Flohr C. The global burden of atopic dermatitis: lessons from the Global Burden of Disease Study 1990-2017. Br J Dermatol. 2021 Feb;184(2):304-309. doi: 10.1111/bjd.19580. Epub 2020 Nov 29. PMID: 33006135.
- 2. Weidinger S, Beck LA, Bieber T, Kabashima K, Irvine AD. Atopic dermatitis. Nat Rev Dis Primers. 2018 Jun 21;4(1):1. doi: 10.1038/s41572-018-0001-z. PMID: 29930242.
- 3. Hill DA, Spergel JM. The atopic march: Critical evidence and clinical relevance. Ann Allergy Asthma Immunol. 2018 Feb;120(2):131-137. doi: 10.1016/j.anai.2017.10.037. Erratum in: Ann Allergy Asthma Immunol. 2018 Apr;120(4):451. doi: 10.1016/j.anai.2018.02.033. PMID: 29413336; PMCID: PMC5806141.
- 4. Sliz E, Huilaja L, Pasanen A, Laisk T, Reimann E, Mägi R; FinnGen; Estonian Biobank Research Team; Hannula-Jouppi K, Peltonen S, Salmi T, Koulu L, Tasanen K, Kettunen J. Uniting biobank resources reveals novel genetic pathways modulating susceptibility for atopic dermatitis. J Allergy Clin Immunol. 2022 Mar;149(3):1105-1112.e9. doi: 10.1016/j.jaci.2021.07.043. Epub 2021 Aug 27. PMID: 34454985.
- 5. Liang Y, Chang C, Lu Q. The Genetics and Epigenetics of Atopic Dermatitis-Filaggrin and Other Polymorphisms. Clin Rev Allergy Immunol. 2016 Dec;51(3):315-328. doi: 10.1007/s12016-015-8508-5. PMID: 26385242.
- 6. Brown SJ, Elias MS, Bradley M. Genetics in Atopic Dermatitis: Historical Perspective and Future Prospects. Acta Derm Venereol. 2020 Jun 9;100(12):adv00163. doi: 10.2340/00015555-3513. PMID: 32412647; PMCID: PMC9189740.
- 7. Brattsand M, et al. A proteolytic cascade of kallikreins in the stratum corneum. J Invest Dermatol. 2005;124(1):198–203.
- 8. Cork MJ, Danby SG, Vasilopoulos Y, Hadgraft J, Lane ME, Moustafa M, Guy RH, Macgowan AL, Tazi-Ahnini R, Ward SJ. Epidermal barrier dysfunction in atopic

- dermatitis. J Invest Dermatol. 2009 Aug;129(8):1892-908. doi: 10.1038/jid.2009.133. Epub 2009 Jun 4. PMID: 19494826.
- 9. Brown SJ, McLean WH. Eczema genetics: current state of knowledge and future goals. J Invest Dermatol. 2009 Mar;129(3):543-52. doi: 10.1038/jid.2008.413. PMID: 19209157.
- 10. Brown SJ, Sandilands A, Zhao Y, Liao H, Relton CL, Meggitt SJ, Trembath RC, Barker JN, Reynolds NJ, Cordell HJ, McLean WH. Prevalent and low-frequency null mutations in the filaggrin gene are associated with early-onset and persistent atopic eczema. J Invest Dermatol. 2008 Jun;128(6):1591-4. doi: 10.1038/sj.jid.5701206. Epub 2007 Dec 20. PMID: 18094728.
- 11. Weidinger S, Rodríguez E, Stahl C, Wagenpfeil S, Klopp N, Illig T, Novak N. Filaggrin mutations strongly predispose to early-onset and extrinsic atopic dermatitis. J Invest Dermatol. 2007 Mar;127(3):724-6. doi: 10.1038/sj.jid.5700630. Epub 2006 Nov 9. PMID: 17096018.
- Weidinger S, O'Sullivan M, Illig T, Baurecht H, Depner M, Rodriguez E, Ruether A, Klopp N, Vogelberg C, Weiland SK, McLean WH, von Mutius E, Irvine AD, Kabesch M. Filaggrin mutations, atopic eczema, hay fever, and asthma in children. J Allergy Clin Immunol. 2008 May;121(5):1203-1209.e1. doi: 10.1016/j.jaci.2008.02.014. Epub 2008 Apr 8. PMID: 18396323.
- 13. NETHERTON EW. A unique case of trichorrhexis nodosa; bamboo hairs. AMA Arch Derm. 1958 Oct;78(4):483-7. doi: 10.1001/archderm.1958.01560100059009. PMID: 13582191.
- 14. Chavanas S, Bodemer C, Rochat A, Hamel-Teillac D, Ali M, Irvine AD, Bonafé JL, Wilkinson J, Taïeb A, Barrandon Y, Harper JI, de Prost Y, Hovnanian A. Mutations in SPINK5, encoding a serine protease inhibitor, cause Netherton syndrome. Nat Genet. 2000 Jun;25(2):141-2. doi: 10.1038/75977. PMID: 10835624.
- 15. Hachem JP, Wagberg F, Schmuth M, Crumrine D, Lissens W, Jayakumar A, Houben E, Mauro TM, Leonardsson G, Brattsand M, Egelrud T, Roseeuw D, Clayman GL, Feingold KR, Williams ML, Elias PM. Serine protease activity and residual LEKTI expression determine phenotype in Netherton syndrome. J Invest Dermatol. 2006 Jul;126(7):1609-21. doi: 10.1038/sj.jid.5700288. Epub 2006 Apr 6. PMID: 16601670.
- 16. Nishio Y, Noguchi E, Shibasaki M, Kamioka M, Ichikawa E, Ichikawa K, Umebayashi Y, Otsuka F, Arinami T. Association between polymorphisms in the SPINK5 gene and atopic dermatitis in the Japanese. Genes Immun. 2003 Oct;4(7):515-7. doi: 10.1038/sj.gene.6363889. PMID: 14551605.
- 17. Stemmler S, Parwez Q, Petrasch-Parwez E, Epplen JT, Hoffjan S. Association of variation in the LAMA3 gene, encoding the alpha-chain of laminin 5, with atopic dermatitis in a German case-control cohort. BMC Dermatol. 2014 Nov 3;14:17. doi: 10.1186/1471-5945-14-17. PMID: 25363238; PMCID: PMC4221780.
- 18. Margolis DJ, Gupta J, Apter AJ, Ganguly T, Hoffstad O, Papadopoulos M, Rebbeck TR, Mitra N. Filaggrin-2 variation is associated with more persistent atopic dermatitis in African American subjects. J Allergy Clin Immunol. 2014 Mar;133(3):784-9. doi:

- 10.1016/j.jaci.2013.09.015. Epub 2013 Nov 1. PMID: 24184149; PMCID: PMC3943564.
- 19. Trzeciak M, Wesserling M, Bandurski T, Glen J, Nowicki R, Pawelczyk T. Association of a Single Nucleotide Polymorphism in a Late Cornified Envelope-like Proline-rich 1 Gene (LELP1) with Atopic Dermatitis. Acta Derm Venereol. 2016 May;96(4):459-63. doi: 10.2340/00015555-2301. PMID: 26608070.
- 20. He JQ, Chan-Yeung M, Becker AB, Dimich-Ward H, Ferguson AC, Manfreda J, Watson WT, Sandford AJ. Genetic variants of the IL13 and IL4 genes and atopic diseases in at-risk children. Genes Immun. 2003 Jul;4(5):385-9. doi: 10.1038/sj.gene.6363985. PMID: 12847555.
- 21. Weidinger S, Klopp N, Wagenpfeil S, Rümmler L, Schedel M, Kabesch M, Schäfer T, Darsow U, Jakob T, Behrendt H, Wichmann HE, Ring J, Illig T. Association of a STAT 6 haplotype with elevated serum IgE levels in a population based cohort of white adults. J Med Genet. 2004 Sep;41(9):658-63. doi: 10.1136/jmg.2004.020263. PMID: 15342695; PMCID: PMC1735893.
- 22. Gao PS, Rafaels NM, Mu D, Hand T, Murray T, Boguniewicz M, Hata T, Schneider L, Hanifin JM, Gallo RL, Gao L, Beaty TH, Beck LA, Leung DY, Barnes KC. Genetic variants in thymic stromal lymphopoietin are associated with atopic dermatitis and eczema herpeticum. J Allergy Clin Immunol. 2010 Jun;125(6):1403-1407.e4. doi: 10.1016/j.jaci.2010.03.016. Epub 2010 May 13. PMID: 20466416; PMCID: PMC2925504.
- 23. Heine G, Hoefer N, Franke A, Nöthling U, Schumann RR, Hamann L, Worm M. Association of vitamin D receptor gene polymorphisms with severe atopic dermatitis in adults. Br J Dermatol. 2013 Apr;168(4):855-8. doi: 10.1111/bjd.12077. Epub 2013 Mar 13. PMID: 23034014.
- 24. Weerheim A, Ponec M. Determination of stratum corneum lipid profile by tape stripping in combination with high-performance thin-layer chromatography. Arch Dermatol Res. 2001 Apr;293(4):191-9. doi: 10.1007/s004030100212. PMID: 11380152.
- 25. t'Kindt R, Jorge L, Dumont E, Couturon P, David F, Sandra P, Sandra K. Profiling and characterizing skin ceramides using reversed-phase liquid chromatography-quadrupole time-of-flight mass spectrometry. Anal Chem. 2012 Jan 3;84(1):403-11. doi: 10.1021/ac202646v. Epub 2011 Dec 7. PMID: 22111752.
- 26. Angelova-Fischer I, Mannheimer AC, Hinder A, Ruether A, Franke A, Neubert RH, Fischer TW, Zillikens D. Distinct barrier integrity phenotypes in filaggrin-related atopic eczema following sequential tape stripping and lipid profiling. Exp Dermatol. 2011 Apr;20(4):351-6. doi: 10.1111/j.1600-0625.2011.01259.x. PMID: 21410766.
- 27. Imokawa G, Abe A, Jin K, Higaki Y, Kawashima M, Hidano A. Decreased level of ceramides in stratum corneum of atopic dermatitis: an etiologic factor in atopic dry skin? J Invest Dermatol. 1991 Apr;96(4):523-6. doi: 10.1111/1523-1747.ep12470233. PMID: 2007790.
- 28. van Smeden J, Janssens M, Kaye EC, Caspers PJ, Lavrijsen AP, Vreeken RJ, Bouwstra JA. The importance of free fatty acid chain length for the skin barrier

- function in atopic eczema patients. Exp Dermatol. 2014 Jan;23(1):45-52. doi: 10.1111/exd.12293. PMID: 24299153.
- 29. Janssens M, van Smeden J, Gooris GS, Bras W, Portale G, Caspers PJ, Vreeken RJ, Hankemeier T, Kezic S, Wolterbeek R, Lavrijsen AP, Bouwstra JA. Increase in short-chain ceramides correlates with an altered lipid organization and decreased barrier function in atopic eczema patients. J Lipid Res. 2012 Dec;53(12):2755-66. doi: 10.1194/jlr.P030338. Epub 2012 Sep 28. PMID: 23024286; PMCID: PMC3494247.
- 30. Mak VH, Potts RO, Guy RH. Percutaneous penetration enhancement in vivo measured by attenuated total reflectance infrared spectroscopy. Pharm Res. 1990 Aug;7(8):835-41. doi: 10.1023/a:1015960815578. PMID: 2235880.
- 31. Mojumdar EH, Helder RW, Gooris GS, Bouwstra JA. Monounsaturated fatty acids reduce the barrier of stratum corneum lipid membranes by enhancing the formation of a hexagonal lateral packing. Langmuir. 2014 Jun 10;30(22):6534-43. doi: 10.1021/la500972w. Epub 2014 May 27. PMID: 24818519.
- 32. Zheng Y, Wang Q, Ma L, Chen Y, Gao Y, Zhang G, Cui S, Liang H, He C, Song L. Alterations in the skin microbiome are associated with disease severity and treatment in the perioral zone of the skin of infants with atopic dermatitis. Eur J Clin Microbiol Infect Dis. 2019 Sep;38(9):1677-1685. doi: 10.1007/s10096-019-03598-9. Epub 2019 May 31. PMID: 31152265.
- 33. Shi B, Leung DYM, Taylor PA, Li H. Methicillin-Resistant Staphylococcus aureus Colonization Is Associated with Decreased Skin Commensal Bacteria in Atopic Dermatitis. J Invest Dermatol. 2018 Jul;138(7):1668-1671. doi: 10.1016/j.jid.2018.01.022. Epub 2018 Feb 8. PMID: 29410379; PMCID: PMC6019637.
- 34. Tauber M, Balica S, Hsu CY, Jean-Decoster C, Lauze C, Redoules D, Viodé C, Schmitt AM, Serre G, Simon M, Paul CF. Staphylococcus aureus density on lesional and nonlesional skin is strongly associated with disease severity in atopic dermatitis. J Allergy Clin Immunol. 2016 Apr;137(4):1272-1274.e3. doi: 10.1016/j.jaci.2015.07.052. Epub 2015 Nov 11. PMID: 26559326.
- 35. Spaulding AR, Salgado-Pabón W, Kohler PL, Horswill AR, Leung DY, Schlievert PM. Staphylococcal and streptococcal superantigen exotoxins. Clin Microbiol Rev. 2013 Jul;26(3):422-47. doi: 10.1128/CMR.00104-12. PMID: 23824366; PMCID: PMC3719495.
- 36. Skov L, Olsen JV, Giorno R, Schlievert PM, Baadsgaard O, Leung DY. Application of Staphylococcal enterotoxin B on normal and atopic skin induces up-regulation of T cells by a superantigen-mediated mechanism. J Allergy Clin Immunol. 2000 Apr;105(4):820-6. doi: 10.1067/mai.2000.105524. PMID: 10756235.
- 37. Iwase T, Uehara Y, Shinji H, Tajima A, Seo H, Takada K, Agata T, Mizunoe Y. Staphylococcus epidermidis Esp inhibits Staphylococcus aureus biofilm formation and nasal colonization. Nature. 2010 May 20;465(7296):346-9. doi: 10.1038/nature09074. PMID: 20485435.

- 38. Sroka-Tomaszewska J, Trzeciak M. Molecular Mechanisms of Atopic Dermatitis Pathogenesis. Int J Mol Sci. 2021 Apr 16;22(8):4130. doi: 10.3390/ijms22084130. PMID: 33923629; PMCID: PMC8074061.
- 39. Tsoi LC, Rodriguez E, Stölzl D, Wehkamp U, Sun J, Gerdes S, Sarkar MK, Hübenthal M, Zeng C, Uppala R, Xing X, Thielking F, Billi AC, Swindell WR, Shefler A, Chen J, Patrick MT, Harms PW, Kahlenberg JM, Perez White BE, Maverakis E, Gudjonsson JE, Weidinger S. Progression of acute-to-chronic atopic dermatitis is associated with quantitative rather than qualitative changes in cytokine responses. J Allergy Clin Immunol. 2020 May;145(5):1406-1415. doi: 10.1016/j.jaci.2019.11.047. Epub 2019 Dec 28. Erratum in: J Allergy Clin Immunol. 2023 May;151(5):1413. doi: 10.1016/j.jaci.2023.02.019. PMID: 31891686; PMCID: PMC7214216.
- 40. Werfel T, Allam JP, Biedermann T, Eyerich K, Gilles S, Guttman-Yassky E, Hoetzenecker W, Knol E, Simon HU, Wollenberg A, Bieber T, Lauener R, Schmid-Grendelmeier P, Traidl-Hoffmann C, Akdis CA. Cellular and molecular immunologic mechanisms in patients with atopic dermatitis. J Allergy Clin Immunol. 2016 Aug;138(2):336-49. doi: 10.1016/j.jaci.2016.06.010. PMID: 27497276.
- 41. Tsakok T, Woolf R, Smith CH, Weidinger S, Flohr C. Atopic dermatitis: the skin barrier and beyond. Br J Dermatol. 2019 Mar;180(3):464-474. doi: 10.1111/bjd.16934. Epub 2018 Oct 10. PMID: 29969827.
- 42. Lee EB, Kim KW, Hong JY, Jee HM, Sohn MH, Kim KE. Increased serum thymic stromal lymphopoietin in children with atopic dermatitis. Pediatr Allergy Immunol. 2010 Mar;21(2 Pt 2):e457-60. doi: 10.1111/j.1399-3038.2009.00919.x. PMID: 20444170.
- 43. Kim J, Kim BE, Lee J, Han Y, Jun HY, Kim H, Choi J, Leung DYM, Ahn K. Epidermal thymic stromal lymphopoietin predicts the development of atopic dermatitis during infancy. J Allergy Clin Immunol. 2016 Apr;137(4):1282-1285.e4. doi: 10.1016/j.jaci.2015.12.1306. Epub 2016 Feb 12. PMID: 26879860.
- 44. Gittler JK, Shemer A, Suárez-Fariñas M, Fuentes-Duculan J, Gulewicz KJ, Wang CQ, Mitsui H, Cardinale I, de Guzman Strong C, Krueger JG, Guttman-Yassky E. Progressive activation of T(H)2/T(H)22 cytokines and selective epidermal proteins characterizes acute and chronic atopic dermatitis. J Allergy Clin Immunol. 2012 Dec;130(6):1344-54. doi: 10.1016/j.jaci.2012.07.012. Epub 2012 Aug 27. PMID: 22951056; PMCID: PMC3991245.
- 45. Gandhi NA, Bennett BL, Graham NM, Pirozzi G, Stahl N, Yancopoulos GD. Targeting key proximal drivers of type 2 inflammation in disease. Nat Rev Drug Discov. 2016 Jan;15(1):35-50. doi: 10.1038/nrd4624. Epub 2015 Oct 16. PMID: 26471366.
- 46. Bieber T. Atopic dermatitis: an expanding therapeutic pipeline for a complex disease. Nat Rev Drug Discov. 2022 Jan;21(1):21-40. doi: 10.1038/s41573-021-00266-6. Epub 2021 Aug 20. PMID: 34417579; PMCID: PMC8377708.
- 47. Furue M, Tsuji G, Mitoma C, Nakahara T, Chiba T, Morino-Koga S, Uchi H. Gene regulation of filaggrin and other skin barrier proteins via aryl hydrocarbon receptor. J

- Dermatol Sci. 2015 Nov;80(2):83-8. doi: 10.1016/j.jdermsci.2015.07.011. Epub 2015 Jul 26. PMID: 26276439.
- 48. Schaper-Gerhardt K, Köther B, Wolff L, Kabatas A, Gehring M, Nikolouli E, Mommert S, Werfel T, Gutzmer R. The H4 R is highly expressed on eosinophils from AD patients and IL-4 upregulates expression and function via the JAK/STAT pathway. Allergy. 2021 Apr;76(4):1261-1264. doi: 10.1111/all.14599. Epub 2020 Oct 7. PMID: 32975872.
- 49. Agrawal R, Woodfolk JA. Skin barrier defects in atopic dermatitis. Curr Allergy Asthma Rep. 2014 May;14(5):433. doi: 10.1007/s11882-014-0433-9. PMID: 24633617; PMCID: PMC4034059.
- 50. Rebane A, Zimmermann M, Aab A, Baurecht H, Koreck A, Karelson M, Abram K, Metsalu T, Pihlap M, Meyer N, Fölster-Holst R, Nagy N, Kemeny L, Kingo K, Vilo J, Illig T, Akdis M, Franke A, Novak N, Weidinger S, Akdis CA. Mechanisms of IFN-γ-induced apoptosis of human skin keratinocytes in patients with atopic dermatitis. J Allergy Clin Immunol. 2012 May;129(5):1297-306. doi: 10.1016/j.jaci.2012.02.020. Epub 2012 Mar 24. PMID: 22445417.
- 51. Zaniboni MC, Samorano LP, Orfali RL, Aoki V. Skin barrier in atopic dermatitis: beyond filaggrin. An Bras Dermatol. 2016 Jul-Aug;91(4):472-8. doi: 10.1590/abd1806-4841.20164412. PMID: 27579743; PMCID: PMC4999106.
- 52. De Benedetto A, Rafaels NM, McGirt LY, Ivanov AI, Georas SN, Cheadle C, Berger AE, Zhang K, Vidyasagar S, Yoshida T, Boguniewicz M, Hata T, Schneider LC, Hanifin JM, Gallo RL, Novak N, Weidinger S, Beaty TH, Leung DY, Barnes KC, Beck LA. Tight junction defects in patients with atopic dermatitis. J Allergy Clin Immunol. 2011 Mar;127(3):773-86.e1-7. doi: 10.1016/j.jaci.2010.10.018. Epub 2010 Dec 15. PMID: 21163515; PMCID: PMC3049863.
- 53. Tokumasu R, Yamaga K, Yamazaki Y, Murota H, Suzuki K, Tamura A, Bando K, Furuta Y, Katayama I, Tsukita S. Dose-dependent role of claudin-1 in vivo in orchestrating features of atopic dermatitis. Proc Natl Acad Sci U S A. 2016 Jul 12;113(28):E4061-8. doi: 10.1073/pnas.1525474113. Epub 2016 Jun 24. PMID: 27342862; PMCID: PMC4948351.
- 54. Bäsler K, Galliano MF, Bergmann S, Rohde H, Wladykowski E, Vidal-Y-Sy S, Guiraud B, Houdek P, Schüring G, Volksdorf T, Caruana A, Bessou-Touya S, Schneider SW, Duplan H, Brandner JM. Biphasic influence of Staphylococcus aureus on human epidermal tight junctions. Ann N Y Acad Sci. 2017 Oct;1405(1):53-70. doi: 10.1111/nyas.13418. Epub 2017 Jul 28. PMID: 28753223.
- 55. Galli E, Cinicola B, Carello R, Caimmi S, Brindisi G, De Castro G, Zicari AM, Tosca MA, Manti S, Martelli A, Calvani M, Cravidi C, Marseglia GL, Cardinale F, Miraglia Del Giudice M, Caffarelli C, Duse M. Atopic dermatitis. Acta Biomed. 2020 Sep 15;91(11-S):e2020011. doi: 10.23750/abm.v91i11-S.10313. PMID: 33004781; PMCID: PMC8023058.
- 56. Seite S, Flores GE, Henley JB, Martin R, Zelenkova H, Aguilar L, Fierer N. Microbiome of affected and unaffected skin of patients with atopic dermatitis before

and after emollient treatment. J Drugs Dermatol. 2014 Nov;13(11):1365-72. PMID: 25607704.