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THE ROLE OF IL-18 IN THE DEVELOPMENT OF DIABETES **MELLITUS**

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Abstract

Background. Currently, diabetes mellitus is the most common of all endocrine diseases with increasing tendency. The purpose of our research was to study the dynamics of serum levels of proinflammatory cytokine-IL-1β in streptozotocin-induced diabetes. Materials and methods. The experiments were performed on 88 white male Wistar rats weighing 170-210 g. Animals were divided into three groups: 1 - intact; 2 - control; 3 experimental with a model of diabetes mellitus, which was reproduced by intraperitoneal injection of streptozotocin by "Sigma" company (USA), diluted in 0.1 M citrate buffer with a pH of 4.5, at a rate of 60 mg/kg body weight. The control group of animals received an intraperitoneal injection with an equivalent dose of 0.1 M citrate buffer solution with a pH of 4.5.

All studies were performed under thiopental-sodium anesthesia at a rate of 60 mg/kg body weight. Serum levels of IL-1β were determined by enzyme-linked immunosorbent assay (ELISA) kit (Elabscience, USA) according to the manufacturer's instructions 14, 28, 42 and 70 days after streptozotocin injection. The STATISTICA 10 program was used for statistical processing of the obtained results. Results. Conducted biochemical analysis showed that in animals with streptozotocin-induced diabetes there was an increase in the content of proinflammatory cytokine IL-1β compared with similar indicators in the control group of animals at

all stages of the experiment: after 14 days by 22.5%, after 28 days by 40.2%, after 42 days by 72.8% and after 70 days by 107.2%. **Conclusion.** The obtained results suggest that pro-inflammatory cytokine interleukin-1 β plays one of the leading roles in the pathogenesis of streptozotocin-induced diabetes, as indicated by a significant increase in serum of this cytokine at all stages of the experiment.

Key words: streptozotocin-induced diabetes, interleukin-1β.

INTRODUCTION

Nowadays, diabetes mellitus (DM) is the most common of all endocrine diseases with increasing tendency [1, 2, 17, 28, 29]. According to the International Diabetes Federation (IDF), the number of people with diabetes will increase to 629 million by 2045 worldwide [19, 25, 30]. In recent years, the prevalence of diabetes has increased due to a large number of patients with type 2 diabetes [6, 10, 15, 16].

Lately, increasing attention has been paid to establishing the pathogenetic role of cytokines in various diseases, including DM [3, 18, 20, 31]. It is known that cytokines are regulators of intercellular and intersystemic interactions, and ensure the coherence of the endocrine, immune, and nervous systems both under normal conditions and in response to pathogenetic factors. To date, it has been established that among many cytokines, a special role in the development of diabetes belongs to proinflammatory interleukin-1 β (IL-1 β) [8, 21, 24, 32, 34].

Major sources of IL-1 β include tissue macrophages, blood monocytes, and dendritic cells [7, 14]. According to the literature data, IL-1 β is a key proinflammatory cytokine that inhibits insulin secretion and stimulates the expression of a gene encoding inducible nitric oxide synthase. The latter leads to the synthesis of NO and the death of β -cells of the pancreas due to necrosis or apoptosis in experimental animals with spontaneous autoimmune diabetes [11]. The cytotoxic effect of IL-1 β on β -cells of the islets of Langerhans is indicated by many other authors [4, 5, 22, 26, 33].

However, the role of proinflammatory cytokines in diabetes remains unclear.

The aim of our research was to study the dynamics of serum levels of proinflammatory cytokine-IL-1β in streptozotocin-induced diabetes.

MATERIALS AND METHODS

The experiments were performed on 88 white male Wistar rats weighing 170-210 g, which were kept on a standard diet with free access to water. Animals were divided into three groups: 1 - intact (n = 10); 2 - control (n = 40); 3 - experimental (n = 38) with a model of diabetes mellitus, which was reproduced by intraperitoneal injection of streptozotocin by "Sigma" company (USA), diluted in 0.1 M citrate buffer with a pH of 4.5, at a rate of 60 mg/kg body weight. The control group of animals received an intraperitoneal injection with an equivalent dose of 0.1 M citrate buffer solution with a pH of 4.5.

Animal husbandry and research were conducted in accordance with the provisions of the "European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes" (Strasbourg, 1986), the Law of Ukraine on the "Protection of Animals from Cruelty" (2006) and the "General Ethical Principles of Experiments on Animals" approved by the Fifth National Congress on Bioethics (Kyiv, 2013). All studies were performed under thiopental-sodium anesthesia at a rate of 60 mg/kg body weight.

Serum IL-1 β levels were determined by enzyme-linked immunosorbent assay (ELISA) kit (Elabscience, USA) according to the manufacturer's instructions 14, 28, 42 and 70 days after streptozotocin injection.

The STATISTICA 10 program was used for statistical processing of the obtained results. Using the possibilities of descriptive statistics, all the quantitative data obtained in the study were first checked for the type of their distribution by the Shapiro-Wilk test. Since the vast majority of these data were

consistent with Gauss's normal law, the arithmetic mean \pm standard error of mean (M \pm m) was chosen to describe the central trend, and a parametric t-test (Student's test) was chosen to assess the reliability of differences in the results obtained in the comparison groups (experimental and control) and to test the null hypothesis. To assess the reliability of data changes in the dynamics (14, 28, 42, 70 days) within each of the comparison groups we used a non-parametric method for three or more comparison groups — Friedman's test and Kendall's coefficient of concordance (Friedman ANOVA and Kenall Coef. of Concordance).

RESULTS AND DISCUSSION

Conducted studies showed that in animals with streptozotocin-induced diabetes there was an increase in the content of proinflammatory cytokine IL-1 β compared with the similar indicators of the control group of animals at all stages of the experiment (Table 1 and Figure 1).

Table 1 The content of IL-1 β (pg / mL) in the serum of white rats in experimental diabetes mellitus.

Group	14 days		28 days		42 days		70 days		10.
	M	±m	M	±m	M	±m	M	±m	p_2
Experimen	115,9	0,63	130,5	0,58	164,5	1 40	194,4	1.05	<0,00
t	*	0,03	*	0,38	*	1,40	*	1,05	1
Control	94,6	0,68	93,1	0,63	94,8	0,63	93,6	0,64	>0,05
p_1	<0,001		<0,001		<0,001		<0,001		X
Intact	$94,8\pm0,70$								

Notes: p_1 – the reliability of the difference between the data of the experimental and control groups;

p₂ – reliability of data within the group in the dynamics;

^{*} – reliability of the data difference compared to the intact group.

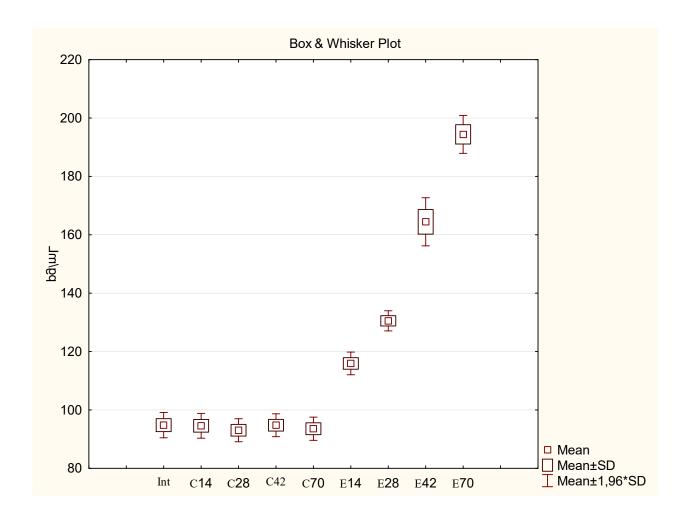


Fig. 1. Dynamics of IL-1 β content (pg/mL) in the serum of white rats in experimental diabetes mellitus.

Notes: groups of animals: Int – intact; C – control; E – experimental. 14, 28, 42, 70 – days of the experiment

It was found that 14 days after the start of the experiment, the level of IL- 1β in the serum significantly exceeded the control group of animals by 22.5% (p<0.001). With increasing study duration (28 days) there was an increase in serum proinflammatory cytokine IL- 1β by 40.2% (p<0.001) compared with the control group of animals. 42 days after the experiment, a further increase in the level of IL- 1β in the serum was determined. It was found that the value of IL- 1β in the serum exceeded that of the control group of animals by 72.8% (p<0.001).

The study of the serum content of IL-1 β after 70 days in the conditions of simulated diabetes showed a further increase in this indicator. In particular, it was found that the serum level of IL-1 β at that period of the study exceeded similar indicator of the control group of animals by 107.2% (p<0.001).

The results of the conducted studies showed that in the conditions of simulated diabetes the serum level of proinflammatory cytokine IL-1 β increased by 22.5%, 40.2%, 72.8% and 107.2%, respectively, on 14, 28, 42 and 70 days after the start of experiment. It was found that the maximum increase in IL-1 β was observed 70 days after modelling of diabetes relative to the control group of animals (p<0.001) and differed significantly from the value of IL-1 β in previous observation periods.

The results of our study are consistent with the data of other researchers, who also indicated a significant increase in serum proinflammatory cytokine IL-1 β in diabetes [9, 20, 23, 27]. The production of significant amounts of IL-1 β can lead to large the number of pullo-proliferative cells and their changes may be proportional to the degree of damage. IL-1 β induces a wide range of biological effects on both systemic level and at the site of localization of the inflammatory response. Activating the cells of innate and acquired parts of the immune system, IL-1 β promotes vessel expansion, production of acute phase proteins, and antibody synthesis [12, 13]. According to the literature data, proinflammatory cytokine IL-1 β induces the migration of proinflammatory cells to the islets of Langerhans of the pancreas, mediates cytokine-induced apoptosis of β -cells, has cytotoxic properties on β -cells and thus promotes development of diabetes mellitus [4, 22].

CONCLUSION

The obtained results suggest that pro-inflammatory interleukin- 1β plays one of the leading roles in the pathogenesis of streptozotocin-induced diabetes,

as indicated by a significant increase in serum of this cytokine at all stages of the experiment.

REFERENCES

- 1. Akasaka H, Ohnishi H, Narita Y, Kameda M, Miki T, Takahashi H, et al. The Serum Level of KL-6 Is Associated with the Risk of Insulin Resistance and New-onset Diabetes Mellitus: The Tanno-Sobetsu Study. Intern Med. 2017; 56: 3009-3018.
- 2. Atkinson MA, Eisenbarth GS, Michels AW. Type 1 diabetes. Lancet. 2014; 383(9911):69-82.
- 3. Bandas IA, Krynytska IYa, Kulitska MI, Kuzmak IP, Korda MM. Changes of blood serum cytokine profile in rats in response to combined introduction of silicon dioxide nanoparticles and chemical toxicant lead acetate [in Ukrainian]. Bulletin of Scientific Research. 2017;3:114-118.
- 4. Berchtold LA, Prause M, Sterling J, Mandrup-Poulsen T. Cytokines and reatic beta-Cell Apoptosis. Adv Clin Chem. 2016;75:99-158.
- 5. Bulboaca AE, Boarescu PM, Bolboaca SD, Blidaru M, Festila D, Dogaru G, et al. Comparative Effect Of Curcumin Versus Liposomal Curcumin On Systemic Pro-Inflammatory Cytokines Profile, MCP-1 And RANTES In Experimental Diabetes Mellitus. Int J Nanomedicine. 2019 Nov 18;14:8961-8972. doi: 10.2147/IJN.S226790. PMID: 31819412; PMCID: PMC6873975.
- 6. Chen X-F, Yan L-J, Lecube A, Tang X. Editorial: Diabetes and Obesity Effects on Lung Function. Fronties in Endocrinology. 2020;11(462):1-2.
- 7. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annu Rev Immunol. 2009;27:519-550.
- 8. Dogan Y, Akarsu S, Ustundag B, Yilmaz E, Gurgoze MK. Serum ILlbeta, IL-2, and IL-6 in insulin-dependent diabetic children. Mediators Inflamm. (2006) 2006:59206. doi: 10.1155/MI/2006/59206

- 9. Feng S, Yu H, Yu Y, et al. Levels of Inflammatory Cytokines IL-1β, IL-6, IL-8, IL-17A, and TNF-α in Aqueous Humour of Patients with Diabetic Retinopathy. J Diabetes Res 2018; 2018: 8546423. doi:10.1155/2018/8546423
- 10. Forgiarini LA Jr, Kretzmann NA, Porawski M, Dias AS, Marroni NA. Experimental diabetes mellitus: oxidative stress and changes in lung structure. J Bras Pneumol. 2009:35(8):788-791
- 11. Gallagher KA, Liu ZJ, Xiao M. Diabetic impairments in NO-mediated endothelial progenitor cell mobilization and homing are reversed by hyperoxia and SDF-1 alpha. J. Clin. Invest. 2007; 117 (5):1249-1259.
- 12. Gandhi NA, Bennett BL, Graham NM, Pirozzi G, Stahl N, Yancopoulos GD. Targeting key proximal drivers of type 2 inflammation in disease. Nat Rev Drug Discov. 2016;15: 35-50.
- 13. Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. Immunity. 2013;39:1003-1018. https://doi.org/10.1016/j.immuni.2013.11.010
- 14. Gonzalez LL, Garrie K, Turner MD. Type 2 diabetes-An autoinflammatory disease driven by metabolic stress. Biochim Biophys Acta Mol Basis of Dis. 2018; 1864(11):3805-3823. doi:10.1016/j.bbadis.2018.08.034
- 15. Holman N, Young B, Gadsby R. Current prevalence of Type 1 and Type 2 diabetes in adults and children in the UK. Diabetic Med. 2015;32:1119-20. doi:10.1111/dme.12791
- 16. Hu JF, Zhang GJ, Wang L, Kang PF, Li J, Wang HJ, et al. Ethanol at low concentration attenuates diabetes induced lung injury in rats model. J Diabetes Res, 2014: 107152.
- 17. Jaacks LM, Siegel KR, Gujral UP, Narayan KM. Type 2 diabetes: a 21st century epidemic. Best Pract Res Clin Endocrinol Metab. 2016; 30: 331-343.
- 18. Krynytska IYa. The features of bronchoalveolar lavage cytokine profile in rats with modeled hepatopulmonary syndrome [in Ukrainian].

Tuberculosis, lung disease, HIV infections. 2013. 1(12):45-50.

- 19. Kuziemski K, Slominski W, Jassem E. Impact of diabetes mellitus on functional exercise capacity and pulmonary functions in patients with diabetes and healthy persons. BMC Endocr Disord. 2019,19:2.
- 20. Kyiak Y, Fartushok N, Onyschuk Y, Fedevych Y, Bashta G. Profile of proinflammatory cytokines in type 1 diabetes mellitus [in Ukrainian]. Physiol. journ. 2012; 58(5):65-69.
- 21. Lombardi A, Tsomos E, Hammerstad SS, Tomer Y. Interferon alpha: The key trigger of type 1 diabetes. J Autoimmun. 2018 Nov;94:7-15. doi: 10.1016/j.jaut.2018.08.003. Epub 2018 Aug 14. PMID: 30115527; PMCID: PMC6235162
- 22. Mandrup-Poulsen T, Pickersgili L, Donath MY. Blockade of interleukin 1 in type 1 diabetes mellitus. Nat Rev Endocrinol. 2010;6:158-66.
- 23. Meretskiy VM, Corda MM. Peculiarities of cytokine status at experimental craniocerebral injury on the background of diabetes mellitus [in Ukrainian]. Bulletin of Scientific Research. 2013;1:96-98.
- 24. Morey M, O'Gaora P, Pandit A, Helary C. Hyperglycemia acts in synergy with hypoxia to maintain the proinflammatory phenotype of macrophages. PLoS One, vol. 14, no. 8, article e0220577, 2019.
- 25. Ogurtsova K, da Rocha Fernandes JD, Huang Y, Linnenkamp U, Guariguata L, Cho NH, Cavan D, Shaw JE, Makaroff LE. IDF diabetes atlas: global estimates for the prevalence of diabetes for 2015 and 2040. Diabetes Res Clin Pract. 2017;128:40-50.
- 26. Pang H, Luo S, Huang G, Xia Y, Xie Z, Zhou Z. Advances in knowledge of candidate genes acting at the beta-cell level in the pathogenesis of T1DM. Front Endocrinol. (2020) 11:119. doi: 10.3389/fendo.2020.00119
- 27. Papinska AM, Soto M, Meeks CJ, Rodgers KE. Long-term administration of angiotensin (1-7) prevents heart and lung dysfunction in a mouse model of type 2 diabetes (db/db) by reducing oxidative stress,

- inflammation and pathological remodeling. Pharmacol Res. 2016; 107: 372-380. doi:10.1016/j.phrs.2016.02.026.
- 28. Pitocco D, Fuso L, Conte EG, Zaccardi F, Condoluci C, Scavone G, et al. The Diabetic Lung A New Target Organ? Rev Diabet Stud. 2012. 9:23-35.
- 29. Rajasurya V, Gunasekaran K, Surani S. Interstitial lung disease and diabetes. World Journal of Diabetes. 2020; 11(8):351-357.
- 30. Rani RE, Ebenezer BSI, Venkateswarlu M. A study on pulmonary function parameters in type 2 diabetes mellitus. National Journal of Physiology, Pharmacy and Pharmacology. 2019; 9(1):53-57.
- 31. Stepan NA, Denysenko OI. Cytokine profile of the peripheral blood in patients suffering from eczema in the acute stage [in Ukrainian]. Clinical & experimental pathology. 2014; 3(49):176-179.
- 32. Sun X, Pang H, Li J, Luo S, Huang G, Li X, Xie Z, Zhou Z. The NLRP3 Inflammasome and Its Role in T1DM. Front Immunol. 2020 Aug 27;11:1595. doi: 10.3389/fimmu.2020.01595. PMID: 32973739; PMCID: PMC7481449.
- 33. Vives-Pi M, Rodriguez-Fernandez S, Pujol-Autonell I. How apoptotic betacells direct immune response to tolerance or to autoimmune diabetes: a review. Apoptosis. (2015) 20:263-72. doi: 10.1007/s10495-015-1090-8
- 34. Wegeberg AL, Okdahl T, Floyel T, Brock C, Ejskjaer N, Riahi S, et al. Circulating Inflammatory Markers Are Inversely Associated with Heart Rate Variability Measures in Type 1 Diabetes. Mediators Inflamm. 2020 Aug 18,2020:3590389. doi: 10.1155/2020/3590389. PMID: 32908447; PMCID: PMC7450314