

Semianiv I.O., Sukholytkyi Yu.R. Analysis of deletion polymorphism of xenobiotics detoxication system genes in patients with tuberculosis and diabetes mellitus. *Journal of Education, Health and Sport*. 2022;12(7):24-29. eISSN 2391-8306. DOI <http://dx.doi.org/10.12775/JEHS.2022.12.07.003> <https://apcz.umk.pl/JEHS/article/view/JEHS.2022.12.07.003> <https://zenodo.org/record/6537402>

The journal has had 40 points in Ministry of Education and Science of Poland parametric evaluation. Annex to the announcement of the Minister of Education and Science of December 21, 2021. No. 32343. Has a Journal's Unique Identifier: 201159. Scientific disciplines assigned: Physical Culture Sciences (Field of Medical sciences and health sciences); Health Sciences (Field of Medical Sciences and Health Sciences).

Punkty Ministerialne z 2019 - aktualny rok 40 punktów. Załącznik do komunikatu Ministra Edukacji i Nauki z dnia 21 grudnia 2021 r. Lp. 32343. Posiada Unikatowy Identyfikator Czasopisma: 201159. Przypisane dyscypliny naukowe: Nauki o kulturze fizycznej (Dziedzina nauk medycznych i nauk o zdrowiu); Nauki o zdrowiu (Dziedzina nauk medycznych i nauk o zdrowiu).

© The Authors 2022.

This article is published with open access at Licensee Open Journal Systems of Nicolaus Copernicus University in Torun, Poland
Open Access. This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author (s) and source are credited. This is an open access article licensed under the terms of the Creative Commons Attribution Non commercial license Share alike. (<http://creativecommons.org/licenses/by-nc-sa/4.0/>) which permits unrestricted, non commercial use, distribution and reproduction in any medium, provided the work is properly cited.

The authors declare that there is no conflict of interests regarding the publication of this paper.

Received: 25.04.2022. Revised: 09.05.2022. Accepted: 10.05.2022.

Analysis of deletion polymorphism of xenobiotics detoxication system genes in patients with tuberculosis and diabetes mellitus

Semianiv I.O.¹, Sukholytkyi Yu.R.¹,

1. Phthisiology and Pulmonology Department, Bukovinian State Medical University, Chernivtsi, Ukraine, 58002

Semianiv I.O., assistant of the Phthisiology and Pulmonology Department of Bukovinian State Medical University; ORCID ID: [0000-0003-0340-0766](https://orcid.org/0000-0003-0340-0766); E-mail: igor_semianiv@bsmu.edu.ua
Sukholytkyi Yu.R., student, Phthisiology and Pulmonology Department of Bukovinian State Medical University; ORCID ID: [0000-0003-2027-1951](https://orcid.org/0000-0003-2027-1951); E-mail: suholitkiy.yuriy@gmail.com

Abstract. An analysis of the occurrence of alleles and genotypes of GSTM1 gene in patients with pulmonary tuberculosis and diabetes mellitus regarding the MBT resistance version allowed to establish that under the conditions of pulmonary tuberculosis infection GSTM1 gene deletion mutation can be found in one out of five (21.87% of cases), and the occurrence due to the MBT resistance version is: with NDTB - 17.39%, with MDR-TB 35.0% and - PRTB 20.0% respectively. According to the nature of the distribution of allelic gene GSTM1 a favorable functional 1 allele prevails (73.29%) in the normal inbreeding among patients and deficiency of heterozygosity among healthy people, which generally forms a normal population distribution for the European race.

Objective. To identify GSTM1 gene polymorphism in patients with tuberculosis and diabetes mellitus regarding the MBT resistance version.

Material and methods. The study involved 100 patients with newly diagnosed pulmonary TB and diabetes mellitus who had been hospitalized in Chernivtsi Regional TB Dispensary. The control group consisted of 50 healthy individuals. Genomic DNA was isolated from the whole venous blood. GSTM1 polymorphic areas were isolated by means of multicomplex polymerase chain reaction, according to the protocol for instantaneous analysis of polymorphism by M. Arana et al (1996). Deletion of gene corresponds to the lack of appropriate strips in the electropherogram.

Results and discussion. Despite the fact that the activity of the enzyme glutathione-S-transferase of class M is encoded by five GST genes of class M (M1-M5), the dominant cause of genetically caused dysregulation of antioxidant activity is deletion (null) polymorphism of the gene GSTM1.

Conclusion. Among the patients with pulmonary tuberculosis and diabetes mellitus one out of five persons (21,87 % of cases) was diagnosed with deletion mutation of GSTM1 gene; and the occurrence due to MBT resistance variation is: in NDTB-17,39 %, in MDR-TB - 35,0 % and in PRTB- 20,0 % respectively.

Key words: tuberculosis, diabetes mellitus, deletion polymorphism, resistance, MBT.

Introduction. The importance of TB and DM's problem is due on the one hand to the growing number of patients with tuberculosis with multiple drug resistance to the pathogen, and on the other – to a steady increase in the number of people with various forms of carbohydrate metabolism [1,3,9]. Thus, modern objective reality increases the urgency of the problem of this combined pathology, as well as necessitates the study and proper understanding of the mechanisms of tuberculosis infection in this category of patients [2,4,13].

GST are enzymes of the second phase of detoxification systems which protect the body against endogenous oxidative stress and exogenous toxins, catalyzing conjugation of sulfhydryl groups of reduced glutathione and rendering harmless various electrophilic compounds, including products of lipid and DNA oxidation [5,7].

Objective. To identify GSTM1 gene polymorphism in patients with tuberculosis and diabetes mellitus regarding the MBT resistance version.

Material and methods. The study involved 100 patients with newly diagnosed pulmonary TB and diabetes mellitus who had been hospitalized in Chernivtsi Regional TB Dispensary. The control group consisted of 50 healthy individuals. Genomic DNA was isolated from the whole venous blood. GSTM1 polymorphic areas were isolated by means of multicomplex polymerase chain reaction, according to the protocol for instantaneous analysis of polymorphism by M. Arana et al (1996). Deletion of gene corresponds to the lack of appropriate strips in the electropherogram. We used the program STATISTICA, version 10.0.228.8 (StatSoft, Inc.) for statistical analysis of the findings. The difference in the distribution of occurrence of genotypes and their combinations between groups were calculated using χ^2 criteria . Differences were regarded as significant at significance level $p < 0.05$. The association of genotypes with susceptibility to tuberculosis was judged by the size of the odds ratio (odds ratio, OR) [18].

Results and discussion. Despite the fact that the activity of the enzyme glutathione-S-transferase of class M is encoded by five GST genes of class M (M1-M5), the dominant cause of genetically caused dysregulation of antioxidant activity is deletion (null) polymorphism of the gene GSTM1 [12, 15]. Due to the above, we have analyzed the occurrence of alleles and genotypes of GSTM1 gene in patients with pulmonary tuberculosis and diabetes mellitus due to MBT resistance version [16].

Results and discussion. Lack of 0-genotype was found in 214 (73.29%) cases out of 292 isolated alleles ($n = 107$), while the "mutant" deletion (0-allele) was observed by 2.74 times less frequently - in 78 (26.71%) cases ($n = 39$) ($\chi^2 = 63,34$, $p < 0.001$) (Table. 1).

Table 1 – Distribution of deletion polymorphism of the gene glutathione-S-transferase of class M1 (GSTM1)

| Study groups | Experimental group, n=96 | Control group, n=50 | χ^2 p | Total, n=146 (%) |
|----------------------|-----------------------------|----------------------------|----------------------------|-----------------------------|
| No 0-genotype, n (%) | 75 (78,13) | 32 (64,0) | $\chi^2=3,35$ $p=0,067$ | 107 (73,29) |
| 0-genotype, n (%) | 21 (21,87) | 18 (36,0) | $\chi^2=3,67$ $p=0,052$ | 39 (26,71) |
| χ^2 p | $\chi^2=60,75$ $p<0,001$ | $\chi^2=7,84$ $p=0,005$ | - | $\chi^2=63,34$ $p<0,001$ |

The relative occurrence of 0-genotype and its absence among TB patients and healthy individuals did not differ significantly ($p > 0.05$). Thus, in both groups the functional allele of gene GSTM1 was found much more frequently: by 3.57 times in the experimental group ($\chi^2 = 60,75$ $p < 0,001$) and by 1.78 times in the control group ($\chi^2 = 7,84$ $p = 0.005$) (Table 3.1). The resulting distribution in observation groups reflected the general one in the surveyed population , which was also dominated by those with wild 1 allele by 2.74 times over those with non-functional 0-genotype ($p < 0,001$).

Race and population analysis of gene GSTM1 null polymorphism showed that the frequency of homozygous null genotype gene appointed above among the examined tuberculosis patients was lower than in European population (PD = 0,42-0,60 vs PD = 0.22, $p < 0.05$) and Asian races (PD = 0,42-0,54, $p < 0.05$), it did not differ significantly from the corresponding figure of the equatorial race (PD = 0,16-0,36, $p < 0.05$). Occurrence of null genotype in the control group of the examined patients did not differ significantly from the rate for Caucasians ($p > 0.05$). In addition, the occurrence of GSTM1 0/0-genotype in our experimental (PD = 0.22) and control groups (PD = 0.36) corresponded to averages in Ukrainian (south-eastern and central Ukraine) and some Eastern European populations (PD = 0,15-30).

Allelic distribution according to the polymorphic variant of gene GSTM1 among TB patients and healthy individuals in general corresponds to the expected population equilibrium Hardy-Weinberg (Table 2). In quantitative terms, an allele without genotype-0 is dominant ($P_1 = 54.0\%$), while the relative occurrence of alleles did not differ significantly. We found statistically significant heterozygote deficiency in the control group ($F = 0,28$, $p = 0.033$), which does not generally cover the entire sample ($F = 0,24$, $p > 0.05$) and shows a normal population distribution.

Table 2 – Analysis of heterozygosity of null polymorphism of the gene glutathione-S-transferase of class M1 (GSTM1)

| Groups | Genotypes, alleles, n (%) | | P _D | P ₁ | H ₀ | H _E | F | χ^2 | P |
|--------------------------|---------------------------|-------------|----------------|----------------|----------------|----------------|------|----------|-------|
| | DD | 1 allele | | | | | | | |
| Experimental group, n=96 | 21 (21,87) | 75 (78,13) | 0,41 | 0,59 | 0,38 | 0,48 | 0,20 | 2,33 | >0,05 |
| Control group, n=50 | 18 (36,0) | 32 (64,0) | 0,54 | 0,46 | 0,36 | 0,50 | 0,28 | 4,56 | 0,033 |
| Total, n=146 | 39 (26,71) | 107 (73,29) | 0,46 | 0,54 | 0,38 | 0,50 | 0,24 | 3,27 | >0,05 |

Notes: 1. P₁ – relative occurrence of 1 allele; P_D – relative occurrence of deletion allele D. 2. H₀ – real heterozygosity; H_E – expected heterozygosity; F – inbreeding factor. 3. $\chi^2 p$ – criterion of correctness of “null” hypothesis between real and expected heterozygosity.

The occurrence of 00-gene GSTM1 genotype in patients with pulmonary tuberculosis depending on the type is shown in Table 3. We found significantly more frequent presence of a functional allele than its absence, in patients with newly diagnosed pulmonary tuberculosis (NDTB) by 4.75 times ($p < 0.001$) and in those with poly-resistant pulmonary tuberculosis (PRTB) by 4 times ($p < 0.001$), respectively. There was no substantial difference in frequency in patients with multi-drug resistant tuberculosis (MDR-TB) ($p = 0.056$). It should be noted that among the carriers of non-functional allele in the experimental group there were more patients with NDTB than those with MDR-TB and PRTB by 2,92 ($\chi^2 = 18,57$, $p < 0.001$) and 1,58 ($\chi^2 = 5,39$, $p = 0.02$) times. At the same time there were more patients with PRTB and without mutated GSTM1 gene than those with MDR-TB: 32.0% vs 17,33% ($\chi^2 = 4,34$, $p = 0.037$), respectively. There were no significant differences between the occurrence of certain types of pulmonary tuberculosis (NDTB, MDR-TB, PRTB) among homozygous carriers of the gene GSTM1 of the deletion genotype (Table 3).

Table 3 – Occurrence of the gene GSTM1 null genotype in patients with TB and diabetes mellitus

| Study groups | No 0-genotype, n=75 (%) | 0-genotype, n=21 (%) | LOD [95% CI] | $\chi^2 p$ |
|--|-------------------------|----------------------|-------------------|-------------------------------|
| Newly diagnosed tuberculosis, n=46 (%) | 38 (82,61) | 8 (17,39) | 22,56 [7,67-66,3] | $\chi^2=39,13$ $p < 0,001$ |
| Multidrug resistant tuberculosis, n=20 (%) | 13 (75,0) | 7 (35,0) | 3,45 [0,94-12,6] | $\chi^2=3,60$ $p=0,056$ |
| Poly-resistant tuberculosis, | 24 (80,0) | 6 (20,0) | 16,0 | $\chi^2=21,60$ |

| | | | | | |
|-------------------|-------------|---------------------------|------------------------|---------------------|-----------------------|
| n=30 (%) | | | | [4,51-56,7] | p<0,001 |
| χ^2 p | NDTB-MDR-TB | $\chi^2=18,57$ p<0,001 | $\chi^2<1,0$ p>0,05 | - | - |
| | NDTB-PRTB | $\chi^2=5,39$ p=0,02 | $\chi^2<1,0$ p>0,05 | | |
| | MDR-TB-PRTB | $\chi^2=4,34$ p=0,037 | $\chi^2<1,0$ p>0,05 | | |
| Control, n=50 (%) | | 32 (64,0) | 18 (36,0) | 3,16 [1,40-7,15] | $\chi^2=7,84$ p=0,005 |

Note. LOD – logarithm of the odds ratio score; CI – confidence interval; p – differences in probability; NDTB – newly diagnosed tuberculosis; MDR-TB – multidrug resistant tuberculosis; PRTB poly-resistant pulmonary tuberculosis.

An analysis of heterozygosity of null polymorphism of the gene GSTM1 heterozygous gene GSTM1, taking into account diagnosed MBT resistance variation (Table 4), showed normal allelic distribution, which corresponded to the scale of population equilibrium by Hardy-Weinberg ($p > 0,05$). In quantitative terms, the dominant allele in the experimental group regardless of the type of tuberculosis is functional variant 1 (75,0-82,61% vs 17,39-35,0%).

Table 4 – Analysis of heterozygosity of null polymorphism of the gene glutathione-S-transferase of class M1 (GSTM1) due to the MBT resistance variation

| Groups | Genotypes, alleles, n (%) | | P _D | P _I | H _O | H _E | F | χ^2 | P |
|------------------|---------------------------|---------------------|----------------|----------------|----------------|----------------|------|----------|-------|
| | DD | I _{allele} | | | | | | | |
| NDTB, n=46 (%) | 8 (17,39) | 38 (82,61) | 0,39 | 0,61 | 0,43 | 0,48 | 0,09 | 1,32 | >0,05 |
| MDR-TB, n=20 (%) | 7 (35,0) | 13 (75,0) | 0,53 | 0,48 | 0,35 | 0,50 | 0,30 | 2,36 | >0,05 |
| PRTB, n=30 (%) | 6 (20,0) | 24 (80,0) | 0,37 | 0,63 | 0,33 | 0,46 | 0,28 | 2,23 | >0,05 |
| Total, n=96 | 21 (21,87) | 75 (78,13) | 0,41 | 0,59 | 0,38 | 0,48 | 0,20 | 2,33 | >0,05 |

Notes: 1. – NDTB – newly diagnosed tuberculosis; MDR-TB – multidrug resistant tuberculosis; PRTB poly-resistant pulmonary tuberculosis. 2. P_I – relative occurrence of I allele; P_D – relative occurrence of deletion allele D. 3. H_O – real heterozygosity; H_E – expected heterozygosity; F – inbreeding factor. 4. χ^2 p – criterion of the correctness of null hypothesis between real and expected heterozygosity.

Conclusions. 1. Among the patients with pulmonary tuberculosis and diabetes mellitus one out of five persons (21,87 % of cases) was diagnosed with deletion mutation of GSTM1 gene; and the occurrence due to MBT resistance variation is: in NDTB-17,39 %, in MDR-TB - 35,0 % and in PRTB- 20,0 % respectively.

2. According to the nature of allele distribution of GSTM1 gene the favorable functional 1 allele prevails (73,29 %) in case of normal inbreeding in patients (F=0,20, p>0,05) and lack of heterozygosity in healthy individuals (F=0,28 p=0,033), which, in general forms a normal population distribution [OR=14,06, p=0,005].

REFERENCES

1. Al-Rifai RH, Pearson F, Critchley J, Abu-Raddad LJ. Association between diabetes mellitus and active tuberculosis: a systematic review and meta-analysis. *PLoS ONE*. 2017; 12: e0187967.

2. American Diabetes Association (2019) Standards of Medical Care in Diabetes-2019 Abridged for Primary Care Providers. *Clin Diabetes*. 2019; 37(1): 11–34.
3. Genetic polymorphism of GSTM1 and GSTP1 in lung cancer in Egypt. / M. Maggie Ramzy, M. Mohei El-Din Solliman, A. Hany Abdel-Hafiz [et al.] // Intern. J. Of Collabor. Research on Intern. Med. &Public Health. – 2018. – Vol. 3 No. 1. – P.41-51.
4. Genetic polymorphisms of NAT2, CYP2E1 and GST enzymes and the occurrence of antituberculosis drug-induced hepatitis in Brazilian TB patients. / R.L.Teixeira, R.G. Morato, P.H. Cabello [et al.] // Mem. Inst. Oswaldo. Cruz. – 2017. - Vol. 106 (6). – P. 716-724.
5. GST M1-T1 null allele frequency patterns in geographically assorted human populations: a phylogenetic approach / S.P. Kasthurinaidu, T. Ramasamy, J. Ayyavoo [et al.] // PLoS One. – 2015. – Vol. 10(4). – P. 23-27.
6. Harries AD, Lin Y, Kumar AMV, Satyanarayana S, Zachariah R, Dlodlo RA. How can integrated care and research assist in achieving the SDG targets for diabetes, tuberculosis and HIV/AIDS? *Int J Tuberc Lung Dis*. 2018; 22: 1117-1126.
7. Jain KK, Thakuria R, Lokesh S. Prevalence of pulmonary diabetes mellitus in tuberculosis patients attending tertiary care institute. *International Medical Journal*. 2015; 2(4):245-8.
8. Koesoemadinata RC, Kranzer K, Livia R, et al. Computer-assisted chest radiography reading for tuberculosis screening in people living with diabetes mellitus. *Int J Tuberc Lung Dis*. 2018; 22: 1088-1094.
9. Li C, Long J, Hu X, Zhou Y. GSTM1 and GSTT1 genetic polymorphisms and risk of anti-tuberculosis drug-induced hepatotoxicity: an updated meta-analysis. *European journal of clinical microbiology & infectious diseases: official publication of the European Society of Clinical Microbiology*. 2013;32(7):859–68. Epub 2013/02/05. 10.1007/s10096-013-1831-y .
10. Liu F, Jiao AX, Wu XR, Zhao W, Yin QQ, Qi H, et al. Impact of glutathione S-transferase M1 and T1 on anti-tuberculosis drug-induced hepatotoxicity in Chinese pediatric patients. *PloS one*. 2014;9(12):e115410 Epub 2014/12/20. 10.1371/journal.pone.0115410 ; PubMed Central PMCID: PMC4272297.
11. Liu Q, Li W, Xue M, et al. Diabetes mellitus and the risk of multidrug resistant tuberculosis: a meta-analysis. *Scientific Reports*. 2017; 7: 1090.
12. *Molecular epidemiology* / [V. Zaporozhan, YI Bazhora, V. Krysyun et al.]. - Odessa: ONMU, 2019. - 356 p.
13. Ruslami R, Aarnoutse RE, Alisjahbana B, van der Ven AJ, Van Crevel R. Implications of the global increase of diabetes for tuberculosis control and patient care. *Trop Med Int Health*. 2010;12(11):1289-99.
14. Semianiv I, Todoriko L, Ieremenchuk I. Prevention of adverse reactions due to pharmacotherapy in MRTB considering polymorphism of glutathione-S-transferase M1 and T1 genes. *Europen Respiratory Journal*. 2017;49: 60.
15. Syal K, Srinivasan A, Banerjee D. VDR, RXR, coronin-1 and interferon γ levels in PBMCs of type-2 diabetes patients: Molecular link between diabetes and tuberculosis. *Ind J Clin Biochem*. 2015;30(3):323-8.
16. Todoriko LD. Alel'nyy stan heniv biotransformatsiyi ksenobiotyktiv hlutation-S-transferazy klasiv T1 (GSTT1) ta M1 (GSTM1) u khvorykh na tuberkul'oz lehen' [Alele of xenobiotics metabolism genes of glutathione-S-transferase classes T1(GSTT1) and M1 (GSTM1) in patients with pulmonary tuberculosis]. *Tuberkul'oz, lehenevi khvoroby, VIL-infektsiya [Tuberculosis, Lung Diseases, HIV-Infection]*. 2016; 25(2):73-8. – http://tubvil.vitapol.com.ua/svizhij_nomer.php?nid=25
17. Todoriko LD. Immunopathogenesis of drug-resistant tuberculosis present position. *Tuberculosis Lung disease HIV-infection*. 2017;3(30):92-8.

18. WHO consolidated guidelines on tuberculosis Module 2: screening - systematic screening for tuberculosis disease, 2021.