

KOSTRZEWA, Patrycja, KOSTRZEWA, Michał, ANIOŁ, Maciej Sergiusz, PALMERSKA, Anna, PIELA, Andrzej, PIELA, Kinga, PRZYBYŁA, Olga Katarzyna, SZURMA, Anna, SZURMA, Rafał and PACEK, Szymon. Let's open our eyes to the importance of vaginal microbiota. Correlation between dysbiosis and HPV infection: A review. *Journal of Education, Health and Sport*. 2024;71:49399. eISSN 2391-8306.

<https://dx.doi.org/10.12775/JEHS.2024.71.49399>

<https://apcz.umk.pl/JEHS/article/view/49399>

The journal has had 40 points in Minister of Science and Higher Education of Poland parametric evaluation. Annex to the announcement of the Minister of Education and Science of 05.01.2024 No. 32318. Has a Journal's Unique Identifier: 201159. Scientific disciplines assigned: Physical culture sciences (Field of medical and health sciences); Health Sciences (Field of medical and health sciences). Punkty Ministerialne 40 punktów. Załącznik do komunikatu Ministra Nauki i Szkolnictwa Wyższego z dnia 05.01.2024 Lp. 32318. Posiada Unikatowy Identyfikator Czasopisma: 201159. Przypisane dyscypliny naukowe: Nauki o kulturze fizycznej (Dziedzina nauk medycznych i nauk o zdrowiu); Nauki o zdrowiu (Dziedzina nauk medycznych i nauk o zdrowiu). © The Authors 2024;

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The authors declare that there is no conflict of interests regarding the publication of this paper.

Received: 13.03.2024. Revised: 10.05.2024. Accepted: 14.05.2024. Published: 20.05.2024.

Let's open our eyes to the importance of vaginal microbiota. Correlation between dysbiosis and HPV infection: A review

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Abstract

Infections with the human papillomavirus (HPV) are one of the most common sexually transmitted diseases and are the main cause of precancerous changes and cervical cancer development. However, mere infection does not guarantee the development of disease or carcinogenesis. For this to occur, high-risk types of HPV must integrate into the host cell genome, transitioning into a persistent form of infection.

Following the discovery of risk factors for HPV infection and factors predisposing infection to a persistent form, the number of studies investigating the correlation between vaginal dysbiosis and infection has significantly increased. Numerous studies document a significant association between the state of vaginal bacterial flora and HPV infection, leading to the development of plentiful studies on infection pathogenesis, risk factors, prevention, and treatment methods using bacteria. In this paper, based on recently published studies and current state of science knowledge, we will outline the topic of vaginal microbiome, emphasize the importance of bacteria inhabiting the vagina for human health, and identify the problem of changes in vaginal microflora. We would like to contribute to the dissemination of knowledge about the vaginal microbiome and general knowledge about taking care of its health. We will also summarize available treatment methods based on bacterial environment and discuss those methods for which research is currently underway or should be initiated.

Keywords: Microbiota; Lactobacillus; Dysbiosis; Cervicovaginal microbiome; Human Papillomavirus; HPV; Cervical cancer; Bacteriotherapy;

Introduction

For years, the bacterial flora associated with the human body has been a frequently discussed topic in the scientific world. In recent years, research on the bacterial flora present in female reproductive organs, which accounts for about 9% of the total human microflora, has been increasingly conducted [1]. The vaginal microflora protects the reproductive system from potentially pathogenic microorganisms such as bacteria, fungi, and viruses, and generally from sexually transmitted diseases [2]. But what about diseases that are the result of changes in vaginal microenvironmental conditions?

Purpose

Below, we will outline the topic of vaginal dysbiosis, HPV infection, and their mutually non-trivial influence on each other, as well as present new perspectives on treatment utilizing our knowledge of vaginal bacterial flora.

Vaginal Microflora

Vaginal Microflora is an environment that changes over the course of a lifetime, e.g., during puberty, menstrual cycle [3,4], pregnancy and postpartum [5], menopause [6], contraceptive use [7], and sexual activity [8-11]. Hygienic practices [12], the number of sexual partners, smoking, intrauterine device use, scented soaps, vaginal infection history, and vaginal irrigation also have a significant impact [13-15]. It has been demonstrated that vaginal irrigation, especially after menstruation, significantly increases the risk of vaginal bacterial infection [16], while discontinuing this practice can reduce it [17]. Diet also influences the composition of vaginal microflora, especially the presence of vitamins A, C, E, and D, as well as beta-carotene, folic acid, and calcium [18].

"Normalcy" of Vaginal Microbiome

On a global scale, we cannot define the concept of normalcy of the vaginal microbiome because women in different countries have unique elements in their microbiomes in terms of bacterial species proportion, as confirmed by studies in Canada [19], China [20], Europe [21], and Africa [22]. Ravel et al. [23], examining vaginal samples taken from 396 ethnically

diverse women of reproductive age, demonstrated that the vaginal microbiological profile could be classified into five "community state types" (CST) [24-26]. CST I, II, III, and V are characterized by low species diversity with a dominance of *Lactobacillus crispatus*, *L. gasseri*, *L. iners*, and *L. jensenii*, respectively, while CST IV characterizes a heterogeneous group, usually lacking *Lactobacillus* spp. and enriched with anaerobic species often associated with bacterial vaginosis, e.g., *Gardnerella*, *Megasphaera*, *Sneathia* and *Prevotella*, but also *Dialister*, *Prevotella*, *Atopobium*, *Gardnerella*, *Megasphaera*, *Peptoniphilus*, *Sneathia*, *Eggerthella*, *Aerococcus*, *Finegoldia*, and *Mobiluncus*. However, sometimes there is a change from CST III to CST IV, which has led to suspicions of differences in providing stability of vaginal microflora by certain bacterial species. Verstraelen et al. [28], based on vaginal microbiome samples from 100 Caucasian women, confirmed that bacterial flora dominated by *L. crispatus* compared to flora dominated by other *Lactobacillus* species is associated with a fivefold decreased risk of vaginal bacterial dysbiosis.

Bacterial Vaginosis

Bacterial vaginosis is caused by a change in vaginal microflora, specifically an increase in facultative and anaerobic bacteria at the expense of *Lactobacillus* species [29-32]. The main "culprits" of this condition are *Gardnerella vaginalis*, *Mycoplasma hominis*, *Mobiluncus* spp., *Bacteroides* spp., *Prevotella* spp., *Peptostreptococcus* spp., *Fusobacterium* spp., *Porphyromonas* spp., *Megasphaera*, and *Sneathia*, corresponding to CST IV [23, 33]. The prevalence rate in 2012 was as high as 29% in the USA [34]. Clinical symptoms include foul-smelling vaginal discharge, burning during urination, and itching in the vaginal and perineal area, although it can also be asymptomatic [35]. There are many risk factors for bacterial vaginosis, including multiple sexual partners, smoking, intrauterine device use, scented soaps, vaginal infection history, vaginal irrigation [13-17], different hygienic practices [12], diet deficiencies (particularly vitamins A, C, E, and D, as well as beta-carotene, folic acid, and calcium) [18].

Bacterial vaginosis is a risk factor for various conditions, such as preterm birth, low birth weight, HIV infection, and increased susceptibility to chlamydia and gonorrhea infections [36-38], and may also contribute to an increased risk of both acquiring and exacerbating HPV infection, which we will further discuss in the later part of the paper.

Human Papillomavirus HPV

Infection with the *Human Papillomavirus* (HPV) is the most common sexually transmitted disease and is the leading cause of precancerous changes and cervical cancer development [39, 40]. It is estimated that over 80% of sexually active women will become infected with or have been infected with this infection at least once in their lifetime, although the vast majority of HPV infections resolve spontaneously [41]. There are over 200 types of HPV, classified into low-risk (6, 11, 40, 42, 43, 44, 54) associated with benign warts, and high-risk (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59) capable of causing epithelial dysplasia and cervical cancer [42-44].

Despite intensive screening efforts and the increasing frequency of individuals vaccinated against HPV, cervical cancer remains the fourth most commonly diagnosed cancer worldwide among women [45]. Factors associated with prolonged HPV infection and progression to cervical cancer include early onset of sexual activity, multiple sexual partners, coexistence of sexually transmitted diseases, lowered immunity, genetic predisposition, oral contraceptive use, and smoking [46-50].

Increasingly, publications confirm that immune status is significantly correlated with HPV infection. Furthermore, scientific studies and research also confirm that the state of vaginal microflora, which plays a role in regulating the female reproductive tract's immune system, also correlates with infection status [51-52]. There is evidence suggesting a possible relationship between vaginal microflora and the development of cervical intraepithelial neoplasia (CIN), suggesting that an abnormal vaginal environment may play a crucial role in the development and progression of CIN [44, 53-55].

Correlation between dysbiosis and HPV infection

The vaginal microbiome dominated by *Lactobacillus* bacteria protects female genital organs from pathogens [56]. Therefore, vaginal flora with low *Lactobacillus* dominance and increased diversity of pathogenic microorganisms, such as *G. vaginalis*, *Atopobium*, *Prevotella*, *Sneathia*, and *Megasphaera*, is commonly observed in bacterial vaginosis or dysbiosis [57-61]. During disruptions in vaginal bacterial flora, damage to the mucous structure, cytoskeleton, mucous membrane, and antimicrobial peptide construction may occur, along with increased cell death and promotion of pro-inflammatory cytokine production [62-66].

Anahtar et al. [67], after examining a group of asymptomatic women in South Africa, discovered that the vaginal bacterial flora of most women was characterized by low levels of *Lactobacillus* and high bacterial diversity, including *Sneathia sanguinigena*, *Sneathia amnii*, *Mobiluncus mulieris*, and *Prevotella amnii*, which were associated with the presence of pro-inflammatory cytokines such as IL-1 α , IL-1 β , and IL-8.

Similar results were obtained by Łaniewski et al. [68] on three-dimensional models of cervical epithelial cells cultured with *Lactobacillus* as well as with *Gardnerella vaginalis*, *Atopobium vaginae*, *Prevotella bivia*, and *Sneathia amnii*. In particular, *Lactobacillus crispatus* increased the protection of the cervical microenvironment, while *Atopobium vaginae* and *Sneathia amnii* induced inducible nitric oxide synthase (iNOS), oxidative stress, and the highest pro-inflammatory cytokines (IL-6, IL-8, TNF- α , etc.). *Gardnerella vaginalis*, *Prevotella bivia*, and *Sneathia amnii* altered the epithelial barrier by reducing the levels of proteins and metabolites, such as polyamines, sialic acid, and mucins. These changes may result in chronic inflammation, a risk factor for cervical carcinogenesis [69].

Additionally, Fan et al. [70] described the relationship between vaginal bacterial dysbiosis and the proliferation and invasion of cervical cancer cells by studying the fucosylation of the mucous membrane epithelium, whose abnormal course favors the development of cervical cancer.

Lee et al. [71], were the first to use Next Generation Sequencing (NGS) to investigate the influence of HPV on the vaginal microbiome composition. They conducted a cross-sectional cohort study involving 912 women, including 26 monozygotic twins. A clear difference in vaginal flora structure was observed between twins. Women infected with HPV had greater bacterial diversity and significantly fewer *Lactobacillus spp.* compared to their uninfected twin. Furthermore, *Sneathia* was identified as a microbiological marker of HPV infection in this study.

Brotman et al. [72], in their cohort study of 32 sexually active premenopausal North American women, demonstrated that women with CST III and IV had the highest likelihood of HPV infection. Additionally, Brotman and colleagues suggested that CST II, dominated by *L. gasseri*, may be associated with the fastest clearance of acute HPV infection. This observation suggests the potential use of *L. gasseri* as a therapeutic species in maintaining cervical health.

Recent cross-sectional studies have been conducted to characterize vaginal microbiota changes in cervical lesions [73]. It was shown that CIN was associated with decreased

Lactobacillus spp. and increased vaginal microflora diversity. Furthermore, a gradual increase in the frequency of CST IV was observed with increasing disease severity. While in healthy control subjects, CST IV occurred at a frequency of 10%, which is consistent with previous studies involving HPV-uninfected individuals [74].

Di Paola et al. [75], aimed to identify CST associated with persistent HPV by collecting samples from the cervix and vagina of 55 HPV-infected women in Italy and followed up with them after 12 months. They found that over 40% of women with persistent HPV infection had CST IV.

It has also been emphasized that *Gardnerella* species may contribute to maintaining HPV status by secreting sialidase enzyme, important in biofilm formation [75, 76]. Donmez et al. [77], confirmed the association between biofilm formation in the vaginal microenvironment and HPV infection. Utilizing these results, Qingqing et al. [78] identified a high abundance of anaerobes, including *Prevotella*, *Sphingomonas*, and *Anaerococcus*, associated with persistent HPV and increased presence of Treg, MDSC, IL-6, and TNF- α in cervical secretions.

Lebeau et al. [79], after an 8-year observation of over 6000 patients for HPV infection and vaginal microflora status, found that persistent HPV infection could lead to the inhibition of significant amounts of antimicrobial peptides, several of which are essential for *Lactobacillus* survival, resulting in a significant decrease in *Lactobacillus* concentration. In other words, the presence of HPV in the immune system leads to a microbial imbalance in the vaginal flora of women.

Treatment

Oral treatment

Considering the frequent correlation between vaginal dysbiosis and HPV infection and the significant role of bacteria in the development of this infection and cervical cancer, bacteriotherapy has become a promising form of treatment. Probiotics are the most commonly used representatives of bacteriotherapy. According to WHO, probiotics are live microorganisms that, when administered in the right amount, exert a beneficial effect on the host's health. [80]

The most common types of probiotics in the human diet are *Lactobacillus*, *Bifidobacterium*, *Lactococcus*, *Streptococcus*, and *Enterococcus*, as well as some strains belonging to *Bacillus* and *Saccharomyces* [81]. Recently, there has been rapid development in the use of probiotics for cervical cancer [82-84]. Researchers believed that probiotics could promote apoptosis of

cancer cells and inhibit their proliferation and metastasis [85]. As a result, probiotics can be used as an additional strengthening or modulating agent during the use of other diagnostic or therapeutic methods.

It has been demonstrated that probiotics can affect the presence of HPV and its development in three ways. The first main way is by promoting an immune response [86]. Modulation of the immune system may occur, for example, through *L. gasseri*, which can influence the anti-inflammatory action of HeLa cells by reducing TNF- α and increasing the level of IL-10 cytokine [87]. The strain *Bifidobacterium adolescentis* SPM1005-A has potential antitumor activity by inhibiting the expression of oncogenes E6 and E7 in SiHa cells [84].

The second way is based on increasing the number of protective strains acting on the vaginal microenvironment, which can prevent and limit HPV infections by releasing inhibitory factors such as lactic acid, bacteriocins, biosurfactants, and aggregating molecules, as well as by competing for space or nutrients with bacteria with pro-inflammatory effects. The acidic environment can protect against the invasion and development of pathogens [88].

The third and direct option is the elimination of viruses by secreting specific metabolites [89].

Vaginal Treatment

Research has been conducted on vaginal suppositories containing *Lactobacillus* to prevent vaginal dysbiosis [90, 91]. In a clinical trial by Tomusiak et al. [91], it was shown that vaginal therapy with *Lactobacillus species* is safe and may increase the number of these bacteria and reduce the pH and Nugent score in women with symptomatic bacterial vaginosis after four visits.

Lactoferrin is also considered, a bacteriocin produced by lactic acid bacteria. Lactoferrin can act antibacterially by producing lactic acid and destroying the bacterial membrane. Combination therapy of lactoferrin with the antitumor drug IFN- α 2b in the treatment of HPV-infected patients has been studied. Positive results were obtained in terms of restoring the normal vaginal bacterial flora and inhibiting inflammatory factors. [93]

Studies are currently underway to evaluate the effectiveness of combining probiotics with chemotherapy. Previous studies have shown that probiotic supplements may reduce the side effects or toxicity of chemotherapy. Cisplatin is a common treatment method for cervical cancer; however, its toxicity also affects undamaged human cells and tissues. Negi et al. [95] decided to combine cisplatin with pessaries containing probiotics (*Lactobacillus rhamnosus*)

in a mouse vaginal model. They found better outcomes with fewer side effects of cisplatin and reduced tumor volume in the treated group.

Probiotics can help not only with the current disease but also with the remotely appearing accompanying symptoms of cervical cancer treatment. Diarrhea is the most common side effect of radiotherapy in the treatment of cervical cancer [96]. Previous studies have suggested that probiotic supplementation may prevent this gastrointestinal problem. Linn et al. [97], examined the effectiveness of *Lactobacillus acidophilus LA-5* and *Bifidobacterium Animalis subsp. lactis BB-12* in 57 patients with cervical cancer with diarrhea after radiotherapy. The study showed a significant reduction in diarrhea symptoms after three weeks of probiotic use in the mild to moderate group and in the severe diarrhea group.

Vaginal Microbiota Transplantation (VMT) is another treatment option being considered. VMT can improve the imbalance of vaginal bacterial flora by transplanting vaginal secretions from a healthy woman with a high abundance of *Lactobacillus* [98]. Lev-Sagie et al. [99] described the use of VMT in the treatment of incurable and recurrent bacterial vaginal infections. Out of five patients who underwent therapy, four achieved remission with improved symptoms and no side effects after 5 to 21 months of transplantation. However, we want to emphasize that the number of participants in the study was very small, so the study should be repeated on a larger group for more reliable result.

Conclusions

As presented in the research cited above, the vaginal microbiome plays a huge role in maintaining human health. As emphasized in one of the first paragraphs, we should not speak of a "normal vaginal microbiota" in a global sense, but this concept is permissible regionally. One can also speak of a "normal bacterial flora," which means non-pathogenic microorganisms predominated by *Lactobacillus spp.*

In several paragraphs, one can notice repeated factors that influence the state of vaginal microbiota, the risk of HPV infection, and the development of infection leading ultimately to cervical cancer. The conclusion from these repetitions can be simply summarize: to take care of women's health, it is necessary to raise awareness about the importance of maintaining a healthy vaginal bacterial flora, limiting risk factors for vaginal dysbiosis, systematically observing one's body, and effectively treating emerging dysbiosis or vaginal infections. HPV infection status is undoubtedly associated with cervical-vaginal dysbiosis. One might wonder whether HPV infection causes dysbiosis or dysbiosis causes HPV infection. To protect the

public from both risk in addition to the aforementioned considerations for maintaining a healthy vaginal bacterial flora, measures to promote HPV vaccination are important.

But what about women who did not have the opportunity to get vaccinated before infection? For them, research on the vaginal microbiome are especially important. In just a few years, we have expanded our knowledge to incredible proportions, and there is still much to strive for. Millions of women are waiting for a cure for HPV infection or a drug to halt the progression of infection, and we hope that research will not cease and that the medical-scientific community will strive to solve the problem of HPV infection and its associated consequences.

Author's contribution

Conceptualization, Kostrzewa P; methodology, Przybyła O, Piela K; check, Kostrzewa P, Palmerska A, and Szurma R; resources, Szurma A and Kostrzewa M; writing - rough preparation, Kostrzewa M and Piela A; writing - review and editing, Kostrzewa P and Aniol M; visualization, Pacek S; supervision, Kostrzewa P; project administration, Kostrzewa P; All authors have read and agreed with the published version of the manuscript.

Funding Statement

The study did not receive any special funding.

Conflict of Interest

The authors declare no conflict of interest.

References

1. Sirota I, Zarek SM, Segars JH (2014) Potential influence of the microbiome on infertility and assisted reproductive technology. *Semin Reprod Med* 32(1):35–42
2. Sobel JD (1999) Is There a protective role for vaginal flora? *Curr Infect Dis Rep* 1(4):379–383
3. Gajer P, Brotman RM, Bai G, Sakamoto J, Schutte UM, Zhong X, Koenig SS, Fu L, Ma ZS, Zhou X, et al. Temporal dynamics of the human vaginal microbiota. *Sci Transl Med.* 2012;4(132):132RA52 [Temporal Dynamics of the Human Vaginal Microbiota | Science Translational Medicine](#)

4. Eschenbach DA, Thwin SS, Patton DL, Hooton TM, Stapleton AE, Agnew K, Winter C, Meier A, Stamm WE. Influence of the normal menstrual cycle on vaginal tissue, discharge, and microflora. *Clin Infect Dis*. 2000;30(6):901–7.
5. MacIntyre DA, Chandiramani M, Lee YS, Kindinger L, Smith A, Angelopoulos N, Lehne B, Arulkumaran S, Brown R, Teoh TG, et al. The vaginal microbiome during pregnancy and the postpartum period in a European population. *Sci Rep*. 2015;5:8988
6. Brotman RM, Shardell MD, Gajer P, Fadrosch D, Chang K, Silver MI, Viscidi RP, Burke AE, Ravel J, Gravitt PE. Association between the vaginal microbiota, menopause status, and signs of vulvovaginal atrophy. *Menopause*. 2014;21(5):450–8.
7. Vodstrcil LA, Hocking JS, Law M, Walker S, Tabrizi SN, Fairley CK, Bradshaw CS. Hormonal contraception is associated with a reduced risk of bacterial vaginosis: a systematic review and meta-analysis. *PLoS One*. 2013;8(9):e73055.
8. Mandar R, Punab M, Borovkova N, Lapp E, Kiiker R, Korrovits P, Metspalu A, Krjutskov K, Nolvak H, Preem JK, et al. Complementary seminovaginal microbiome in couples. *Res Microbiol*. 2015;166(5):440–7.
9. Gajer P, Brotman RM, Bai G, Sakamoto J, Schütte UM, Zhong X et al (2012) Temporal dynamics of the human vaginal microbiota. *Sci Transl Med*. <https://doi.org/10.1126/scitranslmed.3003605>
10. DiGiulio DB, Callahan BJ, McMurdie PJ, Costello EK, Lyell DJ, Robaczewska A et al (2015) Temporal and spatial variation of the human microbiota during pregnancy. *Proc Natl Acad Sci USA* 112(35):11060–11065
11. Huang Y, Merkatz RB, Hillier SL, Roberts K, Blithe DL, SitrukWare R et al (2015) effects of a one year reusable contraceptive vaginal ring on vaginal microflora and the risk of vaginal infection: an open-label prospective evaluation. *PLoS ONE* 10(8):e0134460
12. Water Aid. Menstrual hygiene matters. [<http://www.wateraid.org/what-wedo/our-approach/research-and-publications/view-publication?id=02309d73-8e41-4d04-b2ef-6641f6616a4f>]. Accessed 19 Oct 2016.
13. Holzman C, Leventhal JM, Qiu H, Jones NM, Wang J, Group BS (2001) Factors linked to bacterial vaginosis in nonpregnant women. *Am J Public Health* 91(10):1664–1670
14. Smart S, Singal A, Mindel A (2004) Social and sexual risk factors for bacterial vaginosis. *Sex Transm Infect* 80(1):58–62

15. Georgijević A, Cjukić-Ivancević S, Bujko M (2000) Bacterial vaginosis. Epidemiology and risk factors. *Srp Arh Celok Lek* 128(1–2):29–33
16. Schwebke JR, Desmond RA, Oh MK. Predictors of bacterial vaginosis in adolescent women who douche. *Sex Transm Dis.* 2004;31(7):433–6.
17. Brotman RM, Ghanem KG, Klebanoff MA, Taha TE, Scharfstein DO, Zenilman JM. The effect of vaginal douching cessation on bacterial vaginosis: a pilot study. *Am J Obstet Gynecol.* 2008;198(6):628. e621-627
18. Thoma ME, Klebanoff MA, Rovner AJ, Nansel TR, Neggers Y, Andrews WW et al (2011) Bacterial vaginosis is associated with variation in dietary indices. *J Nutr* 141(9):1698–1704
19. Chaban B, Links MG, Jayaprakash TP, Wagner EC, Bourque DK, Lohn Z et al (2014) Characterization of the vaginal microbiota of healthy Canadian women through the menstrual cycle. *Microbiome* 2:23
20. Xiao BB, Liao QP (2012) Analysis of diversity of vaginal microbiota in healthy Chinese women by using DNA-fingerprinting. *Beijing Da Xue Xue Bao Yi Xue Ban* 44(2):281–287
21. Jespers V, Menten J, Smet H, Poradosú S, Abdellati S, Verhelst R et al (2012) Quantification of bacterial species of the vaginal microbiome in different groups of women, using nucleic acid amplification tests. *BMC Microbiol* 12:83
22. Anukam KC, Osazuwa EO, Ahonkhai I, Reid G (2006) Lactobacillus vaginal microbiota of women attending a reproductive health care service in Benin city Nigeria. *Sex Transm Dis* 33(1):59–62
23. Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SS, McCulle SL et al (2011) Vaginal microbiome of reproductive-age women. *Proc Natl Acad Sci USA* 108(Suppl 1):4680–4687
24. MacIntyre DA, Chandiramani M, Lee YS, Kindinger L, Smith A, Angelopoulos N, Lehne B, Arulkumaran S, Brown R, Teoh TG, et al. The vaginal microbiome during pregnancy and the postpartum period in a European population. *Sci Rep.* 2015;5:8988.
25. Brotman RM, Shardell MD, Gajer P, Fadrosch D, Chang K, Silver MI, Viscidi RP, Burke AE, Ravel J, Gravitt PE. Association between the vaginal microbiota, menopause status, and signs of vulvovaginal atrophy. *Menopause.* 2014;21(5):450–8.
26. Kindinger LM, MacIntyre DA, Lee YS, Marchesi JR, Smith A, McDonald JA, Terzidou V, Cook JR, Lees C, Israfil-Bayli F, et al. Relationship between vaginal

- microbial dysbiosis, inflammation, and pregnancy outcomes in cervical cerclage. *Sci Transl Med.* 2016;8(350):350ra102.
27. Romero R, Hassan SS, Gajer P, Tarca AL, Fadrosh DW, Nikita L, Galuppi M, Lamont RF, Chaemsaitong P, Miranda J, et al. The composition and stability of the vaginal microbiota of normal pregnant women
 28. Verstraelen H, Verhelst R, Claeys G, De Backer E, Temmerman M, Vaneechoutte M (2009) Longitudinal analysis of the vaginal microflora in pregnancy suggests that *L. crispatus* promotes the stability of the normal vaginal microflora and that *L. gasseri* and/ or *L. iners* are more conducive to the occurrence of abnormal vaginal microflora. *BMC Microbiol.* <https://doi.org/10.1186/1471-2180-9-116>
 29. Brotman, R.M.; Shardell, M.D.; Gajer, P.; Tracy, J.K.; Zenilman, J.M.; Ravel, J.; Gravitt, P.E. Interplay Between the Temporal Dynamics of the Vaginal Microbiota and Human Papillomavirus Detection. *J. Infect. Dis.* 2014, 210, 1723–1733. <http://doi.org/10.1093/infdis/jiu330>
 30. Duchi, S.; Onofrillo, C.; O’Connell, C.D.; Blanchard, R.; Augustine, C.; Quigley, A.F.; Kapsa, R.M.; Pivonka, P.; Wallace, G.; Di Bella, C.; et al. Characterization of cervico-vaginal microbiota in women developing persistent high-risk Human Papillo-mavirus infection. *Sci. Rep.* 2017, 7, 5837. <http://doi.org/10.1038/s41598-017-05699-x>
 31. Adebamowo, S.N.; Ma, B.; Zella, D.; Famooto, A.; Ravel, J.; Adebamowo, C. *Mycoplasma hominis* and *mycoplasma genitalium* in the vaginal microbiota and persistent high-risk human papillomavirus infection. *Front. Public Health* 2017, 5, 1–10. <http://doi.org/10.3389/fpubh.2017.00140>
 32. Arokiyaraj, S.; Seo, S.S.; Kwon, M.; Lee, J.K.; Kim, M.K. Association of cervical microbial community with persistence, clearance and negativity of Human Papillomavirus in Korean women: A longitudinal study. *Sci. Rep.* 2018, 8, 15479. <http://doi.org/10.1038/s41598-018-33750-y>
 33. Drell T, Lillsaar T, Tummeleht L, Simm J, Aaspõllu A, Väin E et al (2013) Characterization of the vaginal micro-and mycobiome in asymptomatic reproductive-age Estonian women. *PLoS ONE* 8(1):e54379
 34. Kenyon C, Colebunders R, Crucitti T. The global epidemiology of bacterial vaginosis: a systematic review. *Am J Obstet Gynecol.* 2013;209(6):505–23

35. van de Wijgert JH, Borgdorf H, Verhelst R, Crucitti T, Francis S, Verstraelen H et al (2014) The vaginal microbiota: what have we learned after a decade of molecular characterization? *PLoS ONE* 9(8):e105998
36. Hillier SL, Nugent RP, Eschenbach DA, Krohn MA, Gibbs RS, Martin DH et al (1995) Association between bacterial vaginosis and preterm delivery of a low-birth-weight infant. The vaginal infections and prematurity study group. *N Engl J Med* 333(26):1737–1742
37. Petrova MI, van den Broek M, Balzarini J, Vanderleyden J, Lebeer S (2013) Vaginal microbiota and its role in HIV transmission and infection. *FEMS Microbiol Rev* 37(5):762–792
38. Barbone F, Austin H, Louv WC, Alexander WJ (1990) A follow-up study of methods of contraception, sexual activity, and rates of trichomoniasis, candidiasis, and bacterial vaginosis. *Am J Obstet Gynecol* 163(2):510–514
39. Koshiol, J.; Lindsay, L.; Pimenta, J.M.; Poole, C.; Jenkins, D.; Smith, J.S. Persistent human papillomavirus infection and cervical neoplasia: A systematic review and meta-analysis. *Am. J. Epidemiol.* 2008, 168, 123–137. <http://doi.org/10.1093/aje/kwn036>
40. Vedham, V.; Verma, M.; Mahabir, S. Early-life exposures to infectious agents and later cancer development. *Cancer Med.* 2015, 4, 1908–1922 <http://doi.org/10.1002/cam4.538>
41. Plummer M, Schiffman M, Castle PE, Maucort-Boulch D, Wheeler CM. A 2- year prospective study of human papillomavirus persistence among women with a cytological diagnosis of atypical squamous cells of undetermined significance or low-grade squamous intraepithelial lesion. *J Infect Dis.* 2007; 195(11):1582–9
42. Bzhalava, D.; Eklund, C.; Dillner, J. International Standardization and Classification of Human Papillomavirus Types. *Virology* 2015, 476, 341–344 <https://doi.org/10.1016/j.virol.2014.12.028>
43. McLaughlin-Daurbin, M.E.; Münger, K. Oncogenic Activities of Human Papillomaviruses Margaret. *Virus Res.* 2009, 143, 195–208. <https://doi.org/10.1016/j.virusres.2009.06.008>
44. Castellsagué, X. Natural history and epidemiology of HPV infection and cervical cancer. *Gynecol. Oncol.* 2008, 110 (Suppl. S2), S4–S7 <http://doi.org/10.1016/j.ygyno.2008.07.045>

45. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 2018, 68, 394–424. <http://doi.org/10.3322/caac.21492>
46. Trottier, H.; Franco, E.L. The epidemiology of genital human papillomavirus infection. *Vaccine* 2006, 24 (Suppl. S1), S4–S15. <http://doi.org/10.1016/j.vaccine.2005.09.054>
47. Sreedevi, A.; Javed, R.; Dinesh, A. Epidemiology of cervical cancer with special focus on India. *Int. J. Womens Health* 2015, 7, 405–414.
48. Erickson, B.K.; Alvarez, R.D.; Huh, W.K. Human papillomavirus: What every provider should know. *Am. J. Obstet. Gynecol.* 2014, 208, 169–175. <http://doi.org/10.1016/j.ajog.2012.09.007>
49. Hariri, S.; Unger, E.; Sternberg, M.; Dunne, E.F.; Swan, D.; Patel, S.; Markowitz, L.E. Prevalence of Genital Human Papillomavirus Among Females in the United States, the National Health and Nutrition Examination Survey, 2003. *J. Infect. Dis.* 2011, 204, 566–573. <http://doi.org/10.1093/infdis/jir341>
50. Caruso, S.; Bruno, M.T.; Cianci, S.; Di Pasqua, S.; Minona, P.; Cianci, A. Sexual Behavior of Women With Diagnosed HPV. *J. Sex Marital Ther.* 2019, 45, 569–573. <http://doi.org/10.1080/0092623X.2019.1586019>
51. Audirac-Chalifour, A.; Torres-Poveda, K.; Bahena-Román, M.; Téllez-Sosa, J.; Martínez-Barnette, J.; Cortina-Ceballos, B.; López-Estrada, G.; Delgado-Romero, K.; Burguete-García, A.I.; Cantú, D.; et al. Cervical Microbiome and Cytokine Profile at Various Stages of Cervical Cancer: A Pilot Study. *PLoS ONE* 2016, 11, e0153274. <http://doi.org/10.1371/journal.pone.0153274>
52. Pino, A.; Rapisarda, A.M.C.; Vitale, S.G.; Cianci, S.; Caggia, C.; Randazzo, C.L.; Cianci, A. A clinical pilot study on the effect of the probiotic *Lactobacillus rhamnosus* TOM 22.8 strain in women with vaginal dysbiosis. *Sci. Rep.* 2021, 11, 2592. <http://doi.org/10.1038/s41598-021-81931-z>
53. Schiffman, M.; Castle, P.E. The Promise of Global Cervical-Cancer Prevention. *N. Engl. J. Med.* 2005, 353, 2101–2104. <http://doi.org/10.1056/NEJMp058171>
54. Joura, E.A.; Ault, K.A.; Bosch, F.X.; Brown, D.; Cuzick, J.; Ferris, D.; Garland, S.M.; Giuliano, A.R.; Hernandez-Avila, M.; Huh, W.; et al. Attribution of 12 High-Risk Human Papillomavirus Genotypes to Infection and Cervical Disease. *Cancer*

- Epidemiol. Biomarkers Prev. 2014, 23, 1997–2008. <http://doi.org/10.1158/1055-9965.EPI-14-0410>
55. Tommasino, M. The human papillomavirus family and its role in carcinogenesis. *Semin. Cancer Biol.* 2014, 26, 13–21. <http://doi.org/10.1016/j.semcancer.2013.11.002>
56. Chee, W.J.Y.; Chew, S.Y.; Than, L.T.L. Vaginal Microbiota and the Potential of Lactobacillus Derivatives in Maintaining Vaginal Health. *Microb. Cell Fact.* 2020, 19, 203. <https://doi.org/10.1186/s12934-020-01464-4>
57. Kaelin, E.A.; Skidmore, P.T.; Łaniewski, P.; Holland, L.A.; Chase, D.M.; Herbst-Kralovetz, M.M.; Lim, E.S. Cervicovaginal DNA Virome Alterations Are Associated with Genital Inflammation and Microbiota Composition. *mSystems* 2022, 7, e00064-22. <https://doi.org/10.1128/msystems.00064-22>
58. Sodhani, P.; Gupta, S.; Gupta, R.; Mehrotra, R. Bacterial Vaginosis and Cervical Intraepithelial Neoplasia: Is There an Association or Is Co-Existence Incidental? *Asian Pac. J. Cancer Prev.* 2017, 18, 1289–1292. <https://doi.org/10.22034/APJCP.2017.18.5.1289>
59. Lin, S.; Zhang, B.; Lin, Y.; Lin, Y.; Zuo, X. Dysbiosis of Cervical and Vaginal Microbiota Associated with Cervical Intraepithelial Neoplasia. *Front. Cell. Infect. Microbiol.* 2022, 12, 767693. <https://doi.org/10.3389/fcimb.2022.767693>
60. Xu, X.; Zhang, Y.; Yu, L.; Shi, X.; Min, M.; Xiong, L.; Pan, J.; Liu, P.; Wu, G.; Gao, G. A Cross-Sectional Analysis about Bacterial Vaginosis, High-Risk Human Papillomavirus Infection, and Cervical Intraepithelial Neoplasia in Chinese Women. *Sci. Rep.* 2022, 12, 6609. <https://doi.org/10.1038/s41598-022-10532-1>
61. Onderdonk, A.B.; Delaney, M.L.; Fichorova, R.N. The Human Microbiome during Bacterial Vaginosis. *Clin. Microbiol. Rev.* 2016, 29, 223–238. <https://doi.org/10.1128/CMR.00075-15>
62. Sun, N.; Ding, H.; Yu, H.; Ji, Y.; Xifang, X.; Pang, W.; Wang, X.; Zhang, Q.; Li, W. Comprehensive Characterization of Microbial Community in the Female Genital Tract of Reproductive-Aged Women in China. *Front. Cell. Infect. Microbiol.* 2021, 11, 649067. <https://doi.org/10.3389/fcimb.2021.649067>
63. Torcia, M.G. Interplay among Vaginal Microbiome, Immune Response and Sexually Transmitted Viral Infections. *Int. J. Mol. Sci.* 2019, 20, 266. <https://doi.org/10.3390/ijms20020266>

64. Łaniewski, P.; Barnes, D.; Goulder, A.; Cui, H.; Roe, D.J.; Chase, D.M.; Herbst-Kralovetz, M.M. Linking Cervicovaginal Immune Signatures, HPV and Microbiota Composition in Cervical Carcinogenesis in Non-Hispanic and Hispanic Women. *Sci. Rep.* 2018, 8, 7593. <https://doi.org/10.1038/s41598-018-25879-7>
65. Mitchell, C.; Marrazzo, J. Bacterial Vaginosis and the Cervicovaginal Immune Response. *Am. J. Reprod. Immunol.* 2014, 71, 555–563. <https://doi.org/10.1111/aji.12264>
66. Borgdorff, H.; Gautam, R.; Armstrong, S.D.; Xia, D.; Ndayisaba, G.F.; Van Teijlingen, N.H.; Geijtenbeek, T.B.H.; Wastling, J.M.; Van De Wiggert, J.H.H.M. Cervicovaginal Microbiome Dysbiosis Is Associated with Proteome Changes Related to Alterations of the Cervicovaginal Mucosal Barrier. *Mucosal Immunol.* 2016, 9, 621–633. <https://doi.org/10.1038/mi.2015.86>
67. Anahtar, M.N.; Byrne, E.H.; Doherty, K.E.; Bowman, B.A.; Yamamoto, H.S.; Soumillon, M.; Padavattan, N.; Ismail, N.; Moodley, A.; Sabatini, M.E.; et al. Cervicovaginal Bacteria Are a Major Modulator of Host Inflammatory Responses in the Female Genital Tract. *Immunity* 2015, 42, 965–976. <https://doi.org/10.1016/j.immuni.2015.04.019>
68. Łaniewski, P.; Herbst-Kralovetz, M.M. Bacterial Vaginosis and Health-Associated Bacteria Modulate the Immunometabolic Landscape in 3D Model of Human Cervix. *NPJ Biofilms Microbiomes* 2021, 7, 88. <https://doi.org/10.1038/s41522-021-00259-8>
69. Łaniewski, P.; Barnes, D.; Goulder, A.; Cui, H.; Roe, D.J.; Chase, D.M.; Herbst-Kralovetz, M.M. Linking Cervicovaginal Immune Signatures, HPV and Microbiota Composition in Cervical Carcinogenesis in Non-Hispanic and Hispanic Women. *Sci. Rep.* 2018, 8, 7593. <https://doi.org/10.1038/s41598-018-25879-7>
70. Fan, Q.; Wu, Y.; Li, M.; An, F.; Yao, L.; Wang, M.; Wang, X.; Yuan, J.; Jiang, K.; Li, W.; et al. *Lactobacillus* Spp. Create a Protective Micro-Ecological Environment through Regulating the Core Fucosylation of Vaginal Epithelial Cells against Cervical Cancer. *Cell Death Dis.* 2021, 12, 1094. <https://doi.org/10.1038/s41419-021-04388-y>
71. Lee JE, Lee S, Lee H, Song YM, Lee K, Han MJ, Sung J, Ko G. Association of the vaginal microbiota with human papillomavirus infection in a Korean twin cohort. *PLoS One.* 2013;8(5):e63514.

72. Brotman RM, Shardell MD, Gajer P, Tracy JK, Zenilman JM, Ravel J, Gravitt PE. Interplay between the temporal dynamics of the vaginal microbiota and human papillomavirus detection. *J Infect Dis.* 2014;210(11):1723–33
73. Brotman RM, Shardell MD, Gajer P, Tracy JK, Zenilman JM, Ravel J, Gravitt PE. Interplay between the temporal dynamics of the vaginal microbiota and human papillomavirus detection. *J Infect Dis.* 2014;210(11):1723–33
74. Ravel J, Gajer P, Abdo Z, Schneider GM, Koenig SS, McCulle SL, Karlebach S, Gorle R, Russell J, Tacket CO, et al. Vaginal microbiome of reproductive-age women. *Proc Natl Acad Sci U S A.* 2011;108 Suppl 1:4680–7.
75. Di Paola, M.; Sani, C.; Clemente, A.M.; Iossa, A.; Perissi, E.; Castronovo, G.; Tanturli, M.; Rivero, D.; Cozzolino, F.; Cavalieri, D.; et al. Characterization of Cervico-Vaginal Microbiota in Women Developing Persistent High-Risk Human Papillomavirus Infection. *Sci. Rep.* 2017, 7, 10200. <https://doi.org/10.1038/s41598-017-09842-6>
76. Lin, W.; Zhang, Q.; Chen, Y.; Dong, B.; Xue, H.; Lei, H.; Lu, Y.; Wei, X.; Sun, P. Changes of the Vaginal Microbiota in HPV Infection and Cervical Intraepithelial Neoplasia: A Cross-Sectional Analysis. *Sci. Rep.* 2022, 12, 2812. <https://doi.org/10.1038/s41598-022-06731-5>
77. Donmez, H.G.; Sahal, G.; Akgor, U.; Cagan, M.; Ozgul, N.; Beksac, M.S. The Relationship between the Presence of HPV Infection and Biofilm Formation in Cervicovaginal Smears. *Infection* 2020, 48, 735–740. <https://doi.org/10.1007/s15010-020-01478-5>
78. Qingqing, B.; Jie, Z.; Songben, Q.; Juan, C.; Lei, Z.; Mu, X. Cervicovaginal Microbiota Dysbiosis Correlates with HPV Persistent Infection. *Microb. Pathog.* 2021, 152, 104617. <https://doi.org/10.1016/j.micpath.2020.104617>
79. Lebeau, A.; Bruyere, D.; Roncarati, P.; Peixoto, P.; Hervouet, E.; Cobraiville, G.; Taminiou, B.; Masson, M.; Gallego, C.; Mazzucchelli, G.; et al. HPV Infection Alters Vaginal Microbiome through Down-Regulating Host Mucosal Innate Peptides Used by Lactobacilli as Amino Acid Sources. *Nat. Commun.* 2022, 13, 1076. <https://doi.org/10.1038/s41467-022-28724-8>
80. Report FAO/WHO. 2001. Report of a Joint FAO/WHO Expert Consultation on Evaluation of Health and Nutritional Properties of Probiotics in Food including Powder Milk with Live Lactic Acid Bacteria [Internet]. p. 1–29; [cited 2017

November 15]. Available from: <http://www.fao.org/tempref/docrep/fao/meeting/009/y6398e.pdf>

81. Markowiak, P.; Slizewska, K. Effects of Probiotics, Prebiotics, and Synbiotics on Human Health. *Nutrients* 2017, 9, 1021. <https://doi.org/10.3390/nu9091021>
82. Pourmollaei, S.; Barzegari, A.; Farshbaf-Khalili, A.; Nouri, M.; Fattahi, A.; Shahnazi, M.; Dittrich, R. Anticancer Effect of Bacteria on Cervical Cancer: Molecular Aspects and Therapeutic Implications. *Life Sci.* 2020, 246, 117413. <https://doi.org/10.1016/j.lfs.2020.117413>
83. Verhoeven, V.; Renard, N.; Makar, A.; Van Royen, P.; Bogers, J.P.; Lardon, F.; Peeters, M.; Baay, M. Probiotics Enhance the Clearance of Human Papillomavirus-Related Cervical Lesions: A Prospective Controlled Pilot Study. *Eur. J. Cancer Prev.* 2013, 22, 46–51. <https://doi.org/10.1097/CEJ.0b013e328355ed23>
84. Cha, M.; Lee, D.; An, H.; Lee, S.; Shin, S.; Kwon, J.; Kim, K.; Ha, N. Antiviral Activity of Bifidobacterium Adolescentis SPM1005-A on Human Papillomavirus Type 16. *BMC Med.* 2012, 10, 72. <https://doi.org/10.1186/1741-7015-10-72>
85. Jahanshahi, M.; Maleki Dana, P.; Badehnoosh, B.; Asemi, Z.; Hallajzadeh, J.; Mansournia, M.A.; Yousefi, B.; Yousefi, B.; Moazzami, B.; Chaichian, S. Anti-Tumor Activities of Probiotics in Cervical Cancer. *J. Ovarian Res.* 2020, 13, 68. <https://doi.org/10.1186/s13048-020-00668-x>
86. Azad, M.A.K.; Sarker, M.; Wan, D. Immunomodulatory Effects of Probiotics on Cytokine Profiles. *Biomed Res. Int.* 2018, 2018, 8063647. <https://doi.org/10.1155/2018/8063647>
87. Sungur, T.; Aslim, B.; Karaaslan, C.; Aktas, B. Impact of Exopolysaccharides (EPSs) of Lactobacillus Gasseri Strains Isolated from Human Vagina on Cervical Tumor Cells (HeLa). *Anaerobe* 2017, 47, 137–144. <https://doi.org/10.1016/j.anaerobe.2017.05.013>
88. Pourmollaei, S.; Barzegari, A.; Farshbaf-Khalili, A.; Nouri, M.; Fattahi, A.; Shahnazi, M.; Dittrich, R. Anticancer Effect of Bacteria on Cervical Cancer: Molecular Aspects and Therapeutic Implications. *Life Sci.* 2020, 246, 117413. <https://doi.org/10.1016/j.lfs.2020.117413>
89. Van Baarlen, P.; Troost, F.; Van Der Meer, C.; Hooiveld, G.; Boekschoten, M.; Brummer, R.J.M.; Kleerebezem, M. Human Mucosal in Vivo Transcriptome Responses to Three Lactobacilli Indicate How Probiotics May Modulate Human

- Cellular Pathways. *Proc. Natl. Acad. Sci. USA* 2011, 108, 4562–4569. <https://doi.org/10.1073/pnas.1000079107>
90. van de Wijgert, J.H.H.M.; Verwijs, M.C. Lactobacilli-Containing Vaginal Probiotics to Cure or Prevent Bacterial or Fungal Vaginal Dysbiosis: A Systematic Review and Recommendations for Future Trial Designs. *BJOG Int. J. Obstet. Gynaecol.* 2020, 127, 287–299. <https://doi.org/10.1111/1471-0528.15870>
91. 111. Tomusiak, A.; Strus, M.; Heczko, P.B.; Adamski, P.; Stefański, G.; Mikołajczyk-Cichońska, A.; Suda-Szczurek, M. Efficacy and Safety of a Vaginal Medicinal Product Containing Three Strains of Probiotic Bacteria: A Multicenter, Randomized, Double-Blind, and Placebo-Controlled Trial. *Drug Des. Dev. Ther.* 2015, 9, 5345–5354. <https://doi.org/10.2147/DDDT.S89214>
92. Zhu, Y.; Zhang, S. Antibacterial Activity and Mechanism of Lacidophilin From *Lactobacillus Pentosus* Against *Staphylococcus Aureus* and *Escherichia Coli*. *Front. Microbiol.* 2020, 11, 582349. <https://doi.org/10.3389/fmicb.2020.582349>
93. Sun, Y.; Xu, J.; Zhou, H.; You, L.; Zhu, Y. Influence of Lacidophilin Vaginal Capsules plus Rh-IFN- α 2b on Efficacy, Vaginal Microecology, and Safety of Patients with HPV Infection. *Evid.-Based Complement. Altern. Med.* 2022, 2022, 3632053. <https://doi.org/10.1155/2022/3632053>
94. Rodriguez-Arrastia, M.; Martinez-Ortigosa, A.; Rueda-Ruzafa, L.; Ayora, A.F.; Ropero-Padilla, C. Probiotic Supplements on Oncology Patients' Treatment-Related Side Effects: A Systematic Review of Randomized Controlled Trials. *Int. J. Environ. Res. Public Health* 2021, 18, 4265. <https://doi.org/10.3390/ijerph18084265>
95. Negi, D.; Singh, A.; Joshi, N.; Mishra, N. Cisplatin and Probiotic Biomass Loaded Pessaries for the Management of Cervical Cancer. *Anticancer Agents Med. Chem.* 2019, 20, 589–598. <https://doi.org/10.2174/1871520619666191211110640>
96. Andreyev, J. Gastrointestinal Complications of Pelvic Radiotherapy: Are They of Any Importance? *Gut* 2005, 54, 1051–1054. <https://doi.org/10.1136/gut.2004.062596>
97. 119. Linn, Y.H.; Thu, K.K.; Win, N.H.H. Effect of Probiotics for the Prevention of Acute Radiation-Induced Diarrhoea Among Cervical Cancer Patients: A Randomized Double-Blind Placebo-Controlled Study. *Probiotics Antimicrob. Proteins* 2019, 11, 638–647. <https://doi.org/10.1007/s12602-018-9408-9>
98. 120. Yockey, L.J.; Hussain, F.A.; Bergerat, A.; Reissis, A.; Worrall, D.; Xu, J.; Gomez, I.; Bloom, S.M.; Mafunda, N.A.; Kelly, J.; et al. Screening and Characterization of

Vaginal Fluid Donations for Vaginal Microbiota Transplantation. *Sci. Rep.* 2022, 12, 17948. <https://doi.org/10.1038/s41598-022-22873-y>

99. 121. Lev-Sagie, A.; Goldman-Wohl, D.; Cohen, Y.; Dori-Bachash, M.; Leshem, A.; Mor, U.; Strahilevitz, J.; Moses, A.E.; Shapiro, H.; Yagel, S.; et al. Vaginal Microbiome Transplantation in Women with Intractable Bacterial Vaginosis. *Nat. Med.* 2019, 25, 1500–1504. <https://doi.org/10.1038/s41591-019-0600-6>