Nosenko O. M., Ayzyatulova D. R. Methods of preparation of endometrium in patients with ovarian hyperstimulation syndrome before the transfer of vitrified / warmed embryos. Journal of Education, Health and Sport. 2018;8(8):964-972. eISNN 2391-8306. DOI http://dx.doi.org/10.5281/zenodo.1412472

http://ojs.ukw.edu.pl/index.php/johs/article/view/5959

The journal has had 7 points in Ministry of Science and Higher Education parametric evaluation. Part b item 1223 (26/01/2017). 1223 Journal of Education, Health and Sport eISSN 2391-8306 7 © The Author(s) 2018; This article is published with open access at Licensee Open Journal Systems of Kazimierz Wielki University in Bydgoszcz, Poland Open Access. This article is distributed under the terms of the Creative Commons Attribution Noncommercial Licensee which permits any noncommercial use, distribution, and reproduction in any mediu provided the original author (s) and source are credited. This is an open access article iscensed under the terms of the Creative Commons Attribution Non commercial Licensee the true of the Creative Commons Attribution and reproduction in any medium, provided the work is properly cited. his is an open access article licensed under the terms of the Creative Commons Attribution Non commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted, non commercial License (http://

Methods of preparation of endometrium in patients with ovarian hyperstimulation syndrome before the transfer of vitrified / warmed embryos

<sup>1</sup>O. M. Nosenko, <sup>2</sup>D. R. Ayzyatulova

# <sup>1</sup>Odessa National Medical University: <sup>2</sup>Donetsk National Medical University, Lyman, Ukraine

### Abstract

The aim of the study was to develop a personified method of endometrium preparation for the transfer of vitrified / warmed embryos in women with the development of OHSS in IVF programs. 39 women with early OHSS after IVF, who were undergoing personalized correction of the morphofunctional state of the endometrium, and 39 patients with baseline OHSS II group undergoing standard therapy were examined and treated. Control group consisted of women without OHSS in the IVF cycle. Personalized correction of the morphofunctional state of the endometrium includes traditional hormonal replacement therapy and addition of adjuvants: preparation with extract of horse chestnut and thiamine hydrochloride, vitamin E, serratiopeptidase, myo-inositol, the product of proteolysis of the placenta of cattle, L-arginine aspartate. The application of the developed method for the preparation of endometrium for the transfer of vitrified / warmed embryos by the developed method contributed to the prolongation of pregnancy and the increase in the number of live births in 2,11 times (OR 3.17; 95% CI 1.20-8.39; p<0,02). The increase in absolute benefit (therapeutic benefit) of the proposed person-adjusted correction is 25.64% (95% CI 5.12-46.16%), an increase in relative benefit corresponds to a clinically significant effect of 111.10% (95% CI 9.40 – 307.40 %), the number of patients to be treated is 3.90 (95% CI 2.17-19.52). The developed method is effective for the preparation of endometrium for the transfer of vitrified / warmed embryos.

Key words: in vitro fertilization, ovarian hyperstimulation syndrome, cycle segmentation, vitrified / warmed embryos, method of endometrium preparation, live births.

In case of high risk or development of ovarian hyperstimulation syndrome (OHSS), the *in vitro* fertilization (IVF) treatment cycle can be segmented, all embryos are cryopreserved without the transfer of a fresh embryo. After restoring the morphofunctional state of the ovaries, the embryos are thawed and transferred to the uterus in the natural cycle, a cycle of replacement hormone therapy or stimulated cycle.

A meta-analysis from 11 observational studies has shown that pregnancy after the transfer of vitrified / warmed embryos is associated with a lower risk of preterm delivery, low birth weight and perinatal mortality than pregnancy from transferred a fresh embryo [16, 17]. Subsequently, other studies comparing the two procedures showed better pregnancy rates through the transfer of vitrified / warmed embryos [19, 22].

The causal relationships between the periimplanting environment and the perinatal outcomes are confirmed by several studies demonstrating that the adverse effects of the periimplantation medium are most significant in those patients who are most reactive to COS [9, 13, 14]. A recent pilot study by AN Imudia.et al. (2013) has shown that in patients at high risk of OHSS who have chosen cryopreservation of all embryos, preeclampsia and birth weight of children, defined as <10% for gestational age, were lower than those who decided to continue embryo transfer [13]. This group also showed higher rates of preeclampsia and the birth of children of low gestational age in patients with a level of  $E_2$  greater than 90 percentile, indicating that an energetic response to superovulation may be associated with a higher risk of adverse outcomes of pregnancy (OR for the birth of children with low weight - 9.40, 95% DI 3.22-27.46; OR for preeclampsia - 4.79, 95% DI 1.55-14.84) [12].

**The aim** of the study was to develop a personified method of endometrium preparation for the transfer of vitrified / warmed embryos in women with the development of OHSS in IVF programs.

## Materials and methods

A randomized controlled trial of the effectiveness of the developed measures of personified correction of the morphofunctional state of endometrium compared with the standard approach was conducted. 39 women of the I group (experimental) with early OHSS after IVF, who were undergoing personalized correction of the morphofunctional state of the endometrium, and 39 patients with baseline OHSS II group undergoing standard therapy were examined and treated. Control group K consisted of 30 women without OHSS in the IVF cycle. All groups were homogeneous according to age, anthropometric data, gynecological, reproductive, infectioous and somatic anamnesis, duration of infertility, number of IVF attempts, uterine and ovarian size, ovarian reserve and hormonal status.

The criteria for inclusion of patients in the OHSS group were: the presence of infertility and the development of early moderate or severe OHSS in the IVF cycle during COS with antagonists of gonadotropin-releasing-hormone (antGnRH); informed consent of women to participate in the study. Exclusion criteria are the presence of ovarian tumors, diabetes mellitus, acute infectious diseases.

We used the following COS methodology. From the 2nd day of the menstrual cycle, gonadotropins were started in the average starting dose of FSH 75-150 IU. At 6-8th days, they performed a sonography, when the follicle was detected > 14 mm, antGnRH was administered at a dose of 0.25 mg / day for 4-5 days and gonadotrophin stimulation continued until the leading follicle reached > 18 mm in diameter and at least three follicles sizes > 16 mm. Then a trigger of ovulation was prescribed – agonist GnRH (aGnRH) in a dose of 0.1-0.2 mg; after 34-36 hours, the egg collection and their fertilization were performed. The resulting embryos were vitrified. Women were withdrawn from the fertilization treatment cycle and received the necessary treatment for OHSS. From the first menstruation after the treatment cycle, the endometrium was preparated.

In the comparison group II, endometrial preparation was carried out according to the traditional method as follows: three months after the IVF cycle, replaced cyclic hormone therapy was performed; on the 21st day of the third menstrual cycle, 3.75 mg of tryptoreline was injected subcutaneously, after 14 days, the use of transdermal estrogens was started for 12-14 days. After reaching the thickness of the endometrium of 10-12 mm, they began taking progesterone preparations. On P+7-P+8 day, two frozen embryos were transferred at the stage of the blastocyst. Before transplantation, if necessary, pre-implantation diagnosis was performed to evaluate the genetic material and the numerical chromosome set of the embryonic cells.

For transfer, use soft, biologically inert (non-toxic) catheters, unable to damage the embryo, which made it possible to quickly and delicately place the embryo in the uterine cavity. Penetration into the uterus for the purpose of subcutaneous injection was carried out in two stages. The first – a cavernous catheter, more rigid, passed the cervical canal. The second

– a soft catheter, through the opening in the first catheter, was injected with the germ along with the fluid for cultivation in the uterus cavity. We tried to carry the transfer very smoothly, without sharp movements. We prescribed vaginal forms of progesterone to support implantation and post-implantation during pregnancy.

In group I, the preparation of endometrium was carried out according to the developed methodology, which was conducted similarly to that in group II, but with the addition of adjuvants: 12-15 drops of the preparation, 100 g of which contains 5.4 g dry extract of horse chestnut (1,0 g of triterpenglikozid (triterpensaponin), referenced to escine), 0.5 g of thiamine hydrochloride; drops were prescribed 3 times a day, before meals, with a small amount of fluid, a course of three months; vitamin E in capsules of 400 mg once a day after eating with enough fluid in the second half of the menstrual cycle, three cycles; serratiopeptidase in 10 mg pills 2 times a day, washed with a glass of water for a month; preparations of myo-inositol 2000 mg twice a day for three months; the course of intramuscular injection of the drug, 1 ml of solution containing the product of proteolysis of the placenta of cattle - 15 mg, for a course of 10 injections in a day; oral solution, 5 ml containing L-arginine aspartate 1 g (L-arginine - 0.57 g, acid aspartic acid - 0.43 g) per 5 ml per os 3-6 times a day, course of treatment 15 days.

The processing and analysis of statistical information was carried out using the IBM SPSS Statistics 22 (Statistical Package for the Social Science) software.

# **Results and discussion**

The evaluation of reproductive results in patients with OHSS was performed depending on the method of endometrial preparation before the transfer of vitrified / warmed embryos (Table).

Table

Group	Clinical Pregnancy		Live birth		Miscarriage		Ectopic pregnancy	
	n	%	n	%	n	%	n	%
I (n=39)	23	58,97	19	48,72 <sup>II</sup>	4	10,26	0	0,00
II (n=39)	18	46,15 <sup> k</sup>	9	23,08 <sup> k,I</sup>	8	20,51	1	2,56
K (n=30)	30	100	24	80,00	5	16,67	1	3,33

Clinical results of IVF in patients with OHSS after the transfer of vitrifiedwarmed embryos, depending on the methodology of endometrium preparation

Notes. <sup>k, I, II</sup> - statistically significant difference with the indicators of groups K, I, II, p <0,05.

At the incidence of pregnancy, the groups I studied (58.97% (23) and II (46.15% (18)) did not differ statistically significantly. The number of cases of live birth in group I (48.72% (19)) exceeded that in the group II (23,08%) in 2,11 times (OR 3,17 $\pm$ 0,50; 95% CI 1,20-8,39; p<0,02). Increasing the absolute benefit (therapeutic benefit) of the proposed personified correction was 25.64% (95% CI 5.12-46.16 %), the increase in relative benefit corresponded to a clinically significant effect – 111.10% (95% CI 9.40-307.40 %), the number of patients, which should be treated, – 3.90 (95% CI 2.17-19.52).

The number of miscarriages after IVF in patients with OHSS after the transfer of vitrified / warmed embryos with the preparation of the endometrium according to the developed method was the smallest and was 10.26% (4), whereas in the group II – 20.51% (8) and in the group K 16,67% (5). Ectopic pregnancies were observed in 2.56% (1) of Group II patients and in 3.33% (1) of patients in the K group; in group I, no such cases were reported.

The results obtained can be explained by the therapeutic effect of the developed method of preparing a patient with a transmitted OHSS to the transfer of vitrified / warmed embryos.

The preparation containing the dry extract of horse chestnut Aesculus hippocastanum, the active substance of which is escine, has a pronounced venotonic, anti-inflammatory and anti-edema activity in diseases of the veins. Clinical reports on the vascular efficacy of escine pay particular attention to improving microcirculation, reducing vascular permeability, increasing venous tone and venous return, all that reduces edema [7]. It was suggested that the observed effects are the result of the protection of endothelial cells from hypoxia and inflammatory stimuli. In fact, as shown in pre-clinical studies,  $\beta$ -escine retains ATP in the absence of oxygen, reduces the response of histamine, and releases cytokines, suppresses serotonin-induced capillary permeability [8], suppresses extravasation and migration of leukocytes [6] and preserves the morphology of endothelial cells [15]. It is also worth noting the data indicating the antioxidant potential of  $\beta$ -escine [10, 18] and its inhibitory effect on hyaluronidase, the enzyme is involved in the pathogenesis of chronic venous insufficiency [20]. In later inflammatory work, the softening properties  $\beta$ -escine was associated with its modulating effect on the tumor necrosis factor- $\alpha$ -mediated inflammatory route [21].

Escine is able to break the formed positive feedback, restoring normal hemocytes in venous blood vessels and preventing the development of decompensation of diseases. Anticoagulant and anti-aggregant action of the drug make its use effective for the prevention of thrombosis. The presence of an anti-inflammatory, anti-edema, capillary-resistant effect,

and the ability to reduce the permeability of the plasmolymphatic barrier provides rapid elimination of symptoms [3], which is especially important in the development of OHSS.

Vitamin E and L-arginine improve the function of the luteal body and increase the endometrium in patients by increasing the flow in the uterine artery [5].

Serratiopeptidase is a natural proteolytic enzyme isolated from the non-pathogenic Serratia E15 bacterial bacteria. It has fibrinolytic, anti-inflammatory and anti-edema activity [11]. Serratiopeptidase binds in a ratio of 1:1 to  $\alpha$ 2-macroglobulin, which masks its antigenicity but retains its enzymatic activity. Serratiopeptidase slowly passes into the exudate in the inflammation center and gradually it decreases in blood. With the help of hydrolysis, bradykinin, histamine and serotonin seratoypeptidase directly reduces dilatation of capillaries and control their permeability. Serratiopeptidase blocks the plasma inhibitors, thereby contributing to its fibrinolytic activity.

Mio-inositol is widespread in human tissues and cells and is a precursor of phosphoinositides involved in signal transduction by stimulating membrane receptors and other secondary messengers, including diacylglycerol and inositol triphosphate [23]. The importance of myo-inositol and its derivatives to maintain the physiological flow of a wide range of processes in the human body was established as a result of systematic analysis of more than 37 thousand publications. The main function of myo-inositol and its derivatives is to contribute to the intracellular signaling and to ensure the functioning of such important receptors as insulin receptors, reproductive hormones, growth factors, catecholamines, and others. The derivatives of myo-inositol interact with specific proteins that are involved in the functioning of the reproductive system and the development of the embryo. Mio-inositol is an important synergist of folates and other vitamins (B5, PP) [1].

The complex of placental regulatory peptides has an immunotropic effect: stimulates the functional capacity of phagocytes of the mucous membranes and blood, increases the synthesis of anti-inflammatory cytokines, and affects the activity of regulatory subpopulations of lymphocytes. With autoimmune diseases or syndromes, it reduces the activity of the immune-dependent inflammatory process, increases the number of CD4 + / 25 + and CD8 + / 25 + cells, especially the level of IL-10 in blood plasma. The complex of placental regulatory peptides has a pronounced anti-inflammatory and spasticity effect, reduces the intensity of destructive, infiltrative and proliferative processes in the inflammatory site, reduces the severity of edema, accelerates the processes of epithelization, regeneration, prevents the development of the adhesion process, the effect on the proliferative and exudative phases of inflammation significantly exceeds the placenta extract [4].

L-arginine aspartate has a positive effect on the expression of markers of angiogenesis, thereby participating in the formation of new blood vessels in the endometrium. In this case, the enhanced formation of nitric oxide leads to dilatation of peripheral vessels and a decrease in the overall peripheral vascular resistance, which contributes to lowering blood pressure and reducing tissue hypoxia. Increasing the enzyme activity of NO- synthase against the background of oral administration of the solution of L-arginine aspartate provides a constant basic level of nitric oxide, which in turn contributes to increasing the density of functional vessels of the endometrium, enhancing its blood supply [2].

### Conclusions

Conducting the developed preparation of endometrium for the transfer of vitrified / warmed embryos by the developed method contributes to the continuation of pregnancy and increase the number of live births.

## References

1. Gromova OA, Goncharova EA, Torshin IU, Limanova OA, Kerimkulova NV. Prospects for the use of myo-inositol in the pre-gravity preparation of women with polycystic ovary and insulin resistance. Gynecology. 2014, 1: 58-65.

2. Potapov VA. Expediency of using arginine in therapy of luteal phase insufficiency in patients of reproductive age. Health of Ukraine. 2016; 2: 2-3.

3. Prihozhaya V. Escusane is an effective agent of plant origin for the treatment of venous disease. Acute and urgent conditions in the practice of a doctor. 2010; 3 (22). Published online 2010..

4. Chaika VK, Chaika AV, Nosenko OM. Receptivity of the endometrium in patients with infertility: monograph. Donetsk: Publishing house "Nouvelage", 2011. 243 p.

5. Cicek N, Eryilmaz OG, Sarikaya E, Gulerman C, Genc Ya. Vitamin E effect on controlled ovarian stimulation of unexplained infertile women. J Assist Reprod Genet. 2012 Apr; 29(4): 325–328. Published online 2012 Feb 1. doi: 10.1007/s10815-012-9714-1.

6. Denomme MM, Mann MR. Genomic imprints as a model for the analysis of epigenetic stability during assisted reproductive technologies. Reproduction. 2012;144:393–409.

7. Domanski D, Zegrocka-Stendel O, Perzanowska A, Dutkiewicz M, Kowalewska M, Grabowska I, at al. Molecular Mechanism for Cellular Response to  $\beta$ -Escin and Its Therapeutic Implications. PLoS One. 2016 Oct 11;11(10):e0164365. doi: 10.1371/journal.pone.0164365. eCollection 2016.

970

8. Eroglu A, Layman LC. Role of ART in imprinting disorders. Semin Reprod Med. 2012;30:92–104.

9. Farhi J, Ben-Haroush A, Andrawus N, Pinkas H, Sapir O, Fisch B, et al. High serum oestradiol concentrations in IVF cycles increase the risk of pregnancy complications related to abnormal placentation. Reprod Biomed Online. 2010;21:331–7.

10. Feuer SK, Camarano L, Rinaudo PF. ART and health: clinical outcomes and insights on molecular mechanisms from rodent studies. Mol Hum Reprod. 2013;19:189–204.

11. Gupte V, Luthra U. Analytical techniques for serratiopeptidase: A review. J Pharm Anal. 2017 Aug; 7(4): 203–207. Published online 2017 Mar 22. doi: 10.1016/j.jpha.2017.03.005.

12. Imudia AN, Awonuga AO, Doyle JO, Kaimal AJ, Wright DL, Toth TL, et al. Peak serum estradiol level during controlled ovarian hyperstimulation is associated with increased risk of small for gestational age and preeclampsia in singleton pregnancies after in vitro fertilization. Fertil Steril. 2012;97:1374–9.

13. Imudia AN, Awonuga AO, Kaimal AJ, Wright DL, Styer AK, Toth TL. Elective cryopreservation of all embryos with subsequent cryothaw embryo transfer in patients at risk for ovarian hyperstimulation syndrome reduces the risk of adverse obstetric outcomes: a preliminary study. Fertil Steril. 2013;99:168–73.

14. Joo BS, Park SH, An BM, Kim KS, Moon SE, Moon HS. Serum estradiol levels during controlled ovarian hyperstimulation influence the pregnancy outcome of in vitro fertilization in a concentration-dependent manner. Fertil Steril. 2010;93:442–6.

15. Kajantie E, Hovi P. Is very preterm birth a risk factor for adult cardiometabolic disease? Semin Fetal Neonat Med. 2014;19:112–7.

16. Maheshwari A, Pandey S, Shetty A, Hamilton M, Bhattacharya S. Obstetric and perinatal outcomes in singleton pregnancies resulting from the transfer of frozen thawed versus fresh embryos generated through in vitro fertilization treatment: a systematic review and meta-analysis. Fertil Steril. 2012;98:368-377.

17. Roque M, Valle M, Kostolias A, Sampaio M, Geber S. Freeze-all cycle in reproductive medicine: current perspectives. JBRA Assist Reprod. 2017;21(1):49-53. doi: 10.5935/1518-0557.20170012.

18. Santos MA, Kuijk EW, Macklon NS. The impact of ovarian stimulation for IVF on the developing embryo. Reproduction. 2010;139:23–34.

19. Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C, Thomas S. Evidence of impaired endometrial receptivity after ovarian stimulation for in vitro

fertilization: a prospective randomized trial comparing fresh and frozen-thawed embryo transfer in normal responders. Fertil Steril. 2011;96:344-348.

20. Shapiro BS, Daneshmand ST, Garner FC, Aguirre M, Hudson C, Thomas S. Evidence of impaired endometrial receptivity after ovarian stimulation for in vitro fertilization: a prospective randomized trial comparing fresh and frozen-thawed embryo transfer in normal responders. Fertil Steril. 2011;96:344–8.

21. Shapiro BS, Daneshmand ST, Restrepo H, Garner FC, Aguirre M, Hudson C. Matched-cohort comparison of single-embryo transfers in fresh and frozen-thawed embryo transfer cycles. Fertil Steril. 2013;99:389–92.

22. Shi Y, Wei D, Liang X, et al. Live birth after fresh embryo transfer vs elective embryo cryopreservation/frozen embryo transfer in women with polycystic ovary syndrome undergoing IVF (FreFro-PCOS): study protocol for a multicenter, prospective, randomized controlled clinical trial. Trials. 2014;15:154-154.

23. Simi G, Genazzani AR, Obino MER, Papini F, Pinelli S, Cela V, Artini P G/Inositol and In Vitro Fertilization with Embryo Transfer. Int J Endocrinol. 2017; 2017:5469409. Published online 2017 Feb 28. doi: 10.1155/2017/5469409. 201